

Interleukin-1B (-511) gene polymorphism is associated with acute coronary syndrome in the Turkish population

Ozer Soylu¹, Aydin Yildirim¹, Ajda Coker², Tuna Tezel¹, Edward O. List³, Ahmet Arman⁴

¹ The Department of Cardiology, Dr. Siyami Ersek Hospital, Istanbul, Turkey

² The Department of Biology, Art and Sciences, Marmara University, Istanbul, Turkey

³ The Edison Biotechnology Institute, Ohio University, Athens, OH, USA

⁴ The Faculty of Engineering, Marmara University, Istanbul, Turkey

Correspondence : A. Arman Ph.D., The Faculty of Engineering, Advanced Production and Management Technology Building, Molecular Genetics and Cell Culture Lab, Room #: 405, Goztepe Campus, Marmara University, 34722, Istanbul, Turkey
<aarman@eng.marmara.edu.tr>

Accepted for publication February 10, 2008

ABSTRACT. *Objectives:* acute coronary syndrome (ACS) is defined as an inflammatory disease associated with development of atherosclerosis and instability. IL-1 is a candidate inflammatory cytokine that is thought to trigger ACS. The purpose of this study was to determine the relationship between IL-1 gene family polymorphisms (IL-1RN, IL-1B in positions -511 and +3953) and ACS in the Turkish population. *Methods:* a total of 381 people participated in the study, with 117 control subjects and 264 ACS patients. Of the 264 ACS patients, 112 were diagnosed with stable angina pectoris (SAP) and 152 were diagnosed with unstable angina pectoris (USAP). The polymerase chain reaction (PCR) was used to determine the genotype of IL-1RN. The genotypes of IL-1B (-511 and +3953) were determined by PCR, followed by restriction enzyme digestion of the PCR products. *Results:* there were no significant differences in both IL-1RN, IL-1B (-511 and +3953) genotype distributions and IL-1RN allele frequencies between ACS patients and the control subjects. In addition, no association was observed in the allele frequency of IL-1B (-511 and +3953) between ACS patients and controls ($p = 0.113$ and $p = 0.859$, respectively), or between SAP patients and controls ($p = 0.575$ and $p = 0.359$, respectively). However, IL-1B allele 1 (C) (-511) polymorphism in USAP patients was found to be significantly different from that of control subjects ($p = 0.041$, OR: 2.01; 95% CI: 1.985-3.933). A significant difference was also observed between USAP and SAP patients for IL-1B (+3953) allele 1 (C) polymorphism; ($p = 0.043$, OR: 1.522; 95% CI: 1.012-2.88). *Conclusion:* these results show that IL-1RN gene polymorphism has no association with ACS. However, the allele 1 (C) of IL-1B (-511) may be a risk factor for susceptibility to USAP in the Turkish population.

Keywords: interleukin-1B, interleukin-1 receptor antagonist, polymorphism, acute coronary syndrome

Acute coronary syndrome (ACS) is defined as a spectrum of clinical manifestations of acute coronary artery disease, extending from acute myocardial infarction through minimal myocardial injury to unstable angina. Coronary plaque rupture has been shown to be the real reason for sudden, acute coronary syndromes [1]. While the exact mechanism of plaque rupture remains unclear, it has been suggested that inflammatory responses play an important role in the onset, development, and evolution of atherosclerotic lesions [2] and vulnerable plaques destabilization [3]. One of the candidate inflammatory cytokines thought to play a role in the development of ACS is interleukin-1 (IL-1). Evidence for a role of IL-1 in ACS was observed when the IL-1 receptor antagonist gene (IL-1RN) was disrupted in mice [4], leading to enhancement of atherosclerotic plaque development. It has also been shown that the gene product of IL-1RN, the IL-1 receptor antagonist molecule (IL-1Ra), plays important roles in controlling circulating levels of cholesterol and in foam cell progression [5]. Moreover, the severity of atherosclerosis was shown to be decreased

in the absence of IL-1B in a mouse model of atherosclerosis (apoE-deficient mice), when these mice were also made deficient in IL-1B [6].

Interleukin-1 (IL-1) is a cytokine that is involved in immunity, sepsis, trauma, infection and inflammation [7]. The IL-1 family is made up of three members, two of which are agonists (IL-1 α and IL-1B) and one is an antagonist (IL-1Ra). IL-1Ra inhibits IL-1-induced inflammation action by binding to the IL-1 receptor type I (IL-1RI) and blocking the binding of IL-1 [8]. Both the IL-1RN and the IL-1B (IL-1B) genes are mostly polymorphic. There is a variable number of 86-base pair tandem repeats (VNTR) contained within intron 2 of the IL-1RN gene [9-11]. Single nucleotide polymorphisms (SNPs) have been determined at promoter position -511 C/T [12] and in exon 5 at position +3953 C/T in the IL-1B gene [13]. The polymorphisms within these genes are suggested to influence IL-1 expression [14]. Moreover, polymorphisms resulting in IL-1 overproduction may increase susceptibility to autoimmune diseases such as atherosclerosis [14].

DONORS AND METHODS

Subjects

A total of 264 ACS patients and 117 control subjects from the Turkish population were enrolled in this study. All individuals were of Turkish origin, nationality and all were resident in Turkey. One hundred and fifty two of the ACS patients were diagnosed with unstable angina pectoris (USAP), while 112 were diagnosed with stable angina pectoris (SAP). All of the ACS patients with USAP were scheduled for coronary angiography, experienced ischemic chest pain at rest lasting ≥ 20 minutes and demonstrated at least one of the following: new or presumably new ST-segment deviations on electrocardiogram (electrocardiographic evidence of ST-segment elevation or depression), enzyme abnormalities (creatinine kinase-MB higher than the upper reference limit in ≥ 2 samples obtained with an interval of > 6 hours), and/or troponin T ≥ 0.02 ng/mL, and also non-Q-wave myocardial infarction (MI). Study exclusion criteria were acute Q-wave MI, neoplastic disease, renal failure, acute or chronic infectious or immunological conditions, cancer, cardiogenic shock, acute pulmonary edema, history of active liver disease, recent CABG, PTCA, surgery, or gastrointestinal bleeding. During the same period, 112 patients diagnosed with SAP were enrolled. These patients were scheduled for elective angioplasty. One hundred and seventeen control subjects were recruited from healthy individuals with no history of coronary artery disease, and from patients whose coronary angiography results showed less than 30% lumen diameter reduction.

Risk factor assessment

Information on age, gender, familial coronary artery disease (CAD), smoking habit and history of hypertension was obtained by a questionnaire. Weight and height of case and control subjects were measured and body mass index (BMI) was calculated. Triglyceride (TG), total cholesterol (TC), high density lipoprotein (HDL) and low density lipoprotein (LDL) levels were determined enzymatically, and were also measured enzymatically after dextrane sulfate magnesium precipitation as previously described [15]. The presence of *diabetes mellitus* (DM) was defined as a repeated fasting glucose level $>$ glucose 126 mg/dL, or the use of anti-diabetic drugs or both. Artery blood plasma samples were collected just before the index procedure and stored at -80°C until use. hs-CRP was measured by ELISA (DSL-10-42100UsCRP ELISA KIT, Diagnostic Systems Laboratories, Inc). Plasma fibrinogen concentrations were detected using a Dade Thrombin Reactive kit (Dade Bhering).

Genotyping

Blood samples were collected and kept in tubes with EDTA. Genomic DNA isolation was performed using GENTRA DNA isolation kits (Gentra Systems, D5500). The VNTR region contained within intron 2 of the IL-1RN gene, IL-1B (-511) and IL-1B (+3953) was amplified by PCR; genotyping of the IL-1RN gene was determined by PCR as previously described [16]. Genotyping of the

IL-1B (-511) polymorphism was performed by PCR followed by restriction endonuclease digestion of the PCR products overnight with 5 U *Ava*I at 37°C [16]. Genotyping of the IL-1B (+3953) polymorphism was performed by PCR followed by restriction endonuclease digestion of the PCR products overnight with 5U *Taq*I at 65°C [16].

Statistical analysis

Mann-Whitney U and χ^2 tests were used for comparison of two groups of individuals according to ACS risk factors. Allele and genotype frequencies among cases and controls were compared with Hardy-Weinberg predictions using χ^2 analysis. Multivariate logistic regression was used to assess the independent association of the IL-1RN, IL-1B allele status with the presence of ACS and to adjust for confounding factors. The model was adjusted for age, gender, hypertension, and smoking habits. The results were expressed as odds ratio (OR) and 95% confidence interval (CI). Probability of values of $p < 0.05$ (2-sided) were considered statistically significant. SPSS 13.0 software program was used for all statistical analyses.

ETHICS APPROVAL

This research was approved by the ethics committee of Marmara University in Turkey. All human subjects' rights in this research are protected, and any necessary approval was secured from the ethics committee.

RESULTS

Baseline characteristics of the controls, ACS, SAP and USAP groups are given in *table 1*. There was a significant difference between the groups with respect to age, diabetes, gender, weight, BMI, hypertension and smoking habits. Both CRP ($p = 0.001$) and fibrinogen ($p = 0.022$) levels were significantly higher in ACS patients than healthy controls (*table 1*). A significant difference was found between USAP patients and the control subjects for CRP ($p = 0.001$), diabetes ($p = 0.01$) and fibrinogen ($p = 0.007$) levels.

None of the observed genotype frequencies deviated from the Hardy-Weinberg equilibrium. The genotype distribution and allele frequency of IL-1RN VNTR are shown in *table 2*. Three of the six alleles of the IL-1RN gene were observed in patients and the control subjects. Among these three alleles, no significant difference was observed between ACS patients and controls ($p = 0.348$), SAP patients and controls ($p = 0.577$) or USAP patients and controls ($p = 0.327$). For IL-1RN allele frequencies, no association was detected between the control group and patients: ACS ($p = 0.154$), SAP ($p = 0.400$), USAP ($p = 0.120$) (*table 2*). Genotype distribution and allele frequency for IL-1B (-511) in ACS, SAP, USAP and the control groups are shown in *table 3A*. No difference was observed in the genotype distribution of IL-1B (-511) between ACS patients and the controls ($p = 0.292$), SAP patients and controls ($p = 0.837$) or USAP patients and the control ($p = 0.129$). In addition, no significant difference was observed in frequency of the IL-1B (-511) allele between ACS patients and controls ($p = 0.113$), SAP patients and

Table 1
Baseline characteristics of the control, ACS, SAP, USAP groups in a Turkish population

	Controls (n = 117)	ACS (n = 264)	SAP (n = 112)	USAP (n = 152)
Age (years ± SD)	52.4 ± 10.9	58.3 ± 11.3*	58.9 ± 10.6*	57.8 ± 11.9
Gender				
- Male (%)	48.7	73.1*	68.8*	76.3*
- Female (%)	51.3	26.9	31.3	23.7
Height (cm)	167.3 ± 8.4	167.2 ± 6.7	167.2 ± 6	168 ± 7.2
Weight (kg)	69.7 ± 8.7	77.6 ± 10*	77.7 ± 9.2*	77.5 ± 10.6*
BMI (kg/m ²)	25.6 ± 4.5	28.1 ± 3.1*	27.2 ± 3.2*	29.1 ± 3.1*
Familial CAD (%)	33.3	40.5	38.4	42.1
Hypertension (%)	44.4	59.8*	65.2*	55.3*
DM (%)	8.9	24.2**	23.2**	25**
Smoking				
Never (%)	66.7	29.9	36.6	25
Former (%)	18.8	23.9	25	23
Current (%)	14.5	46.2*	38.4*	52*
Triglyceride (mg/dL)	132.7 ± 37.6	135.6 ± 45.6	134.4 ± 42.4	136.6 ± 48.3
HDL (mg/dL)	42.4 ± 8.4	40.3 ± 9	40.5 ± 10	40.4 ± 8.1
LDL (mg/dL)	120.7 ± 33.5	128.3 ± 36.9	127.8 ± 31.2	125.8 ± 32.6
TC (mg/dL)	192.8 ± 40.6	204.4 ± 33.1	198.6 ± 38.2	195.3 ± 39.6
Triglyceride /HDL ratio	3.3 ± 1.3	3.5 ± 1.3	3.4 ± 1.3	3.5 ± 1.3
LDL/HDL ratio	3 ± 0.9	3.2 ± 1.1	3.1 ± 1.1	3.3 ± 1.1
TC/HDL ratio	4.8 ± 1.4	5 ± 1.3	4.9 ± 1.1	5.1 ± 1.2
CRP (mg/L)	0.8 ± 0.5	1.3 ± 0.7*	0.9 ± 0.6	1.4 ± 0.7*
Fibrinogen (mg/dL)	297.7 ± 59.9	316 ± 56.8*	309.9 ± 63.7	320.5 ± 50.9*

* p < 0.05 is taken to be statistically significant, ** p < 0.001 is taken to be statistically significant.

BMI: body mass index; CAD: coronary artery disease; DM: *diabetes mellitus*; HDL: high density lipoprotein; LDL: low density lipoprotein; TC: total cholesterol; CRP: C-reactive protein.

controls (p = 0.575) or SAP and USAP patients (p = 0.155). However, a significant difference was seen for allele 1 (C) of IL-1B (-511) between USAP patients and the controls (p = 0.041).

The genotype distribution and allele frequency for IL-1B (+3953) in ACS, SAP, USAP and the control groups are shown in *table 3B*. When the genotype distribution of IL-1B (+3953) was compared between groups, no significant difference was observed between ACS patients and the controls (p = 0.965), SAP patients and the controls (p = 0.630) or USAP patients and the controls (p = 0.569). In addition, no significant difference was observed in the frequency of the IL-1B (+3953) allele between ACS pa-

tients and the controls (p = 0.859), SAP patients and controls (p = 0.359) or USAP patients and the controls (p = 0.291). However, allele 1 (C) of the IL-1B (+3953) was found to be significantly different between USAP and SAP patients (p = 0.043).

According to multiple logistic regression analysis, there was no significant correlation between IL-1 gene family genotypes and acute coronary disease when adjusted for age, gender, hypertension and smoking habits (*table 4*). However, analysis of IL-1B -511 homozygous mutants compared to homozygous wild-type showed an association between IL-1B -511 gene polymorphisms and USAP (OR: 2.451; 95% C.I.: 1.049-5.75) (*table 5*).

Table 2
Genotype distribution and allele frequency of the IL-1RN VNTR polymorphism in a Turkish population

IL-1RN	Genotypes			
	Control	ACS	SAP	USAP
1/1	59 (50.4)	149 (56.4)	60 (53.6)	89 (58.6)
1/2	45 (38.5)	96 (36.4)	44 (39.3)	52 (34.2)
2/2	13 (11.1)	19 (7.2)	8 (7.1)	11 (7.2)
P versus control		0.348	0.577	0.327
P versus USAP		0.730		
IL-1RN	Alleles			
	Control	ACS	SAP	USAP
IL-1RN 1	0.696	0.746	0.733	0.756
IL-1RN 2	0.304	0.254	0.267	0.244
P versus control		0.154	0.400	0.120
P versus USAP		0.524		

Genotypes are expressed as number of patients (proportion as a % within brackets), p values are from Fisher's exact test.

Table 3
Genotype distribution and allele frequency of the IL-1B (A) (-511) and (B) (+3953) SNP polymorphisms in ACS, SAP, USAP and the control groups in a Turkish population

A		Genotypes			Alleles	
IL-1B (-511)		C/C	C/T	T/T	C allele	T allele
Control		28 (23.9)	58 (49.6)	31 (26.5)	0.488	0.512
ACS		81 (30.7)	128 (48.5)	55 (20.8)	0.549	0.451
SAP		29 (25.9)	57 (50.9)	26 (23.2)	0.514	0.486
USAP		52 (34.2)	71 (46.7)	29 (19.1)	0.576 ^a	0.424 ^a
B		Genotypes			Alleles	
IL-1B (+3953)		C/C	C/T	T/T	C allele	T allele
Control		69 (59)	41 (35)	7 (6)	0.764	0.236
ACS		157 (59.5)	93 (35.2)	14 (5.3)	0.771	0.229
SAP		59 (52.7)	45 (40.2)	8 (7.1)	0.728	0.272
USAP		98 (64.5)	48 (31.6)	6 (3.9)	0.803 ^b	0.197 ^b

Genotypes are expressed as number of patients (proportion as a % within brackets), p values are from Fisher's exact test. USAP^a versus controls, $\chi^2 = 4.163$, 1 df ; p = 0.041, USAP^b versus SAP, $\chi^2 = 4.102$, 1 df ; p = 0.043.

DISCUSSION

The pathophysiology of unstable coronary artery disease involves inflammatory mediators in the development of an atherosclerotic plaque and in thrombus formation by platelet aggregation [1]. A number of epidemiological studies have shown that various inflammatory agents such as adhesion molecules, chemokines, growth factors and cytokines have been associated with ACS and are even implicated as markers for ACS [17]. It has been shown that both C-reactive protein (CRP) [18] and fibrinogen plasma levels are high in patients with acute coronary syndrome (ACS) when compared to control subjects [19]. In our study, a statistically significant difference was found between ACS patients and controls for CRP (p = 0.001) and fibrinogen levels (p = 0.022). Moreover, both CRP (p = 0.001) and fibrinogen levels (p = 0.007) were significantly elevated in USAP patients compared to control subjects.

One of the candidate inflammatory agents is IL-1, which stimulates the synthesis of acute phase proteins (APP) such as interleukin-6 (IL-6), fibrinogen and CRP, and is a

central mediator of inflammation [4]. The IL-1 family genes, IL-1A, IL-1B, IL-1RN are clustered and mapped to chromosome 2q [20], and are known to be largely polymorphic [9-13]. An association between IL-1 gene family polymorphisms and various autoimmune diseases has been reported among different populations [21].

The IL-1RN allele 2 has been associated with several disease states and populations. An association between the IL-1RN allele 2 and carotid atherosclerosis [22], as well as CAD in patients with type 2 diabetes [23] has been shown. A protective effect of IL-1RN allele 2 has been described for restenosis after percutaneous transluminal coronary angioplasty [24]. Moreover, an association between the IL-1RN allele 2 and myocardial infarction (MI) at a young age in an Italian population has been reported [25]; however, no significant difference was found between IL-1RN allele 2 and myocardial infarction in other Caucasian populations [26]. Although, an association between single-vessel disease (SVD) and the IL-1RN allele 2 was observed in a Caucasian population from Sheffield, UK, no association was observed for single vessel CAD

Table 4

Multiple logistic regression analysis assessing the independent association of IL-1 gene polymorphisms and the presence of ACS

All subjects		OR	95% CI	p
IL-1RN genotypes compared				
2/2 and 1/2 versus 1/1		1.220	0.724-2.056	0.4
2/2 versus 1/1		0.448	0.172-1.168	0.1
2/2 versus 1/2 and 1/1		0.463	0.183-1.172	0.1
All subjects				
IL-1B -511 genotypes compared		OR	95% CI	p
T/T and C/T versus C/C		1.420	0.782-2.578	0.2
T/T versus C/C		1.838	0.881-3.837	0.1
T/T versus C/T and C/C		0.631	0.344-1.157	0.1
All subjects				
IL-1B +3953 genotypes compared		OR	95% CI	p
T/T and C/T versus C/C		0.974	0.574-1.651	0.9
T/T versus C/C		0.958	0.299-3.072	0.9
T/T versus C/T and C/C		0.945	0.312-2.865	0.9

OR: odd ratio; CI: confidence interval. All models matched for age, gender, hypertension, smoking habits and adjusted for randomized treatment assignment.

Table 5

Multiple logistic regression analysis assessing the independent association of IL-1 gene polymorphisms and the presence of USAP

All subjects			
IL1 RN genotypes compared			
2/2 and 1/2 versus 1/1	OR	95% CI	p
2/2 versus 1/1	0.133	0.211-1.78	0.346
2/2 versus 1/2 and 1/1	0.585	0.216-1.585	0.292
	0.492	0.187-1.294	0.151
All subjects			
IL1B -511 genotypes compared			
T/T and C/T versus C/C	OR	95% CI	p
T/T versus C/C	1.898	0.889-4.051	0.6
T/T versus C/T and C/C	2.451	1.049-5.75	0.04
	1.218	0.640-2.319	0.1
All subjects			
IL1B +3953 genotypes compared			
T/T and C/T versus C/C	OR	95% CI	p
T/T versus C/C	0.134	0.076-2.236	0.6
T/T versus C/T and C/C	0.615	0.183-2.071	0.5
	0.803	0.288-2.826	0.7

OR: odd ratio; CI: confidence interval. All models matched for age, gender, hypertension, smoking habits and adjusted for randomized treatment assignment.

or multiple-vessel disease (MVD) [27] in a London, UK Caucasian population. In addition, no association was found between the IL-1RN allele 2 and CAD [27], which is consistent with our results as we observe no significant difference between IL-1RN genotypes and ACS ($p = 0.348$). While the IL-1RN 1/1 genotype was seen more frequently in the USAP group (58.6%) than in the healthy control group (50.4%), this difference did not reach statistical significance ($p = 0.327$). Furthermore, no association was found between IL-1RN alleles and ACS ($p = 0.154$), SAP ($p = 0.400$) or USAP ($p = 0.120$).

IL-1B plays an important a role in the pathogenesis of atherosclerosis. Polymorphisms in the IL-1B gene can affect the severity of and susceptibility to different diseases. One of the important SNPs located at the promoter region of the IL-1B gene is -511 C/T. It has been reported that individuals who have the IL-1B (-511) allele 2 have higher levels of IL-1RA [28]. In addition, LPS-induced IL-1B production has been shown to be increased 2-3 fold by T at -511 position [29]. IL-1B, IL-1RA and IL-6 regulate CRP production used to predict cardiovascular disease in healthy people and patients with CHD, and the increased CRP level is strongly predictive of a high risk of cardiovascular morbidity and mortality. It has been reported that males carrying IL-1B -511 T/T and -31C/C showed a higher body mass index and lean body patients with end stage renal disease and, but no association was seen with CRP level [30]. In addition, patients with hypertensive CHD carrying CRP +1059 and IL-1B -511 C/T or T/T showed reduced CRP levels compared to those carrying IL-1B-511 C/C [31]. Japanese researchers have described a relationship between IL-1B -511 polymorphism and atherosclerosis in the subclavian and intracranial arteries [32]. The relationship between IL-1B (-511) and the severity of coronary heart disease has been studied in a Chinese population [33], but no significant difference was found between IL-1B (-511) polymorphism and either myocardial infarction or CAD [26, 27, 34]. In our study, no significant difference between IL-1B (-511) genotypes and ACS ($p = 0.292$) was observed. Although the frequency of the C/C genotype of IL-1B (-511) was high in USAP groups (34.2%) compared to the controls (23.9%), it did

not reach statistical significance ($p = 0.129$). Furthermore, no significant difference was observed either between IL-1B (-511) genotypes and SAP ($p = 0.837$) or between ACS and IL-1B (-511) alleles ($p = 0.113$). However, an association was found between IL-1B (-511) allele 1 (C) and USAP ($\chi^2 = 4.163$, 1 df; $p = 0.041$). This result demonstrates that IL-1B (-511) C allele carriers had a significantly increased risk (approximately two-fold) of USAP than T allele carriers in the Turkish population (OR: 2.01; 95% CI: 1.985-3.933) (table 3A).

Another IL-1B polymorphism is located at position +3953 in exon 5, and is thought to influence IL-1 expression. While studies have found an association between IL-1B (+3953) and increased plasma levels of IL-1B [13, 35], others have found no effect or a reduction in IL-1 levels [36, 37]. It has been reported that the patients with CHD carrying the T/T genotype of IL-1B +3953 showed two-fold higher CRP levels as compared to patients carrying the T/T genotype [38]. In addition, it was shown that people carrying the heterozygote IL-1B C/T genotype showed higher level of CRP [39]. Also, lower serum hsCRP levels were detected in the carriers of the +3954T allele [30]. Furthermore, no association was found between CAD and IL-1B gene polymorphism (+3953), or between either CAD and IL-1B +3953 allele frequency or CAD and the genotype distribution of IL-1B +3953 [24, 26, 34]. In this study, we also observed no significant difference between ACS and the control groups either in genotype distribution ($p = 0.965$) or in the frequency ($p = 0.859$) of the IL-1B (+3953) allele. However, we did observe a significant difference between USAP and SAP patients as regards the allele 1 (C) of IL-1B (+3953) ($\chi^2 = 4.102$, 1 df; $p = 0.043$). Thus, IL-1B (+3953) allele 1 (C) carriers were more likely to develop USAP than T allele carriers (OR: 1.522; 95% CI: 1.012-2.88) (table 3B). It has been reported that carrying the IL-1RN allele 2 is significantly associated with coronary atherosclerosis in patients with type 2 diabetes [23]. However, other researchers did not find an independent effect of IL-1 gene polymorphisms on CAD [26, 34]. To assess an independent association of IL-1 gene family polymorphisms with ACS, we performed a multiple logistic regression model

adjusted for age, gender, hypertension and smoking habits (table 4). Although did not find an additional effect of IL-1 gene polymorphisms on ACS, we did find a significant effect on the development of USAP. In our analysis of IL-1B (-511) homozygous mutant (C/C) compared to homozygous wild-type (T/T), an association between the IL-1B (-511) gene polymorphisms and USAP was found (OR: 2.451; 95% CI: 1.049-5.75).

In conclusion, neither the IL-1RN genotype nor the IL-1RN allele was shown to have an association with ACS, but the presence of the IL-1B (-511) C allele appears to be a significant risk factor for USAP in the Turkish population. This is the first report showing an association between USAP and IL-1B (-511) gene polymorphisms.

Acknowledgments. This work was supported by a grant from The Marmara University Research Foundation. **Competing Interest Statement:** the authors of this manuscript have no competing interest or conflict of interest for any aspect of this study.

REFERENCES

1. Fuster V, Badimon L, Badimon JJ, et al. The pathogenesis of coronary artery disease and the acute coronary syndromes. *N Engl J Med* 1992; 30: 310-8.
2. Elabbassi W, Al-Nooryani A. Acute coronary syndrome. An acute inflammatory syndrome. *Saudi Med J* 2006; 12: 1799-803.
3. Ding SF, Zhang Y, Zhang M, et al. The role of atherosclerotic plaque stability and inflammation in the pathogenesis of acute coronary syndrome. *Zhonghua Xin Xue Guan Bing Za Zhi* 2006; 34: 512-4.
4. Nicklin MJ, Hughes DE, Barton JL, et al. Arterial inflammation in mice lacking the interleukin 1 receptor antagonist gene. *J Exp Med* 2000; 191: 303-12.
5. Devlin CM, Kuriakos G, Hirsch E, et al. Genetic alterations of IL-1 receptor antagonist in mice affect plasma cholesterol level and foam cell lesion size. *Proc Natl Acad Sci USA* 2002; 99: 6280-5.
6. Kirri H, Niwa T, Yamada Y, et al. Lack of Interleukin-1 β decreases the severity of Atherosclerosis in ApoE- deficient Mice. *Arterioscler Thromb Vasc Biol* 2003; 23: 656-60.
7. Dinarello CA. Proinflammatory cytokines. *Chest* 2000; 118: 503-8.
8. Auron PE. The interleukin 1 receptor: ligand interactions and signal transduction. *Cytokine Growth Factor Rev* 1998; 9: 221-37.
9. Steinkasserer A, Koelble K, Sim RB. Length variation within intron 2 of the human IL-1 receptor antagonist gene (IL-1 RN). *Nucleic Acids Res* 1991; 19: 5095.
10. Tarlow JK, Blakemore AI, Lennard A, et al. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by variable numbers of an 86-bp tandem repeat. *Hum Genet* 1993; 91: 403-4.
11. Vamvakopoulos JE, Taylor CJ, Morris-Stiff GJ, et al. The IL-1 Receptor Antagonist gene: A single-copy variant of the intron 2 variable number tandem repeat (VNTR) polymorphism. *Eur J Immunogenet* 2002; 29: 337-40.
12. di Giovine FS, Takhsh E, Blakemore AI, et al. Single base polymorphism at -511 in the human interleukin-1 beta gene (IL1 beta). *Hum Mol Genet* 1992; 1: 450.
13. Pociot F, Molvig J, Wogensen L, et al. A TaqI polymorphism in the human interleukin-1 beta (IL-1 beta) gene correlates with IL-1 beta secretion *in vitro*. *Eur J Clin Invest* 1992; 22: 396-402.
14. Arend WP. The balance between IL-1 and IL-1 Ra in disease. *Cytokine Growth Factor Rev* 2002; 13: 323-40.
15. Friedewald WT, Levi RI, Fredrickson DJ. Estimation of the concentration of low density lipoprotein cholesterol in plasma without use of the ultracentrifuge. *Clin Chem* 1972; 18: 499-502.
16. Arman A, Yilmaz B, Coker A, et al. Interleukin-1 Receptor Antagonist (IL-1RN) and Interleukin-1B gene Polymorphisms in Turkish patients with Rheumatoid Arthritis. *Clin Exp Rheumatol* 2006; 24: 643-8.
17. Corti R, Fuster V, Badimon JJ. Pathogenetic concepts of acute coronary syndromes. *Am J Coll Cardiol* 2003; 41: 7S-14S.
18. Heinisch RH, Zanetti CR, Comin F, et al. Serial changes in plasma levels of cytokines in patients with coronary artery disease. *Vasc Health Risk Manag* 2005; 1: 245-50.
19. Brunetti ND, Correale M, Pellegrino PL, et al. Acute phase proteins in patients with acute coronary syndrome: Correlations with diagnosis, clinical features, and angiographic findings. *Eur J Intern Med* 2007; 18: 109-17.
20. Nicklin MJH, Weith A, Duff WG. A physical map of the region encompassing the human interleukin-1 and the interleukin-1 receptor antagonist genes. *Genomics* 1994; 19: 382-4.
21. Dinarello CA. Interleukin-1. *Cytokine & Growth Factors Rev* 1997; 8: 253-65.
22. Worrall BB, Azhar S, Nyquist PA, et al. Interleukin-1 Receptor Antagonist gene polymorphisms in carotid atherosclerosis. *Stroke* 2003; 34: 790-3.
23. Marculescu R, Endler G, Schillinger M, et al. Interleukin 1 receptor antagonist genotype is associated with coronary atherosclerosis in patients with type 2 diabetes. *Diabetes* 2002; 51: 3582-5.
24. Francis SE, Camp NJ, Burton AJ, et al. IL-1Ra gene polymorphism and re-stenosis after coronary angioplasty. *Heart* 2001; 86: 336-40.
25. Manzoli A, Androttì F, Varlotta C, et al. Allelic polymorphism of the IL-1Ra gene in patients with acute or stable presentation of ischemic heart disease. *Cardiologia* 1999; 44: 825-30.
26. Zee RY, Lunze K, Lindpaintner K, et al. A prospective evaluation of the interleukin-1 receptor antagonist intron 2 gene polymorphism and the risk of myocardial infarction. *Thromb Haemost* 2001; 86: 1141-3.
27. Francis SE, Camp NJ, Dewberry RN, et al. Interleukin-1 receptor antagonist gene polymorphism and coronary artery disease. *Circulation* 1999; 99: 861-6.
28. Hurme M, Santtila S. IL-1 receptor antagonist (IL-1Ra) plasma levels are co-ordinately regulated by both IL-1Ra and IL-1beta genes. *Eur J Immunol* 1998; 28: 2598-602.
29. Hall SK, Perregaux DG, Gabel CA, et al. Correlation of polymorphic variation in the promoter region of the interleukin-1 beta gene with secretion of interleukin-1 beta protein. *Arthritis Rheum* 2004; 50: 1976-83.
30. Maruyama Y, Nordfors L, Stenvinkel P, et al. Interleukin-1 gene cluster polymorphisms are associated with nutritional status and inflammation in patients with end-stage renal disease. *Blood Purif* 2005; 23: 384-93.
31. Lin J-W, Dai D-F, Chou Y-H, et al. Exploring the Effects of C-Reactive Protein (CRP) and Interleukin-1 β Single Nucleotide Polymorphisms on CRP Concentration in Patients With Established Coronary Artery Disease. *Int Heart J* 2007; 48: 463-75.
32. Oda K, Tanaka N, Arai T, et al. Polymorphisms in pro- and anti-inflammatory cytokine genes and susceptibility to atherosclerosis: a pathological study of 1503 consecutive autopsy cases. *Hum Mol Genet* 2007; 16: 592-9.
33. Zhang YM, Zhong LJ, He BX, et al. The correlation between polymorphism at position -511C/T in the promoter region of interleukin 1B and the severity of coronary heart disease. *Zhonghua Yi Xue Za Zhi* 2006; 23: 86-8.

34. Vohnout B, Di Castelnuovo A, Trotta R, *et al.* IL-1 gene cluster polymorphisms and risk of coronary artery disease. *Haematologica* 2003; 88: 54-60.
35. Hall SK, Perregaux DG, Gabel CA, *et al.* Correlation of polymorphic variation in the promoter region of the interleukin-1 beta gene with secretion of interleukin-1 beta protein. *Arthritis Rheum* 2004; 50: 1976-83.
36. Dominici R, Malferrari G, Mariani C, *et al.* The Interleukin 1-beta exonic (+3953) polymorphism does not alter *in vitro* protein secretion. *Exp Mol Pathol* 2002; 73: 139-41.
37. Buchs N, di Giovine FS, Silvestri T, *et al.* IL-1B and IL-1Ra gene polymorphisms and disease severity in rheumatoid arthritis: interaction with their plasma levels. *Genes Immun* 2001; 2: 222-8.
38. Berger P, McConnell JP, Nunn M, *et al.* C-reactive protein levels are influenced by common IL-1 gene variations. *Cytokine* 2002; 17: 171-4.
39. Latkovskis G, Licis N, Kalnins U. C-reactive protein levels and common polymorphisms of the interleukin-1 gene cluster and interleukin-6 gene in patients with coronary heart disease. *Eur J Immunogenet* 2004; 31: 207-13.