

Clinical significance of matrix metalloproteinases activity in acute myocardial infarction

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ABSTRACT. Matrix metalloproteinases (MMP) degrade myocardial fibrillar collagen in acute myocardial infarction (MI) patients. Their activity is tightly controlled in normal myocardium by a family of closely related tissue inhibitors known as TIMP. An imbalance in their activity might contribute to post-MI remodeling. Plasma levels of MMP-1, TIMP-1 and MMP-1/TIMP-1 complex were measured, using relevant ELISA kits, in 24 (22 males-2 females), acute MI patients with a mean age 59 ± 14 years. Blood samples were taken on admission (0h), and 3h, 6h, 9h, 18h, 24h, 36h, 48h, 3rd, 4th, 5th, 7th, 15th, 30th days after MI. All patients underwent coronary arteriography with ventriculography for estimation of left ventricular ejection fraction (LVEF) and extent of coronary artery diseases, and echocardiographic study for measuring end-diastolic diameter (EDD). Ten patients with an LVEF < 45%, an EDD > 47.5mm, and heart failure symptoms were included in group A and compared against 12 patients with an LVEF > 45% an EDD < 47.5mm in group B. Mean plasma concentrations of MMP-1 were higher by 21% in group A (1.3 ± 0.2 ng/mL) compared to group B (1 ± 0.1 ng/mL) over the total study period. TIMP-1 plasma concentrations showed very little difference between the 2 groups, (704 ± 213 ng/mL versus 691 ± 165 ng/mL, (6%)) Finally, plasma concentrations of MMP-1/TIMP-1 complex were lower by -36% in group A with a mean value of 2.7 ± 0.6 ng/mL versus 3.7 ± 0.5 ng/mL in group B. Mean values for the differences were significant at time points 0, 6, 18, 24 and 48 hours for MMP-1 ($p < 0.036$), and on 48h and the 4th day for MMP-1/TIMP-1 complex ($p < 0.031$). Moreover, a good correlation was found between plasma concentrations of creatine kinase (CK) and MMP-1 at 18h ($r = 0.422$, $p = 0.041$) and on the 4th day ($r = 0.67$, $p = 0.046$), and TIMP-1 on the 4th day ($r = 0.67$, $p = 0.047$). Additionally, mean values for LVEF were $35.8 \pm 8.8\%$ in group A versus $51.2 \pm 1.8\%$ ($p = 0.00014$) in group B. Also, the EDD in-group A was 52.1 ± 6.9 mm versus 42.9 ± 3.2 mm in group B ($p = 0.00013$). In acute MI patients, increased MMP-1, with no change in TIMP-1, is associated with left ventricular dysfunction and dilatation, suggesting that increased collagenolytic activity contributes to loss of LV function.

Keywords: matrix metalloproteinases, acute myocardial infarction, extracellular matrix

Left ventricular dysfunction and remodeling after myocardial infarction (MI) are thought to be among the first steps to progression to heart failure, the degree of remodeling predicting morbidity and mortality [1-6].

The combination of cellular and extracellular events that occur during the post-MI period results in changes in left ventricular geometry, and has been called infarct expansion [3-6]. Previous studies have demonstrated that a structural determinant of infarct expansion is the extracellular collagen matrix (ECCM) remodeling [1-6]. The matrix metalloproteinases are a family of proteolytic enzymes, that contribute to the left ventricular remodeling process [7, 8]. Endogenous MMP activity is controlled by tissue inhibitors of MMP (TIMP) [7, 8]. Another role of the TIMP that is independent of modulating MMP actional

states, is through their effects on cell growth. TIMP-1 and TIMP-2 have been shown to stimulate a growth response in fibroblast cell cultures in a concentration-dependent manner [9]. Past clinical studies have demonstrated alterations in MMP and TIMP after MI [10, 11]. Recent experimental data have provided evidence for a cause-and-effect relationship between MMP-induced- and the post-MI myocardial-remodeling process [7, 12-14].

The purpose of this study was to examine whether a differential expression of plasma concentration occurs between matrix metalloproteinase-1 (MMP-1) and its natural tissue inhibitor (TIMP-1) in patients with acute MI. This unequal expression of plasma concentrations affects left ventricular chamber dimensions and global contractile performance.

MATERIALS AND METHODS

Study population

The study population consisted of 24 patients (22 males, 2 females, mean age 59 ± 14 years) who had been admitted to our department of Cardiology in Laiko Hospital, with a first attack of acute MI, and 12 healthy subjects as controls (8 males, 4 females, mean age 43 ± 14), who were studied in order to estimate normal levels of collagenolytic enzymes. The control subjects had no past history or any evidence of cardiovascular disease, hypertension or diabetes mellitus. The present study did not include any patients or control subjects with a history of neoplastic, hepatic, or infectious disease, severe renal failure, serum creatinine > 2.5 mg/dL or any surgical procedure in the preceding 6 months. The local scientific ethics committee approved the study and each participant gave their informed written consent.

The diagnosis of acute MI was confirmed by typical, prolonged chest pain, accompanied by serial changes on the standard, 12-lead electrocardiogram (ECG), or significant (two-fold greater than the normal range) increases in markers of myocardial damage creatine kinase (CK) or its isoenzyme MB (CK-MB).

All patients were admitted less than 6h from the onset of chest pain (mean 4.2 ± 0.6 h). Depending upon the indications, patients were thrombolized using alteplase, and were treated with angiotensin-converting enzyme inhibitor (ACEI) (table 1). All patients received a combination of standard medication including nitrates, beta-adrenergic blockers, calcium antagonists, and low molecular weight heparin and antiplatelet drugs such as aspirin or clopidogrel.

Table 1
Baseline characteristics

| | Group A | Group B | Control group |
|--|-------------------------------|-------------------------------|-------------------------------|
| Patient number | 10 | 12 | 12 |
| Age (years): | | | |
| Mean \pm SD | 55 ± 13 | 57 ± 11 | 43 ± 14 |
| Range | 37-70 | 41-77 | 31-77 |
| Sex: | | | |
| Male/Female | 10/0 | 10/2 | 8/4 |
| Infarct location: | | | |
| Anterior/Inferior | 9/1 | 6/6 | |
| Extent of coronary artery disease: | | | |
| 0 vessel disease | 0 | 0 | |
| 1 vessel disease | 1 pt | 7 pts | |
| 2 vessel disease | 5 pts | 4 pts | |
| 3 vessel disease | 3 pts | - | |
| No catheterization | 1 pt | 1pt | |
| Total Cholesterol: | | | |
| > 200 mg% | 6 pts | 10 pts | |
| < 200 mg% | 4 pts | 2 pts | |
| Thrombolysis: | | | |
| Yes | 9 pts | 5 pts | |
| No | 1 pt | 7 pts | |
| Angiotensin converting enzyme inhibitor | | | |
| Yes | 8 pts | 5 pts | |
| No | 2 pts | 7 pts | |

Two out of the 24 patients died on the 4th post-infarction day, one from acute pulmonary edema due to rupture of the chordae tendinae followed by acute mitral regurgitation and death, and the other as a result of extension of the acute MI from the inferior towards the lateral and anterior wall, followed by cardiogenic shock and ventricular fibrillation. These two patients are not included in the study. In sixteen patients, the infarct location was the anterior wall, and in eight, the inferior wall.

Patients were assigned to two groups according to the normal lower limits of LVEF = 45%, and the mean value (47.5mm) of EDD measured at papillary muscle level by two-dimensional echocardiography. Ten patients with an ejection fraction of $< 45\%$, heart failure symptoms (third heart sound (S_3) and basal rales on auscultation, mild congestion on chest X-ray) during the hospitalization in the coronary care unit, and left ventricular end-diastolic diameter (LVEDD) > 47.5 mm were included in group A. This group was compared to the remaining 12 patients with an LVEF $> 45\%$, without heart failure symptoms, and LVEDD < 47.5 mm (group B).

Hemodynamic and echocardiographic evaluation

On 6th post-MI day, twenty out of the 22 patients (two patients refused) underwent coronary arteriography to estimate the extent of coronary artery disease and single-plane ventriculography in the right anterior oblique (RAO) view in order to estimate ejection fraction [15] and regional wall movement abnormalities in the anteriorbasal, and posteriorbasal segments [16]. We evaluated LVEDD, on the 6th and 30th days [17, 18], using a Hewlett Packard Sonos 1000 ultrasound system with a 2.5 MHz transducer.

Serum plasma concentrations of MMP-1, TIMP-1 and MMP-1/TIMP-1 complex

Blood samples were collected on admission (0 hours), at 3h, 6h, 9h, 12h, 18h, 24h, 36h and 48 hours and on the 3th, 4th, 5th, 7th, 15th and 30th day after the acute MI. They were centrifuged for 10 minutes at 3000 rpm and stored at -20° to 30°C until the assay (within 6 months).

Plasma concentrations of MMP-1, TIMP-1, and MMP-1/TIMP-1 complex antigen levels were measured using the relevant ELISA kits provided by Amersham International plc, Buckingham, UK. The matrix metalloproteinase-1 human ELISA system code 2610 [19] measured total human MMP-1, both free and that complexed with TIMP-1. The lower limit of detection was 0.5 ng/mL and the coefficients of variation of the intra-assay/ inter-assay were 6.7% and 12.4% respectively. The immuno-assay system for TIMP-1, code 2611 [20], recognize total human TIMP-1, free TIMP-1 and that complexed with MMP-1. The lower detection limit and the intra-assay/inter-assay variation coefficients were 45 ng/mL, 10.5% and 13.7% respectively. Also, the immuno-assay systems for MMP-1/TIMP-1 complex, code 2612 [21], recognize activated MMP-1 that had subsequently been complexed with the specific inhibitor TIMP-1. It did not recognize free, active MMP-1, TIMP-1, or Pro MMP-1 or MMP-1/TIMP-1 complex from other species. The lower detection limit was 1.5 ng/mL, and the intra-assay/inter-assay variation coefficients were 8.2% and 12.7% respectively. The immuno-

assay is based on a two-side ELISA sandwich format. Concentrations in ng/mL were determined by interpolation from a standard curve for each collagenolytic enzyme. Blood samples for assay of CK and its isoenzyme MB were obtained at the same time points, and measured using Medicon, Olympus and Bayer ELISA Kits respectively.

Statistical analysis

We employed analysis of variance (ANOVA repeated measurements) to assess time-dependent alterations in serum MMP-1, TIMP-1, and MMP-1/TIMP-1 complex concentrations. If the analysis indicated a statistically significant change, between subjects and within subjects, then the difference between the mean values was compared by paired and unpaired Student's t-test and the Mann-Whitney test for the same period of time. Blood values are given as mean \pm SD. $p < 0.05$ was considered statistically significant.

RESULTS

The clinical and demographic characteristics of both groups of patients are presented in *table 1*. There were no significant differences as regards age and sex. However, clear baseline differences existed in blood plasma cholesterol levels, extent of coronary artery disease, site and size of infarct, number of patients having been thrombolysed ($n = 14$) and treated with ACEI ($n = 13$).

Time course of MMP-1, TIMP-1, MMP-1/TIMP-1 complex plasma concentration changes

MMP-1

Figure 1 shows the time-dependent changes in plasma concentrations of MMP-1 in both groups of acute MI patients. Group A expressed higher levels of MMP-1 plasma concentrations, average percentage increase (21%)

for the total study period, while the mean \pm SD value was 1.3 ± 0.2 ng/mL in group A versus 1 ± 0.1 ng/mL in group B. Furthermore, at certain time points (6h, 18h, 24h, 48h), the mean values of the differences between the groups, were statistically significant ($p < 0.006$).

TIMP-1

Figure 2 outlines the time-dependent changes in TIMP-1 plasma concentrations. Note that both curves for TIMP-1 plasma concentrations are characterized by fluctuations of mean values throughout the study period, while the mean average plasma concentrations were 704 ± 213 ng/mL in group A versus 691 ± 165 ng/mL in group B, and the percentage difference between them was 6%.

MMP-1/TIMP-1 complex

Figure 3 demonstrates the time-dependent changes in MMP-1/TIMP-1 complex plasma concentrations for each time point in both groups of acute MI patients. Note that the MMP-1/TIMP-1 complex, reflecting activated MMP-1 and expressed as TIMP-1 component, shows a lower average expression in group A compared to group B (-38%) over the total study period, while the mean \pm SD values were 2.7 ± 0.6 ng/mL versus 3.7 ± 0.5 ng/mL in both groups, respectively. Furthermore, at 48h and on the 4th day, the mean values for the differences between groups were statistically significant, with $p < 0.031$ and $p < 0.0052$ respectively.

Healthy subjects

Mean \pm SD plasma values for these subjects were, MMP-1 1.2 ± 0.2 ng/mL, TIMP-1 222 ± 69 ng/mL and MMP-1/TIMP-1 complex 1.3 ± 0.3 ng/mL. Statistical analysis between the mean values for MMP-1, TIMP-1, MMP-1/TIMP-1 complex, and the corresponding mean values for normal healthy subjects showed no statistical significance for MMP-1, while TIMP-1 and MMP-1/TIMP-1

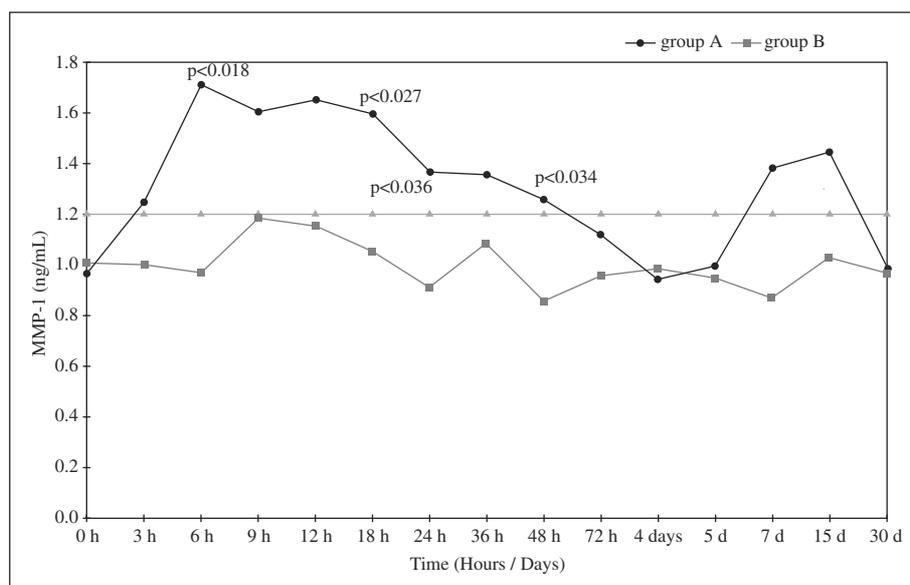


Figure 1

Mean changes in MMP-1 as a function of time in acute MI patients allocated to group A, compared to group B. Note the significant increase in MMP-1 activity in the first 48 hours after the MI. The straight line represents mean value of MMP-1 in healthy subjects.

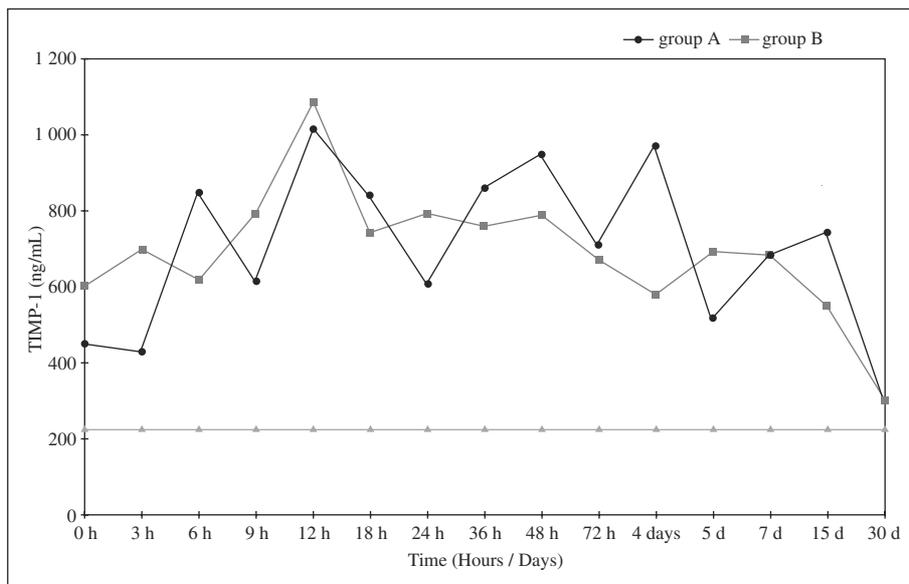


Figure 2

Mean changes in TIMP-1 as a function of time in acute MI patients allocated to group A, compared to group B. The straight line represents mean values of TIMP-1 in healthy subjects.

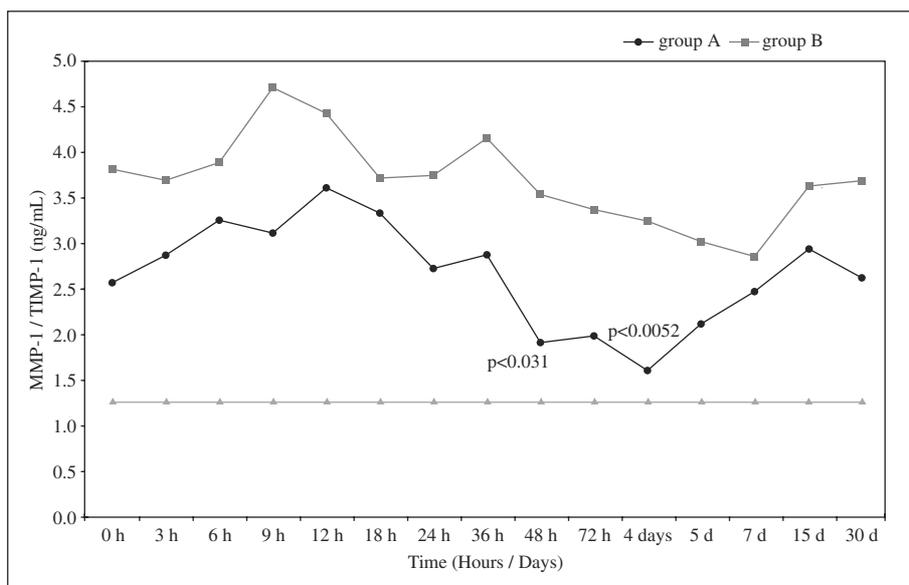


Figure 3

Mean changes in MMP-1/TIMP-1 complex as a function of time in acute MI patients allocated to group A, compared to group B. Note the lower mean values for MMP-1/ TIMP-1 complex throughout the study period, and the significant changes on 48h and the 4th day ($p < 0.031$ and $p < 0.052$). The straight line represents the mean values of MMP-1/ TIMP-1 complex of healthy subjects.

were statistically significant with $p < 0.048$ and $p < 0.049$ respectively at most time points.

Left ventricular ejection fraction

Characteristics are the mean values for ejection fraction (figure 4) between the two groups of acute MI patients. Group A had an ejection fraction of $35.8 \pm 8.8\%$, whereas group B had an ejection fraction of $51.2 \pm 1.8\%$. This -43% percentage reduction in the ejection fraction in group A compared to group B was statistically significant ($p < 0.00014$).

Left ventricular dimensions

Echocardiographic study on the 6th day, showed a mean value for LVEDD of 52.2 ± 6.9 mm in group A, whereas

group B patients demonstrated a mean value for LVEDD of 42.9 ± 3.2 mm (figure 5). This 17% difference between the two groups was statistically significant ($p < 0.001319$). Moreover, echocardiographic study on the 30th day, showed a value for LVEDD of 53.2 ± 7.4 mm for group A versus 43.3 ± 2.4 mm for group B. This 18% difference was also statistically significant ($p < 0.0009$).

Creatine kinase

Additionally, figure 6 demonstrates the mean values for the changes in CK (IU/mL), as a function of time, in both groups of acute MI patients. Note the significance of changes between time points 6h to the 5th day ($p < 0.024$). Also, a good correlation was found between plasma con-

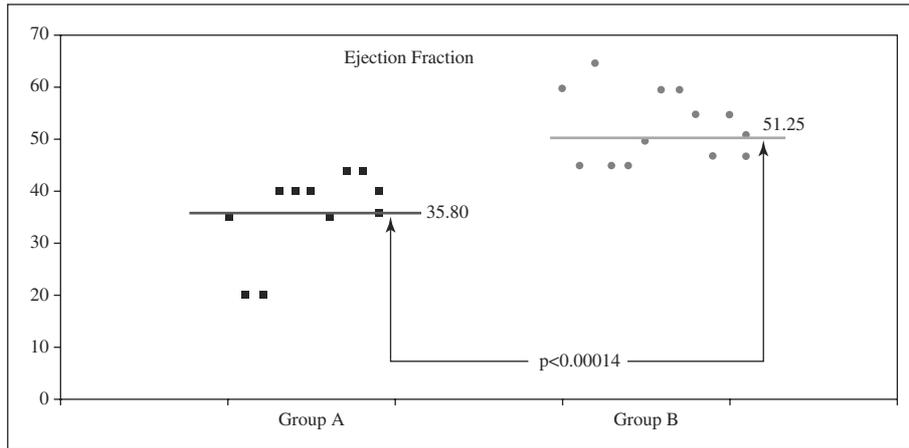


Figure 4

Mean value for ejection fraction between group A and group B. Note the characteristic (43%) statistically significant ($p < 0.00014$) reduction of ejection fraction.

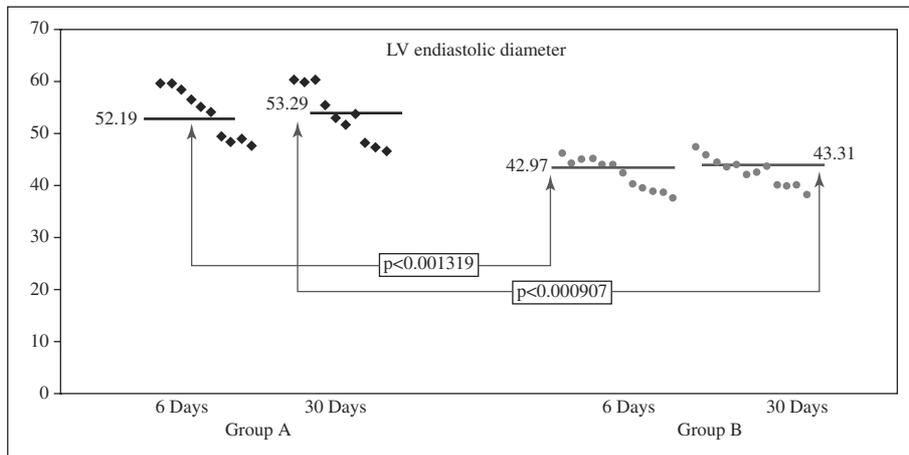


Figure 5

Mean value for left ventricular end diastolic diameter measured at the 6th and at 30th day, by echocardiographic study, in group A and group B. Note the left ventricular chamber dimensions, which were unchanged in the two echocardiographic measurements for both groups.

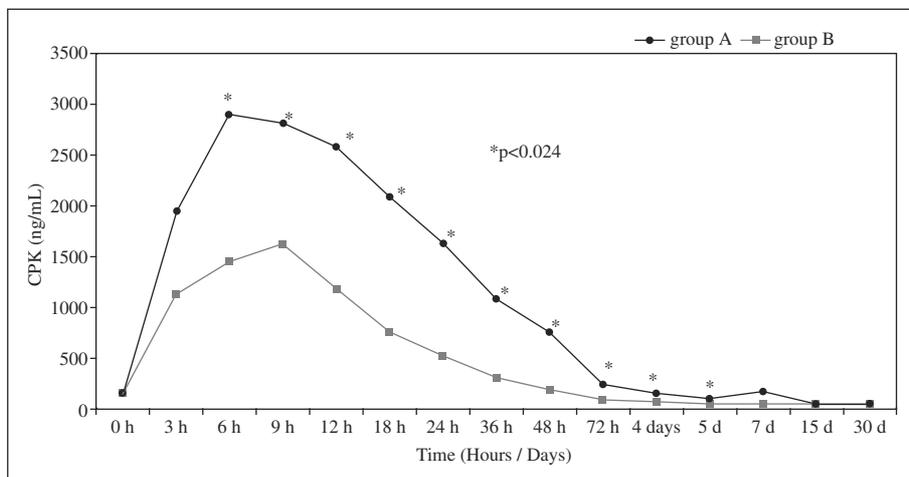


Figure 6

Mean changes in creatine kinase IU/mL as a function of time in acute MI patients allocated to group A, and group B. Note that the significance ($0.0027 < p < 0.034$) changes from 6h to 5 days.

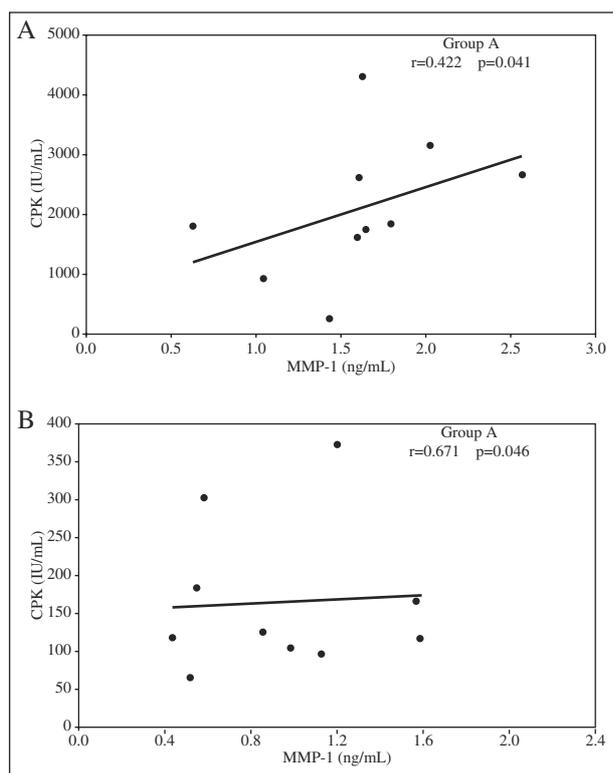


Figure 7

Graphs showing a positive correlation between plasma concentrations of MMP-1 and CK at 18h ($r = 0.422$, $p = 0.041$) (A), and on the 4th day ($r = 0.67$ $p < 0.046$) (B), in acute MI patients.

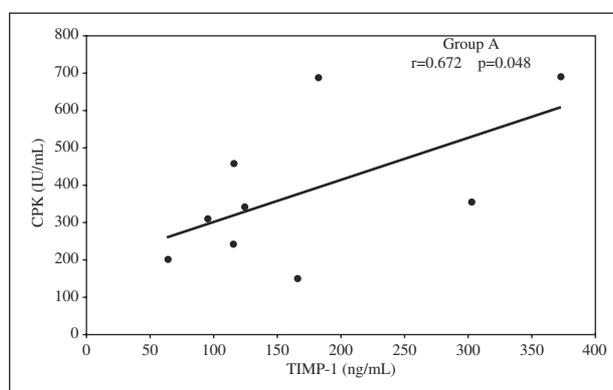


Figure 8

Correlation between plasma concentrations of TIMP-1 and CK on the 4th day in acute MI patients ($r = 0.67$ $p < 0.047$).

centrations of CK and MMP-1 at 18h ($r = 0.422$, $p = 0.041$) (figure 7a), and on the 4th day ($r = 0.67$ $p < 0.046$) (figure 7b), and of TIMP-1 on the 4th day ($r = 0.67$ $p < 0.047$) (figure 8).

DISCUSSION

The findings of our study demonstrated two patterns of collagenolytic enzyme activity (collagenolysis) in acute MI patients.

Pattern I collagenolysis of group A was characterized by higher expression of MMP-1, by different expressions of TIMP-1, and by lower expression of the MMP-1/TIMP-1 complex. In contrast, pattern II collagenolysis of group B

was represented by lower expression of MMP-1, by equivalent expression of TIMP-1, and by higher expression of the MMP-1/TIMP-1 complex.

These quantitative differences in expression of collagenolytic enzyme plasma concentrations, between the two groups of acute MI patients, were accompanied by two kinds of post-MI left ventricular function. More specifically, pattern I collagenolysis, seen in group A, was followed by left ventricular dysfunction and a greater left ventricular chamber dimensions, whereas pattern II collagenolysis seen in group B, was associated with normal left ventricular function and smaller left ventricular chamber dimensions. Additionally, at certain time points the mean values for the differences in MMP-1 and MMP-1/TIMP-1 complex plasma concentrations, between group A and group B, were statistically significant.

Furthermore, the mean values for the differences in ejection fraction and LVEDD, between both groups, were statistically significant. These changes might be attributable, at least in part, to the differential expression levels of collagenolytic enzymes, as well as to other parameters of left ventricular dysfunction such as location and size of infarction.

Relevant to our findings are recent reports indicating that MMP-1 plays a role in post-MI left ventricular remodeling. Hirohara *et al.* showed a time-dependent change in serum MMP-1 levels after acute MI that may contribute to post-MI ventricular remodeling by promoting the degradation of the extracellular matrix [10]. Additionally, Soejima *et al.* [22] have reported a negative correlation, between MMP-1 plasma levels at 7 day and 2 weeks, and left ventricular ejection fraction. Furthermore, Hojo *et al.* [23] showed a significant correlation between maximum peripheral blood mononuclear cell MMP-1 levels and maximum plasma C-reactive protein levels, and left ventricular end diastolic volume index. So our results are generally in agreement with their findings, except for the fact that we could not find any correlation, between changes in MMP-1, TIMP-1, and MMP-1/TIMP-1 complex plasma concentrations and changes in LVEF. The only correlation was between plasma concentrations of MMP-1 at 18h and day 4 and TIMP-1 at day 4, and plasma concentrations of CK. The correlation between MMP-1, TIMP-1 and MMP-1/TIMP-1 complex and the clinical parameters LVDD, and EF was estimated by performing multivariate analysis. No significant correlation was observed.

Also, one other important finding emerging from our echocardiographic study was the fact that left ventricular chamber dimension changes occurred early, on the 1st or 2nd day post-MI. Furthermore, the very early initiation of ACEI in acute MI patients, minimized post-infarction left ventricular dilatation, through a reduction in metalloproteinase enzyme plasma concentrations. This mode of action of ACEI may be considered as an alternative mechanism responsible for their beneficial effects on left ventricular expansion in the chronic phase [24].

It is well known, that cardiac myocytes and the fibrillar extracellular collagen matrix (ECCM) play a critical role in determining cardiac performance [25, 26]. In particular, ECCM provides structural support and integrity to the myocardium [27], and facilitates the conversion of myocyte contraction into pump function. The integrity of the original ECCM is thought to play an important role in determining the extent of dysfunction after myocardial

infarction [28], whereas, parallel to myocyte cell death is damage of the existing ECCM [29, 30].

Degradation of ECCM follows the activation of MMP that are secreted, as zymogen [31]. Recent reports have documented the time-dependent activation of MMP after ischemia or acute MI. MMP activation can occur within minutes of ischemia, with significant increases occurring as early as 15 minutes, and peaking 1 to 2 days after MI [9, 32, 33]. Different mechanisms have been implicated in MMP activation. It is believed that the early (< 48 hours) MMP activation is associated with zymogen activation, whereas subsequent (> 48 hours) increases are associated with newly synthesized protein [32-35].

In our results, the curve for the time-dependent changes of MMP activation (*figure 1*) showed an early (< 24 hours) and a late (5th-30th days) increase in mean values for MMP-1 plasma concentrations. This early increase is due to activation of latent, pro-MMP, to active MMP by the urokinase plasminogen activator (uPA) plasmin system [36]. Also, the late increase in MMP-1 activation may be due to the newly synthesized protein resulting from macrophage infiltration of the infarcted myocardial area [32, 33, 35].

Many studies have shown that remodeling of ECCM plays a major role in left ventricular remodeling [37, 38]. Differential regional remodeling of the ECCM contributes significantly to global left ventricular structural dysfunction after acute MI [37, 38], and plays a pivotal role in infarct expansion [37], infarcted dilation [12, 37], and progressive global left ventricular dilatation [38, 39]. The function of MMP during the healing and remodeling process of the left ventricle after acute MI has been clarified in studies using broad range MMP inhibitors and genetically modified mice. Rohde *et al.* [12] first demonstrated that *in vivo* MMP inhibition attenuates early, left ventricular dilatation, four days after experimental, acute MI in mice. Also Creemers *et al.* [40], have studied the effects of a broad range MMP inhibitor on left ventricular remodeling and infarct healing, one and two weeks after acute MI in mice. In this study, infarcted mice allocated to ilomast treatment, showed a significant decrease in left ventricular dilatation after MI, as measured by echocardiography.

Also, other studies have shown a fine balance between matrix metalloproteinases that degrade ECCM and endogenous tissue inhibitors (TIMP) that inhibits MMP, [41] and maintains normal remodeling and function, while an imbalance in their expression can result in adverse remodeling [41-44]. The functional role of TIMP-1 in the control of left ventricular geometry and cardiac function was recently studied in TIMP-1-deficient mice [45]. Echocardiography in these mice demonstrated an 18% increased in left ventricular end diastolic volume, and a 38% increase in cardiac mass at four months of age. Reduced myocardial collagen content probably accounted for the increased left ventricular dilatation. These results suggest that constitutive TIMP-1 expression contributes to the maintenance of normal left ventricular myocardial structure [45]. Furthermore, several transgenic models have been constructed that disrupt normal myocardial MMP and TIMP levels [7, 45, 46]. Kim *et al.* [46] demonstrated that cardiac-restricted, over expression of MMP-1 resulted in changes in the myocardial ECCM structure, which was accompanied by alterations in left ventricular function. Finally, in a rat MI model, MMP-1 mRNA levels increased soon after

MI, but this increase was not associated with a concomitant increase in TIMP-mRNA levels [7].

The MMP-1/TIMP-1 complex is formed in a stoichiometric, 1:1 molar ratio and forms an important endogenous system for regulating MMP activity. Its plasma levels might express the power of endogenous MMP inhibitory control of human myocardium TIMP-1 to form a complex with several MMP [47]. In the present study, we found a discrepancy between MMP-1 levels and MMP-1/TIMP-1 complex levels, while TIMP-1 levels fluctuated. The lower MMP-1/TIMP-1 levels found in group A compared to group B in acute MI patients might be explained by the loss of TIMP-mediated inhibitory control that has been reported recently in post-MI remodeling [7], in dilated cardiomyopathy [48], and in an *in vitro* system of ischemia and reperfusion [49].

Taken together, the above data suggest that the increased MMP expression and activation, coupled with the loss of endogenous MMP inhibitory control occurring early in the post-MI period can potentially contribute to post-MI left ventricular remodeling and dysfunction.

Study limitations

We could not compare plasma concentrations of collagenolytic enzymes occurring in acute MI patients with comparable MI involving the same wall, because of the small size of the study population.

A larger patient population and a longer period of observation are required to clarify the exact role of the differential expression of collagenolytic enzymes in the pathophysiology of post-infarction cardiac remodeling, and to establish correlations, between the patterns of collagenolysis and progressive cardiac enlargement.

CONCLUSION

In acute MI patients, increased MMP-1, with no change in TIMP-1, is associated with left ventricular dysfunction and dilatation, suggesting that increased collagenolytic activity contributes to loss of left ventricular function.

REFERENCES

1. Pfeffer MA, Braunwald E. Ventricular remodeling after myocardial infarction : experimental observations and clinical implications. *Circulation* 1990; 81: 1161.
2. Chareonthaitawee P, Christian TF, Hirose K, Gibbons RJ, Rumberger JA. Relation of initial infarct size to extent of left ventricular remodeling in the year after acute myocardial infarction. *J Am Coll Cardiol* 1995; 25: 567.
3. St John Sutton M, Pfeffer MA, Moye L, Plappert T, Rouleau JL, Lamas G, *et al.* Cardiovascular death and left ventricular remodeling two years after myocardial infarction: baseline predictors and impact of long-term use of captopril: information from the Survival and Ventricular Enlargement (SAVE) trial. *Circulation* 1997; 96: 3294.
4. Jugdutt BI, Michorowski BL. Role of infarct expansion in rupture of the ventricular septum after acute myocardial infarction: a two-dimensional echocardiographic study. *Clin Cardiol* 1987; 10: 641.
5. St John Sutton M, Sharpe N. Left ventricular remodeling after myocardial infarction. *Circulation* 2000; 101: 2981.

6. Jugdutt BI. Effect of captopril and enalapril on left ventricular geometry, function and collagen during healing after anterior and inferior myocardial infarction in a dog model. *J Am Coll Cardiol* 1995; 25: 1718.
7. Peterson JT, Li H, Dillon L, Bryant JW. Evolution of matrix metalloproteinase and tissue inhibitor expression during heart failure progression in the infarcted rat. *Cardiovasc Res* 2000; 46: 307.
8. Spinale FG. Matrix metalloproteinases: regulation and deregulation in the failing heart. *Circ Res* 2002; 90: 520.
9. Kikuchi K, Kadono T, Furue M, Tamaki K. Tissue inhibitor of metalloproteinase 1 (TIMP-1) may be an autocrine growth factor in scleroderma fibroblasts. *J Invest Dermatol* 1997; 108: 281.
10. Hirohata S, Kusachi S, Murakami M, Murakami T, Sano I, Watanabe T, et al. Time dependent alterations of serum matrix metalloproteinase-1 and metalloproteinase-1 tissue inhibitor after successful reperfusion of acute myocardial infarction. *Heart* 1997; 78: 278.
11. Tyagi SC, Kumar SG, Haas SJ, Reddy HK, Voelker DJ, Hayden MR, et al. Post-transcriptional regulation of extracellular matrix metalloproteinase in human heart end-stage failure secondary to ischemic cardiomyopathy. *J Mol Cell Cardiol* 1996; 28: 1415.
12. Rohde LE, Ducharme A, Arroyo LH, Aikawa M, Sukhova GH, Lopez-Anaya A, et al. Matrix metalloproteinase inhibition attenuates early left ventricular enlargement after experimental myocardial infarction in mice. *Circulation* 1999; 99: 3063.
13. Mukherjee R, Brinsa TA, Dowdy KB, Scott AA, Baskin JM, Deschamps AM, et al. Myocardial infarct expansion and matrix metalloproteinase inhibition. *Circulation* 2003; 107: 618.
14. Lindsey ML, Gannon J, Aikawa M, Schoen FJ, Rabkin E, Lopresti-Morrow L, et al. Selective matrix metalloproteinase inhibition reduces left ventricular remodeling but does not inhibit angiogenesis after myocardial infarction. *Circulation* 2002; 105: 753.
15. Greene DG, Carlisle R, Grant C, Bunnell IL. Estimation of left ventricular volume by one-plane cineangiography. *Circulation* 1967; 35: 61.
16. Bell RM, Smith CH, Reeder SG. Cardiac catheterization and angiography. In: Mayo Clinic practice of cardiology. 3rd Edition Edited by Emilio R Ciuliami, Bernard J Gerth, Michael D, Mc Goon, David L Hayes, Hartsell V. Schaff, 1966 Editor Mosby St Louis, Baltimore, Boston, London, Tokyo, Singapore, Mexico City, Sidney, Toronto, Wiesbaden Chapter 10A; 357.
17. Schiller NB, Shah PM, Crawford M, DeMaria A, Devereux R, Feigenbaum H, et al. Recommendations for the left ventricle by two dimensional echocardiography. *J Am Soc Echocardiogr* 1989; 2: 358.
18. Feigenbaum H, Popp RL, Wolfe SB, Troy BL, Pombo JF, Haine CL, et al. Ultrasound measurements of the left ventricle. A correlative study with angiography. *Arch Intern Med* 1972; 129: 461.
19. Zhang J, Fujimoto N, Iwata K, Sakai T, Okada Y, Hayakawa T. A one-step sandwich enzyme immunoassay for human matrix metalloproteinases-1 (interstitial collagenase) using monoclonal antibodies. *Clin Chim Acta* 1993; 219: 1.
20. Woolley DE, Crossley MJ, Evanson JM. Collagenase at sites of cartilage erosion in the rheumatoid joint. *Arch Rheu* 1977; 20: 1231.
21. Clark IM, Powell LK, Cawston TE. Tissue inhibitor of metalloproteinases (TIMP-I) stimulates the secretion of collagenase from human skin fibroblasts. *Biochem Biophys Res Commun* 1994; 203: 874.
22. Soejima H, Ogawa H, Sakamoto T, Miyamoto S, Kajiwa I, Kojima S, et al. Increased Serum Matrix Metalloproteinase-1 Concentration Predicts Advanced Left Ventricular Remodeling in Patients With Acute Myocardial Infarction. *Circ J* 2003; 67: 301.
23. Hojo Y, Ikeda U, Ueno S, Arakawa H, Shimada K. Expression of Matrix Metalloproteinases in Patients With Acute Myocardial Infarction. *Circ J* 2001; 65: 71.
24. Papadopoulos DP, Economou EV, Makris TK, Kapetanios KJ, Moysakakis I, Votteas VE, et al. Effect of angiotensin-converting enzyme inhibitor on collagenolytic enzyme activity in patients with acute myocardial infarction. *Drugs Exp Clin Res* 2004; 30: 55.
25. Weber KT, Anversa P, Armstrong PW, Brilla CG, Burnett Jr. JC, Cruickshank JM, et al. Remodeling and reparation of the cardiovascular system. *J Am. Coll Card* 1992; 20: 3.
26. Pelouch V, Dixon IML, Golfman L, Beamish RE, Dhalla NS. Role of extracellular Matrix Proteins in heart function. *Mol Cell Bioch* 1993; 129: 101.
27. Caulfield JB, Borg TK. The collagen network of the heart. *Lab Inves* 1979; 40: 364.
28. Thompson MM, Squire IB. Matrix metalloproteinase-9 expression after myocardial infarction: physiological or pathological? *Cardiovasc Res* 2002; 54: 495.
29. Takahashi S, Barry AC, Factor SM. Collagen degradation in ischemic rat hearts. *Bioch J* 1990; 265: 233.
30. Sato S, Ashraf M, Millard RW, Fujiwara H, Schwartz A. Connective tissue changes in early ischemia or porcine myocardium: an ultra structural study. *J Mol Cel Cardiol* 1983; 15: 261.
31. Nagase H, Woessner Jr. JF. Matrix metalloproteinases. *J Biol Chem* 1999; 274: 21491.
32. Etoh T, Joffs C, Deschamps AM, Davis J, Dowdy K, Hendrick J, et al. Spinale FG Myocardial and interstitial matrix metalloproteinase activity after acute myocardial infarction in pigs. *Am J Physiol Heart Circ Physiol* 2001; 281: H987.
33. Romanic AM, Burns-Kurtis CL, Gout B, Berrebi-Bertrand I, Ohlstein EH. Matrix metalloproteinase expression in cardiac myocytes following myocardial infarction in the rabbit. *Life Sci* 2001; 68: 799.
34. Jugdutt BI. Prevention of ventricular remodeling post myocardial infarction: timing and duration of therapy. *Can J Cardiol* 1993; 9: 103.
35. Tziakas DN, Chalikias GK, Parissis JT, Hatzinikolaou EI, Papadopoulos ED, Tripsiannis GA, et al. Serum profiles of matrix metalloproteinases and their tissue inhibitor in patients with acute coronary syndromes. The effects of short-term atorvastatin administration. *Intern J Cardio* 2004; 94: 269.
36. Jugdutt BI. Ventricular Remodeling after Infarction and the Extracellular Collagen Matrix. When is Enough Enough? *Circulation* 2003; 108: 1395.
37. Olivetti G, Capasso JM, Sonnenblick EH, Anversa P. Side-to-side slippage of myocytes participates in ventricular wall remodeling acutely after myocardial infarction in rats. *Circ Res* 1990; 67: 23.
38. Jugdutt BI. Remodeling of the myocardium and potential targets in the collagen degradation and synthesis pathways. *Curr Drug Targets Cardiovasc Haematol Disord* 2003; 3: 1.
39. Jugdutt BI, Tang SB, Khan MI, Basualdo CA. Functional impact on remodeling during healing after non-Q-wave versus Q-wave anterior myocardial infarction in the dog. *J Am Coll Cardiol* 1992; 20: 722.
40. Creemers EEJM, Cleutjens JPM, Smits JFM, Daemen MJAP. Inhibition of matrix metalloproteinase (MMP) activity in mice reduces LV remodeling and depresses cardiac function after myocardial infarction. *Circulation* 1998; 100(suppl): 1.
41. Woessner Jr. JF. The matrix metalloproteinase family. In: Parks WC, Mecham RP, eds. *Matrix Metalloproteinases*. San Diego, Calif: Academic Press, 1998: 1.
42. Mann DL, Spinale FG. Activation of matrix metalloproteinases in the failing human heart: breaking the tie that binds. *Circulation* 1998; 98: 1699.

43. Mann DL. Inflammatory mediators and the failing heart: past, present, and the foreseeable future. *Circ Res* 2002; 91: 988.
44. Fedak PW, Altamentova SM, Weisel RD, Nili N, Ohno N, Verma S, *et al.* Matrix remodeling in experimental and human heart failure: a possible regulatory role for TIMP-3. *Am J Physiol* 2003; 284: H626.
45. Roten L, Nemoto S, Simsic J, Coker ML, Rao V, Boucu S. Effects of gene deletion of the tissue inhibitor of the matrix metalloproteinase-type-1 (TIMP-1) on ventricular geometry and function in mice. *J Mol Cell Cardiol* 2000; 32: 109.
46. Kim HE, Dalal SS, Young E, Legato MJ, Wesfeldt ML, D'Armiento J. Disruption of the myocardial extracellular matrix leads to cardiac dysfunction. *J Clin Invest* 2000; 106: 857.
47. Nagase H. Activation mechanisms of matrix metalloproteinases. *Biol Chem* 1997; 378: 151.
48. Li YY, Feldman AM, Sun Y, McTiernan CF. Differential expression of tissue inhibitors of metalloproteinases in the failing human heart. *Circulation* 1998; 27: 1728.
49. Baghelai K, Marktanner R, Dattilo JB, Dattilo MP, Jakoi ER, Yager DR, Makhoul RG, Wechsler AS. Decreased expression of tissue inhibitor of metalloproteinase 1 in stunned myocardium. *J surg Res* 1998; 77: 35.