

REVIEW

When wrinkles appear on the immune system can it be reversed?

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ABSTRACT. During aging, physiological and physical frailty occur, which is accompanied by a decline in adaptive and innate immunity, termed '*immunosenescence*' characterized by marked changes in the composition, function, and competence of the human immune system. Moreover, the capabilities of the immune system to defend the human body against infections, to detect and destruct malignant or autoreactive cells decline, resulting in an increase in the susceptibility to infection, development of cancer, as well as autoimmune disorders. The study of age-related changes in immune function is an important area of investigation. In this review, the function of the immune system during aging, as well as the different ways to rejuvenate the aging immune system, is explored, as medical intervention, balanced nutrition, and a healthy life style will be discussed.

Key words: immunosenescence, adaptive immunity, innate immunity, rejuvenation

INTRODUCTION

In a context of physiological and physical integrity changes with aging, immune function declines in terms of both adaptive and innate [1] with marked changes in the composition, function, and competence of the human immune system [2], all together termed as "*immunosenescence*" [3]. Functions of the immune system as defense against infections, response against new invaders, and detection and destruction of malignant or autoreactive cells, decline by aging [4]. The availability of naive T cells is substantially affected by age-related thymic atrophy, especially naive CD8+ T-cell reservoir, ending up in a status of increased susceptibility to infections and cancer and an increased incidence of autoimmune disorders [4]. In addition a condition of chronic low-grade inflammation occurs [5], leading to an imbalance between the inflammatory and antiinflammatory states due to elevated levels of pro-inflammatory mediators [6] released by effector memory and senescent T cells [7]. Moreover, dysregulation of monocytes takes place leading to reduced phagocytosis [5]. **Wagner et al.** in 2018 stated that severe infections, symptoms tend to be less in the elderly, and fever is absent in 20 to 30 percent [8]. **In 2000 Norman** suggested that there is a decrease in the ability of immune system to mount an inflammatory cytokine response when facing infection, and the signs of infections are not specific [9]. The efficient immune response is created by a complex network of cells and soluble factors grouped into the innate and adaptive

parts of the immune system, which are formed by hematopoietic stem cells (HSC) [10]. In the present review, the function of the immune system during aging is explored as well as the different ways of rejuvenating the aging immune system as medical intervention, good nutrition, and a healthy life style.

IMPACT OF AGING ON THE IMMUNE SYSTEM

Hematopoiesis and mechanism of aging

Aging is associated with profound alterations of the hematopoietic system [11] responsible for producing lymphoid and myeloid cells. The lymphoid branch produces B and T cells that orchestrate the adaptive immune response; the myeloid branch orchestrates the innate immune defense to infection through general pathogen surveillance, phagocytosis, and the inflammatory response [12] as shown in *figure 1* [13]. With aging, a decrease in the volume of hematopoietic tissue in the bone marrow occurs [14], and the regenerative capacity of the hematopoietic stem cells (HSCs) declines [15]. Also, production of myeloid and megakaryocytic cells increases resulting in impaired immune functions and elevated incidence of hematological malignancies [16]. The DNA damage response (DDR) in cells is involved in the regulation of cell cycle, cell apoptosis, and senescence [17]. A study released by Li and colleague revealed that HSCs are exposed to DDR stress because of their continuous self-renewal

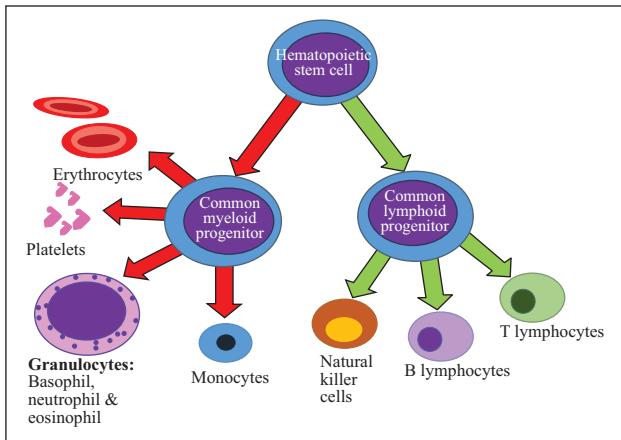


Figure 1

Hematopoiesis from a multipotent stem cell. In the elderly there is skewing toward the myeloid cell differentiation pathway (red arrows) in comparison with the less favored common lymphoid cell differentiation pathway (green arrows); this will result in a decreased production of lymphocytes [13].

process, they proposed that DNA damage may be a principal mechanism that regulates age-dependent stem cell decline [17]. As a consequence of HSC aging a lineage skewing occurs; an upregulation of myeloid-specific genes and a down regulation of lymphoid-specific genes are observed [16]. This skewing toward the myeloid lineage is thought to be due to the decreased number of progenitor cells and colony stimulating factors. Additionally, age-associated changes in the endocrine system lead to reduction in lymphopoiesis, as a result of lowered concentration of growth hormone [18].

Innate immunity

Innate immunity is an unspecific cellular response. Skin and mucous membranes constitute the first line of defense against invaders [19]. The innate immune response is mediated by monocytes, macrophages, neutrophils, dendritic cells, natural killer (NK), and Toll-like receptors [19, 20].

Skin and mucus membrane. By aging the decline of skin cell replacement and epidermal thinning inhibit the barrier function and initiate a pro-inflammatory state [21]. Subcutaneous tissue atrophy [22] and decrease in sweat and sebum production are also observed [23]. In mucous membranes, ciliated cells that remove pathogens, are reduced [24]. Many contributors are involved in skin aging such as alterations in DNA repair and stability, mitochondrial function, cell cycle and apoptosis, proteolysis induced by ubiquitin, cellular metabolism, and hormone decline [25].

Monocytes and Macrophages. The phagocytic activity of monocytes is highly affected and reduced by aging [5]. The ability of cells to produce pro-inflammatory cytokines like TNF- α , IL-1, IL-6, and IL-8 declines [26, 27]. Metcalf *et al.* in 2017 observed that cytokine secretion in response to stimulation via TLRs is greatly reduced by aging [28].

Processing and presenting antigens to T cells decline [27], and they become susceptible to the accumulation of reactive oxygen species ROS [29]. These declined functions were attributed to the telomeres shortening of macrophage leading to a decreased GM-CSF but not M-CSF-dependent proliferation of these cells as a result of decreased phosphorylation of STAT5 [29]. In addition, a decline in classical monocytes ($CD14^{+ve} CD16^{-ve}$) and an increase in intermediate ($CD14^{+ve} CD16^{+ve}$) and nonclassical monocytes ($CD14^{+ve} CD16^{++ve}$) were observed by aging [30]. Interestingly, nonclassical monocytes express high levels of miR-146a and exhibit a senescence associated with a secretory phenotype (SASP), and involved in inflamming [31].

Neutrophils. Aging affects functions of neutrophils in many aspects such as migration route [32], efficiency of phagocytosis [33], and the production of ROS “oxidative burst” [34]. Inaccurate migration was found to be associated with increased constitutive phosphoinositide 3-kinase (PI3K) signaling [35]. PI3K-blocking strategies, specifically inhibition of PI3K γ or PI3K δ , restored neutrophil migratory accuracy [35] and might help improve outcomes during infection and reduce inflammation [36]. Neutrophils showed a reduced chemotaxis to the different stimuli [37] and a reduced ability to extrude their DNA as networks of extracellular fibers (NETs) responsible to entrap bacteria extracellularly [34], causing an alteration in pathogen destruction mechanism [38]. In addition, the significant decrease in T-cell-like receptor repertoire diversity (referred to as TCRL) in old age is a mechanism of immunosenescence in neutrophils [39]. Moreover, an age-associated reduction in TLR1 expression on neutrophils was found to be associated with reduced chemokine (IL8) production, reduced rescue from apoptosis, and lower expression of activation markers, resulting from reduced bioenergetics in neutrophils [40], which explains the recurring infections [35], especially of the skin and respiratory tract in seniors [32]. Also, it was reported that the lowered production of ROS due to aging [34] leads to a reduced ability to eliminate bacteria and fungi [41].

Dendritic cells. Dendritic cells (DCs) are professional antigen-presenting cells, responsible for the first recognition of pathogens, migration to regional lymph nodes, phagocytosis, priming of naive T cells, and regulation of B and NK cells' response [42]. They are strategically placed between internal and external environment, which enables them to link innate and adaptive immunity [42]. Impaired functions of DCs by aging [43] include defective migration, phagocytosis [44], and signaling pathways [45]. They also exhibit mitochondrial dysfunction, illustrated by reduced ATP turnover and coupling efficiency, decreased baseline oxidative phosphorylation, and greater proton leak and reactive oxygen species (ROS) production [46]. It was observed that DC from older mice have a poor ability to stimulate a CD8+ T cell-mediated cytotoxic response [47], because of the reduced production of TNF- α which is a determining factor in the DC that mediate CD8+ T cell response [48].

Natural Killer cells. The NK cells are key component of innate immunity which are involved in the immune response to influenza infection [49] and vaccine [50]. They are responsible for eliminating virus-infected or tumor cells and in the regulation of the immune response by producing cytokines and chemokines that activate other cellular components of innate and adaptive immunity [51]. Changes in NK cell biology that accompany human aging are manifested as an increase in NK cell numbers, due to the increase of CD56^{dim} NK cells [52] and a decrease of CD56bright cells [53], leading to an altered NK cell subset distribution [54] and a decrease of about 75% in the amount of IFN- γ secreted by NK cells [55]. Additionally, the expression of activating NK cell receptors NKp46 and NKp30 declines with age, NK cell cytotoxicity toward cancer cells is reduced with age [52, 54] due to reduced release of perforin [54], increased reactivation rates of latent Mycobacterium tuberculosis [56], CMV seropositivity, and proinflammatory status occur [7] leading to an increased incidence of bacterial and fungal infection [57]. Conversely, aging-related functional NK cell deficiency was completely reversed by injecting soluble IL-15/IL-15R α complexes [58].

Toll-like receptors. Toll-like receptors (TLRs) are receptors that are distributed in the body [59]. By aging, a down regulation of toll-like receptors (TLRs) is observed, a process that contributes to the lack of effective recognition of invaders [60]. Research studies have implicated specific signaling pathways in altera-

tions in TLR function in the context of human aging. These include decreased PI3-kinase activity in MDDCs (associated with increased PTEN phosphorylation) [45] and an impairment in downregulation of STAT1 phosphorylation in macrophages [61]. Microglia are mononuclear immune cells of the central nervous system (CNS) [62], they express all TLRs but their expression is affected by aging, which is a risk factor of neurodegenerative diseases, such as Alzheimer's disease (AD) [63].

Adaptive immunity

Adaptive immunity is characterized by the generation of highly antigen-specific memory after encountering an antigen [64]. By aging, a reduced B and T cell responses [8] and a decrease in the production of T and B cells occur [65]. As a result the immune system becomes less able to distinguish self-belongings from nonself one [66], and autoimmune disorders become more common [67].

Thymic involution. One of the most dramatic changes that occur in the aged immune system is the involution of the thymus gland. As a result, a decrease in both stromal and thymocyte cellularity and a loss of tissue organization occur [68], the thymus function decreases [69] and production of naive T cells declines [69] along with the accumulation of memory T cells that contributes to a shift in T-cell population toward memory T-cell dominance (figure 2) [68]. Therefore, the ability to respond to new immunological challenges, including vaccines, is compromised, and

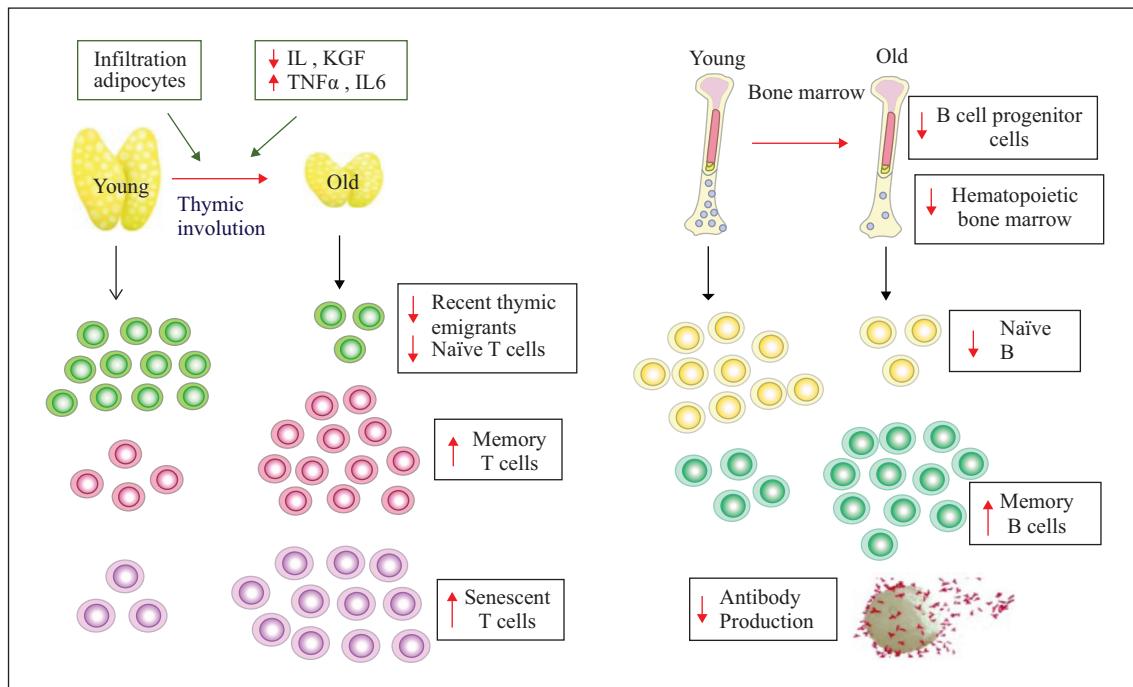


Figure 2

Age-related alterations in adaptive immune cells [68]. The thymus is located in the right upper side of the figure in yellow color as shown in young age it is bigger than that of old (getting smaller with age) the young thymus release a large amount of naive T cells shown as (green circles) small amount of memory T cells shown as (red circles) and very little amount of senescent T cells purple cells arrow to the upper side increase arrow directed to down decrease in the upper left side of the figure in aged individuals the bone marrow produces a very little amount of naive B cells shown in the figure as (yellow circles) a decrease in the antibody production and a large amount of memory B cells shown (turquoise circles).

susceptibility to infection, autoimmune diseases, and cancer increases [70].

T lymphocytes. The hallmarks of T cell immunosenescence include: accumulation of $CD28^{-ve}CD57^{+ve}$ T cells with shortened telomeres and reduced proliferative capacity [71], the lymphocyte potassium channel inhibitory pattern is altered, regulators of calcium influx kinetics contribute to the development of age-related changes of T cell function [72] and a decrease in intracellular calcium levels and alterations in the signal transduction pathways NF κ B and MAPK have been observed [73, 74], a reduction of the membrane receptors CD28 (important in lymphocyte activation) and CD27 (limited proliferative capacity by T lymphocytes) [75] occurs. It was found that decreased miR-181a is associated with age, it controls lymphocyte activation via their T cell receptor. This decline results in poor cell activation and also autoantigen recognition [76]. Endogenous p53 isoforms $\Delta 133p53$ and $p53\beta$ are physiological regulators of proliferation and senescence in human T lymphocytes. Conversely, the knockdown of $\Delta 133$ and the over expression of $p53\beta$ in CD8 + CD28+ cells inhibits cell proliferation and induced senescence [77]. T cells with senescent characteristics increase with age and exhibit constitutive p38 MAPK activation [78], due to the formation of a complex between MAP kinases and the sestrin family of proteins which results in kinase activation [79]. Furthermore, knockdown of sestrins [79] or inhibition of p38 MAP kinase restored T cell proliferative capacity [80].

Naive T cells. Naive T cells are long-lived cells and can circulate for years if they did not encounter the appropriate antigen. In old age they may die before finding their appropriate antigen [81]; therefore, the net outcome of naive T-cell output becomes reduced by aging [70]. Additionally an altered expression of costimulatory and inhibitory receptors occurs [82] causing many functional defects including impaired ability to multiply and become “effector” cells. As a result, the ability of the elderly to mount a successful immune response to new antigens is diminished, their ability to respond to infections increases, vaccine efficacy decreases [83].

Senescent Memory T cells. Memory T cells are generated when encountering an antigen. Upon reexposure to the same antigen, memory T cells recognize it and launch a vigorous response. By time, repeated exposure to antigens increases the number of protective memory T cells [84] memory T cells of seniors are often senescent, and dysfunctional as they lose the ability to proliferate and divide [83]; and secrete high levels of pro-inflammatory cytokines such as tumor necrosis factor-alpha and IL-6 [57, 83]. **Chou and Effros 2013** stated that aging of memory T cells may result from lifetime exposure to common viral infections such as cytomegalovirus (CMV) and Epstein-Barr virus (EBV) [85] that remain inactive in the body for decades, triggering a chronic, low-intensity response from memory cells and other adaptive immune cells and inducing chronic inflam-

mation. There is also an evidence that senescent memory T cells may suppress other types of immune cells [85]; they accumulate in tissues with age, causing degeneration and malignant transformation [86].

Regulatory T cells. **Jagger in 2014** revealed that regulatory T cells $CD4^{+ve}CD25^{+ve}Foxp3^{+ve}$ T cells play a pivotal role in maintaining immune homeostasis [87] by suppressing immune responses [88] they accumulate with age and weaken the immune response to pathogens or malignant cells [87]. Furthermore, their function may change with age [87, 89] as high suppressive activity could allow a greater risk of cancer, whereas impaired suppressive activity could confer a greater risk of autoimmune diseases. Thus, a balanced functioning of regulatory T cells is necessary to avoid chronic inflammation, maintaining immune protection against infections and cancer [87].

Helper T cells. CD4+ T-helper cells in the context of aging exhibit a Th2 profile (i.e., IL-4 producing cells) dominance over a Th1 profile (i.e., IFN- γ producing cells) [90]. In addition, Th17 cells that are responsible for IL-17 production and are associated with autoimmunity and inflammatory diseases accumulate by aging [91].

Cytotoxic T cells. CD8+ cytotoxic T cells mediate lytic reactivity once activated and can kill infected cells directly by the production of cytotoxins such as perforins and granzymes [92]. The accumulation of late-stage potentially differentiated cytotoxic T cells as a result of the substantial telomere erosion that affects the replicative capacity of CD8+ T cells during aging [93] is often considered a hallmark of immunosenescence [94].

B lymphocytes. Functions of B cells are affected at a mechanistic level. B-cell lymphopoiesis decreases [95], a reduced expression of genes for the differentiation occurs [96] leading to a decline in the frequency of naive B cells ($CD27^{-ve} IgD^{+ve}$) and an increase in memory B cells [97]. In addition, a reduction in crucial gene products for immunoglobulin class switch occurs which in turn affects the efficacy of humoral immune responses [96]. A decline in CD4 T cell and dendritic cell functioning, along with intrinsic changes in B cells resulting in age-associated reduction in number and size of germinal centers, are all contributing factors toward the decline in antibody production by older individuals with age [98]. One of the major manifestations of B-cell aging is the changes in their antibody repertoire including a shift in antibody production from IgG to IgM, from high affinity to low affinity, and from specificity for foreign antigens to specificity for self- antigens [71, 99]. Duggal *et al.* in 2012 revealed that immunosuppressive $CD19^{+ve} CD24^{hi} CD38^{hi}$ B cells subsets are affected in their number and function which might be a factor contributing toward increased risk of systemic autoimmunity by aging [100].

Antibody diversity. It has been stated that B-cell diversity is collapsed and decreased by aging [101]. The

immune system becomes disabled to mount an effective and specific antibodies production where a nonspecific antibody production prevails, making seniors vulnerable to infections and affecting their general health status [102]. Despite being quantitatively preserved, antibodies show impaired qualitative attachment to antigens, consequently infectious diseases such as pneumonia, influenza, infective endocarditis, and tetanus are more common among the elderly. Also, this explains why vaccines are less effective among them and why they must take booster shots [103].

PHENOTYPICAL AND FUNCTIONAL ALTERATIONS IN ELDERLY (TABLE 1)

How can immune system survive aging?

Rejuvenation strategies

Impact of lifestyle modifications on immunesenescence.

Stress Management.

Stress can be managed by meditation, mindfulness, and yoga that can promote positive emotional states [120]. Interestingly, it was found that the mind-body therapies decrease the markers of inflammation and increase the anti-viral-related immune responses and cell-mediated immunity [121], and increase IgA [122]. A recent study revealed that yoga reduces pro-inflammatory markers, as CRP and IL-1beta increases antiinflammatory cytokines such as IL-10 and could mediate inflammation at the genomic level, by changing the levels of proteins that control the DNA transcription of proinflammatory cytokine [123]. Another study found that it also improves the levels of cytotoxic T cells, B lymphocytes, and natural killer cells [124].

Sleep. Unfortunately, deprivation of sleep increases expression of genes responsible for DNA damage [125] and thus increases senescence, along with the enhancement of the proinflammatory profile [125] and a decrease in natural killer cell responses and T-cell cytokine production [125], but regular sleep patterns correlate with better immunological responses [126] and promote the redistribution of antigenic memories initially those held by antigen-presenting cells[127].

Oxytocin (Cuddle hormone). Oxytocin (OT) plays an important role in the functioning of the immune system, because the oxytocin secretory system (OSS) has a pivotal role in the development of thymus and bone marrow, strengthens immune defense, performs immune surveillance, inhibits immune damages, and maintains immune homeostasis. As a result OT inhibits inflammation, exerts antibiotic-like effect, promotes wound healing, and regenerates and suppresses immune disorders associated with stress, the OSS can release OT that act on immune system by activating OT receptors [128].

Sunlight. Sunlight is an important factor that positively impacts immune functioning and reduces the incidence of autoimmune diseases [129] and cancer [130].

Interestingly, it was revealed that sunlight energizes T cells through a separate mechanism from vitamin D production and that low levels of blue light, found in sun rays, make T cells move faster, allowing them to report T cells as either helper or killer and as the first immune cells that respond to sunlight. In addition they proved that T lymphocytes possess intrinsic photosensitivity which may enhance their motility in skin. They showed that blue light triggers the production of H₂O₂ that stimulates signaling pathway enhancing T-cell motility in vitro [131].

Water. Succeeding cold exposure (CE) leads to the increase of leukocytes, granulocytes, circulating levels of IL-6, and NK cells responses by pretreatment with exercise in water (18 °C) and thus acute CE has immune-stimulating effects [132]. **Shevchuk and Radoja** revealed that a daily brief cold stress can increase both numbers and activity of peripheral cytotoxic T-lymphocytes and NK cells, which are the major effectors of adaptive and innate tumor immunity [133].

Fasting. During normal and healthy operation, the immune system detects and removes senescent cells; this operation ceases and fails by aging, leading to the accumulation of senescent cells, which are partially responsible for aging [134]. Upon fasting, the immune system tries to save energy, by recycling a lot of the immune cells which are not needed, especially the damaged ones, fasting can help body get back to normal, and possibly even better than before fasting [134]. Another recent study by **Choi et al.**,2017 showed that long-term fasting, followed by refeeding, promotes the antiinflammatory effects, decreases the aging biological rate, and reverses a variety of autoimmune disorders as well as immunosenescence by killing old and damaged cells and replacing them with young and functional ones and causes apoptosis of autoreactive T cells, which are replaced by newly generated naive T cells during the refeeding period [135], because it reduces circulating Insulin-like Growth Factor-1 (IGF-1) levels and protein kinase A (PKA) activity in various cell populations [136]. Recently, **Rangan et al. in 2019** revealed that fasting by water only increased regenerative activity of immune system and reduces inflammatory markers [137].

Calorie restriction. A new study released by **Lee and Longo** stated that caloric restriction program of a 20–40% reduction of food consumption relative to normal intake is very effective in the regulation of aging [138]. Many studies revealed that long-term caloric restriction results in reduction of the risk of age-related diseases, including type 2 diabetes mellitus, cardiovascular disease, and cancer [139]. Other experimental studies have shown that caloric restriction maintains a youthful function of the thymus gland, reduces immune senescence during aging, increases proliferation and diversity of T cells [140], and improves multiple aspects of immune activity, particularly T-cell function [141], response to mitogens [142], and enhances the activity of both cytotoxic T-lymphocytes [143], NK cells and the ability of mononuclear cells to produce proinflammatory cytokines [142]. Periodic

Table 1
Phenotypical and functional alterations in elderly individuals

Alterations in elderly	Phenotypical	Functional
Skin	Subcutaneous tissue atrophy [22] epidermal thinning ultra-structural changes of the mucus membranes of hair cells [24]	Decreased barrier function, pro-inflammatory state, and gradual deterioration of the epidermal immune response [21]
Peripheral lymphoid tissue	Shrinkage of the volume of the peripheral lymphoid tissue [104]	Decline of absolute and relative concentrations of naive T cells in the blood [104]
Monocytes	1-No change in the number [105] 2- Increase in nonclassical monocytes express high levels of miR-146a and exhibit a senescence associated secretory phenotype (SASP) [31]	Migratory/ chemotaxis, Phagocytic, Cytotoxic abilities were reduced [105]
Dendritic cell	1-Reduced number [105] and lower maturation [48] 2-Reduction in plasmacytoid DCs and unaltered frequency of myeloid DCs [106] decline in CD141 ⁺ DCs, inducer of T helper 1 cell and cytotoxic T lymphocyte responses [107] [108]	1-chemotaxis, Phagocytic, abilities were affected [105] 2-T helper 1 cell responses and cytotoxic T lymphocyte responses were affected [108]
Neutrophils	No change in the number [105]	Migratory/ chemotaxis, Phagocytic, Cytotoxic abilities were reduced [105]
Natural killer cell	Impaired maturation [109] increase in NK cell numbers, due to expansion of CD56 ^{dim} NK cells [105, 110]	Migratory and chemotacticity were reduced [105] Impaired function of NK cells [109], slower resolution of inflammatory responses increased incidence of bacterial and fungal infection [57]
Toll like receptor	changes in expression of costimulatory markers CD80 and CD86 on monocytes showed a generalized, highly significant decrease in TLR-induced upregulation of CD80 [111]	Inappropriate persistence of TLR activation lack of effective antigen recognition [112], dysregulation of TLR signaling [112]
Peripheral lymphoid tissue	Shrinkage of the volume of the peripheral lymphoid tissue [104]	Decline of absolute and relative concentrations of naive T cells in the blood [104]
Thymus	Involution of thymus [104] replaced by connective and lipid tissues [113]	Lower number of naive T- cell and thymic hormone [114]
T- cell	Shortening of the average telomere length of T cells [104] Increase in the Number of memory and effector cells increase in the expanded clones of CD8+ and CD4+ memory T cells (oligoclonal expansion) and regulatory T cells [18] Decrease in the Number of naïve T cells [18]	Lymphocyte decline in their proliferative capacity [115, 116] Increase in the levels of pro- inflammatory cytokines [18] Ability of the elderly to mount a successful immune response to new antigens is diminished [83]
B cell	Decrease in the Expression of costimulatory molecules (CD27, CD40) [18] Decline in the frequency of naïve B cells (CD27 ^{-ve} IgD ⁺) and an increase in memory B cells [97] [118] Decline in the immunosuppressive CD19 ⁺ CD24 ^{hi} CD38 ^{hi} B cells [100]	Antibody affinity and specificity as well as diversity, decreases [117] Impaired B cell responses and defective antibody production reflected in a reduced ability to effectively respond against viruses and bacteria [119] Decrease of Isotype switching leading to the risk of autoantibody responses and autoimmunity [18, 100]

dietary restrictions were found to reverse age-dependent immune dysfunction by killing autoimmune cells [144] and could rejuvenate aged HSCs by reduction of the nutrient supply and the clearance of senescent cells [15]. Calorie restriction reduces oxidative stress by SIRT3-mediated SOD2 activation [145].

Mediterranean Diet. The Mediterranean diet is a dietary pattern based on foods and drinks traditionally consumed by people in the region surrounding the Mediterranean Sea [146]. This diet has been shown to protect against several age- and inflammation-related conditions including diabetes, atherosclerosis, obesity, cancers, and neurodegenerative diseases. It is characterized by inclusion of olive oil, fruits, vegetables, legumes, whole grains, nuts, and seeds; with moderate amounts of fish, poultry, cheese, yogurt, and eggs; limited inclusion of red meat, processed food, and foods rich in refined sugars [147]. The adoption of the Mediterranean diet by older adults has also been

associated with improvement in immune responses, particularly dendritic cell function [148].

Exercise. Reduced physical activity leads to increased adiposity and systemic inflammation [149]. There is a link between aging of immune system and inflammation [150], but regular physical activity has been associated with exert antiinflammatory and anti immunosenescence effects, potentially delaying the health declines [151] lowering the levels of pro-inflammatory cytokines such as IL-6, TNF- α in the elderly [152] and improving neutrophil chemotaxis [153]. Regular exercise has been shown to improve neutrophil microbial functions which reduces the risk of infectious disease [151], with increased immune cell telomere length, improved vaccine responses and immune system function [154]. **Takahashi and colleague** revealed that moderate walking reduces basal levels of ROS leading to an improved immune-surveillance in seniors [155]. Exercise and aerobics

are thought to mobilize aged T cells into the blood [156] leading to their degradation, allowing more space for naive T cells [126] and induce changes in immune cell numbers and functions. A clear response of CD4+ T cells, rather than CD8+ T cells or NK cells, was noted [157]. In addition aerobics improved immune response and reduced inflammatory cytokines [6]. Recently, **Duggal *et al.*** observed that aged cyclists had higher levels of IL-7, IL-15 which are thymo protective cytokines that maintain naive T cells in the periphery and lower levels of IL-6 which causes thymus involution [158].

Vitamins. Micronutrients are important for natural defenses on three levels:

- To support physical barriers (skin/mucosa): Vitamins A, C, E and the trace element zinc assist in enhancing the skin barrier function [111, 159].
- To support cellular immunity: vitamins A, B6, B12, D, C, E, folic acid and the trace elements zinc, iron work in synergy to support cellular and humoral immune responses [159].
- Antibody production: The vitamins A, B6, B12, D, E and folic acid and the trace elements zinc are essential for antibody production [111, 159].

Vitamin A

By aging skin becomes thinner resulting in a decreased barrier function because of the decrease in the protein transforming growth factor beta that controls many cellular functions. As a result dermal fibroblasts will not be converted into fat cells, and cells will not produce the antimicrobial peptide cathelicidin, which protects against bacterial infections [160]. Vitamin A increases the epidermal thickness by the upregulation of transcription factors, collagen genes responsible for wound healing, improving extracellular matrix micro-environment, extracellular matrix production, and activation of dermal fibroblasts in elderly [161]. Vitamin A deficiency leads to a poor immune response to infection especially trans retinoic acid (RA), which is an active form of the vitamin that ensures immune homeostasis through regulating cell homing and differentiation. During infections and autoimmune disease, it activates and enhances T-cell responses [162] to cancer, infections, intestinal inflammation, and immune-mediated diseases as autoimmune diseases associated with aging [162].

Vitamin B12 and folate

Vitamin B₁₂ and folate are associated with preventing chronic diseases associated with aging through the methylation of homocysteine [163]. Elevated homocysteine and lower vitamin B12 levels impact the immune system, causing increased inflammation and antioxidant agents damage that catalyzes physiological aging in all systems [164]. A recent study by **Laird *et al.*, (2018)** revealed that adults over 50 are at risk of deficiency in vitamin B12 and folate because their ability to absorb B12 from food decreases; thus, they recommended that B12 must be obtained from supplements and B12-fortified foods like cereals [165]. It is worth noting that most of elderly suffer

from megaloblastic anemia, due to vitamin B12 deficiency [166].

Vitamin E

Recently, it was found that aged people have lower levels of vitamin E and magnesium that was found to associate age-related diseases [167]. Many studies recommend daily vitamin E supplementation in the elderly, as it enhances the differentiation of immature T cells in thymus [168], the function of immune cells as neutrophils, T-cells, B-cells, and NK-cells [169], and the age-associated decline in naive T-cell function [170]. Moreover, it was found that vitamin E regulates lipid rafts and membrane fluidity on the surface of immune cells in naive CD4⁺ T cells of old mice; it also improves age-related early T-cell signaling events in naive helper T cells [171]. As a result, resistance to infection increases [169] and reduces the risk of influenza virus in elderly [168].

Vitamin D

Vitamin D (Vit. D) is an important factor for immune functioning because it stimulates the clearance and phagocytosis of macrophages, and protects immune cells against apoptosis by regulating both extranuclear protein functions and gene expression signaling [172]. Vitamin D intake decreases with age [173], in early life, exposure to the sun activates pre-vitamin D under the skin, by aging, the body composition changes and thus pre-vitamin D is lost. As a result, older people produce less Vit. D even if they get plenty of sunlight [173], where supplementation with Vit. D modulates aging-related systemic inflammation including immunosenescence [174]. In addition it was found that magnesium is essential for vitamin D metabolism. Otherwise vitamin D will remain stored and inactive [175]. Vit. D has also emerged as a key modulator of a range of immune functions including monocyte differentiation into macrophages, less production of pro-inflammatory cytokines by macrophages inhibition of Th1 and Th17 responses, and regulation of B-cell proliferation [176].

Vitamin C

Vitamin C supplementation maintains immune function as we age [177], supports the functions of both the innate and adaptive immune systems, plays an important role in the defense against bacteria and viruses [178], and reduces the duration of common cold [177]. In addition to stimulating immunity, vitamin C also appears to restrain excessive immune activity, perhaps in part by interfering with the synthesis of inflammatory cytokines [178].

Zinc. Zinc is a trace element required for multiple immune cell tasks including suppression of production of pro-inflammatory cytokines (IL1 β , TNF α) by monocytes/macrophages and reduction of reactive oxygen species (ROS) [179]. Additionally, it was found that supplementation of Zinc for six months improves antiinflammatory cytokines profile [180], and decreases the markers of inflammation like IL-6 and C-reactive protein [181]. Another study on older individuals in nursing homes residents observed that

older individuals with normal zinc levels had lower incidence of both the pneumonia and antibiotic usage compared with zinc-deficient individuals [182]. Zinc supplementation has been shown to reduce infection incidence in older adults [180] and has many effects indicative of reversed immunosenescence including: improved NK cell cytotoxicity [183], modification of Th1/Th2 balance [184], restores the serum thymulin activity [185] and improves vaccine responses [186].

Iron. **Mocchegiani et al., 2012** revealed that iron is essential for maintaining immune and antioxidant function during aging [187]. In chronic states of inflammation such as obesity or aging, iron status becomes impaired [188] and malnutrition exacerbates these effects of inflammation [189].

Protection strategy

Improving vaccine efficacy in elderly. Elderly individuals are more prone to severe infections and less responsive to vaccination than the young [190], in terms of the titer, efficacy, and affinity of antibody production [191] and responses which are characterized by shorter duration [192] due to immunosenescence [193]. The vaccination against seasonal influenza, pneumococcal disease, and reactivation of varicella zoster virus (VZV), in addition to regular booster shots against tetanus, diphtheria, pertussis, polio are recommended in the elderly. An early study revealed that the vaccination to influenza drops to 30–50% for those over 65 years of age [194], in addition vaccination to pneumococcal disease is recommended for seniors [195] and it could reduce 63.8% of pneumococcal pneumonia in elderly [196] and that 23-valent pneumococcal polysaccharide vaccine reduces the incidence of all pneumococcal pneumonia [197]. Herpes Zoster (HZ) is a major cause of hospitalization in the elderly, the recombinant vaccine containing AS01B adjuvant, has been licensed in 2017 [198], it elicits a robust and persistent memory response in older adults [199], because it induces antibody and T-cell responses in the elderly [200]. Efficacy of HZ vaccine is about 97% in the prevention of HZ in older adults [201]. Adjuvants such as AS01 can strongly activate elements of innate immunity [202]. These effects may underlie the strong, long-lasting efficacy demonstrated against herpes zoster even in very old individuals [203]. Vaccine efficacy against hepatitis B virus (HBV) and A virus (HAV), tick-borne encephalitis (TBE), is significantly reduced in vaccinated individuals \geq 70–80-year-old [204]. Booster intervals should be shortened for persons over 65 years because of the rapid decline in antibodies with advancing age [103]. Development of vaccine adjuvants specifically designed to optimally stimulate the aging immune system, the most currently used in the elderly are MF59 and AS03, included in influenza vaccines, and AS02 used for the recombinant HZ vaccine [205].

Cytokines, hormones, and pharmacological intervention strategies. Restoration of the T-cell pool balance should be done by stimulating the production of naive T cells and exporting them to the periphery by using

cytokines and hormones such as growth hormone and interleukin 7 [84]. **Montecino-Rodriguez and colleagues 2013** revealed that the manipulation of the concentration of various cytokines including interleukin-7, IL-2 and hormones such as sex steroids, growth hormone and keratinocyte growth factor (KGF) has been shown to be promising for the rejuvenation of aged thymus [84].

IL-7. IL-7 is a cytokine that is produced by stromal cells and the thymus. It plays a pivotal role in supporting thymocytes development, stimulates peripheral T-cell survival and expansion through the induction of Bcl-2. It is thought to influence T-cell progenitors directly [206]. Many experimental studies carried out in old animals reported that IL-7 reversed thymic atrophy, increased thymopoiesis improved thymic output, boosting immune function [207].

IL-2. Interleukin-2 aids in the development of thymocytes, the survival and proliferation of T cells, increases the proportion of thymocytes that differentiate into mature T lymphocytes. Many studies suggested that it reverses thymic atrophy [208].

IL-15/IL-15R α . IL-15 plays a critical role in the development and homeostasis of naive CD8 T cells, memory T cells, NK cells [209]; aging-related functional NK cell deficiency was completely reversed by injecting soluble IL-15/IL-15R α complexes [210]. **Gangemi et al., (2005)** found higher levels of IL-15 in ultralongevel subjects denoting their strong immune system that is able to defend itself from infections through efficient immune-inflammatory responses which is crucial for their longevity [211].

Gene therapy intervention. Telomere length could be restored by reprogramming telomeres to increase the cellular life span which can be induced by using pharmacological agents or gene therapy in order to increase telomerase activity [212].

Adoptive therapy. Adoptive therapy is based on the replacement of senescent cells with effector or naive cells [213].

Hormone treatment

Fibroblast growth factor 7 (FGF7) .

Aging is associated with increased expression of the tumor suppressor gene Ink4a, (**Montecino-Rodriguez et al., 2013**) and his working group showed that fibroblast growth factor 7 administration (FGF7) which is a hormone that acts on thymocytes, down-regulates (Ink4a) in T cell progenitors and rejuvenates a partial part of the involuted thymus [84] and reverses the effects of aging [214].

Dehydroepiandrosterone (DHEA)

Dehydroepiandrosterone (DHEA) is a steroid hormone that is important for the functioning of immune system [215], by aging DHEA levels decline markedly [216]. A study released by **Prall and Muchlenbein, 2015** revealed that the presence of DHEA levels in the saliva

has a bactericidal activity, and acts as a measure of innate immune function [217]. Supplementation with DHEA improved immune parameters, including monocyte levels, B- and T-cell function, and NK-cell levels [218].

Natural products strategy that improves immune system in elderly

Probiotics. Probiotics may have a particular application in elderly populations, especially in terms of protection against infections and perhaps also in the prevention of several age-related diseases [219]; they restore the gut flora and reduce chronic inflammation and pathogen colonization within the host [220]. Many clinical trials have shown that probiotics enhance immune function in reducing the recurrency and the severity of infectious diseases [83]. A study by Toward *et al.* observed that *Bifidobacteria* decreases after 60 years of age, and this decrease was correlated with the occurrence of immunosenescence [221]. The administration of *B. bifidum* exerts an anti-senescence by reducing p16 expression in thymus and spleen; additionally, it has antiinflammatory effects (lower IL6 and TNF α levels) in old mice [222].

Yogurt. Yogurt has immunostimulatory effects. **Maydani and Han** stated that increased yogurt consumption, in immune compromised populations such as the elderly, enhances the immune response, and increases resistance to immune-related diseases [223]. A study by **Nagai** revealed that yogurt intake increases interferon- γ production and augments the natural killer cell activity in his experimental design [224]. Another study in humans showed that yogurt's bacterial components were able to decrease the incidence of the common cold in elderly people when administered daily [225].

Beta-Glucan. Beta-glucans are polysaccharides found in the cell walls of fungi, bacteria, oats, and algae. They are immunomodulators of immune activity with notable anti-tumor and antimicrobial properties [226], medicinal mushrooms such as Reishi are rich in Beta-glucans [227]; they have anti-tumor, and cell-killing effects. A portion of beta-glucan reishi polysaccharides has been found to stimulate both innate and adaptive immune responses [228], regarding innate immunity it was found to enhance both the killing and phagocytic activities of NK cells [229] and macrophages [230]. Regarding adaptive immunity it was found that beta glucan have the ability to influence T-cells activation and proliferation [231, 232] and B cell to proliferate and increase the secretion of immunoglobulin [233].

Cistanche deserticola (C. deserticola). It is a plant used in traditional medicine as a remedy for chronic infections. Many studies on aged animals observed that an extract of this plant extended lifespan and reversed many laboratory indicators of immune senescence, such as the increases in the population of naive T cells and NK cells, reductions in memory T cells in the peripheral blood, and decreased levels of the

inflammatory cytokine interleukin-6 (IL-6) [234]; increased helper T cells, and improved NK cell activity [235].

Pu-erh tea extract. An early study on aged mice [236] revealed that supplementation with Pu-erh tea extract increased the fraction of naive T cells CD8(+)CD28(+) whether helper or cytotoxic T cells, and NK cells. In addition, elevated levels of the inflammatory cytokine IL-6 decreased by 43% [236], increased the resistance to infection and cancer in aging individuals [236]. Another study by (**Chu, 2011**) found that the Pu-erh tea extract decreased CRP, TNF- α , and IL-6 while levels of IL-10, an anti-inflammatory molecule, increased [237].

Garlic Extract. Garlic has immune-modulating and immune-stimulatory properties, as well as anti-tumor effects [238], it stimulates immune function by increasing macrophage activity, and production of T and B cells [239] as well as increased proliferation and activity of cytotoxic T-cells and NK-cells [240]. A recent study found that garlic supplements reduce the number, duration, and severity of upper respiratory tract infections [239] and it was associated with reduced cold and flu severity [240].

Quercetin. Quercetin, a bioflavonoid found in foods such as onions, apples, berries, and green tea, has potent antiinflammatory and free radical-scavenging properties. It has been shown to induce cell death in senescent cells, decreasing their numbers in human fat tissue cultures [241].

Tomatoes. Tomatoes are rich in lycopene [242]. **Riso *et al.*** stated that lycopene found in tomatoes slows down aging by stopping free radicals from binding with oxygen, a process that slows immune building, cleansing, and repair, and that consumption of tomatoes has been shown to be associated with a lower risk of several types of cancer [243]. Another study found that TNF- α levels were 34.4% lower in the subjects who drank tomato juice in comparison to those who do not [244]. These studies pointed out to the potential antiinflammatory effect of tomato [245].

Berries. Berries and their phytochemicals are immunomodulators that delay cancer development and contain a wide spectrum of phytochemicals that influence the functions of multiple immune cells and many aspects of cancer immunity [246] by boosting T cells to recognize tumor cells and destrucit it. Recognition occurs after dendritic cells present antigen, such as tumor antigen, to T cells, generating an adaptive response [246]. NK cells are an essential component of innate immunity against cancer development [247]. **Pan *et al.*, 2018** demonstrated that an increased number of NK cells and enhancement in their cytotoxicity occur after black raspberry intervention [246].

Broccoli. **Carr and Maggini , 2017** showed that sulforaphane, which is a key bioactive compound in broccoli, has the capacity to slow the biochemical

process of aging [248] by increasing the expression of antioxidant genes [249] in addition it was demonstrated that Kaempferol, which is a flavonoid in broccoli, has a strong anti-inflammatory capacity [250].

CLINICAL TRIALS TO REJUVENATE IMMUNESENESCENCE

A National Institute of Aging sponsored randomized trial of a 2 year caloric restriction (CR) regimen in healthy humans (CALERIE) revealed that CR slow of biological ageing by reducing cardiovascular disease risk biomarkers and decreasing the levels of pro-inflammatory cytokines [251, 252]. Another study with 125 older subjects, RISTOMED, observed that Mediterranean diet has an anti-inflammaging effect [253]. Regarding probiotics a study on aged individuals reported that probiotic consumption for 6 months increased the number of naive T cells and decreased the number of senescent CD8 CD28^{null} T cells [254]. Another trial revealed that healthy seniors that consume fermented dairy product for 3 months have a reduced risk of respiratory infection [255]. Regarding vitamins, a study by (Shelbaya *et al.*, 2017) observed that the majority of elderly that live in nursing home or homebound are at risk of developing vitamin D deficiency because of lack of exposure to sunlight [256]. Concerning exercise (El-Sabbagh and colleagues, 2015) revealed that TNF alpha and CRP that are responsible for low grade inflammation in elderly were decreased significantly after training in aged healthy Egyptians [257]. Another study showed that the exercise intervention gave a significant benefit with respect to antibody titer prior to an influenza vaccination and that exercise can improve immune responses and specifically vaccination in the aged [258].

CONCLUSIONS

The fountain of youth is not a myth. After all methods explored in this review as current evidence underlines the importance of consuming a healthy diet best exemplified in the Mediterranean diet “which contains all the essential micronutrients”, exercising, avoiding stress and sleeping well would slow down immunosenescence but are unlikely to reverse the decline of immunity completely, because they are only targeting a specific aspect of immunosenescence rather than stopping it from occurring. Nevertheless, improvement in immune function is beneficial to the elderly population, hence improving the quality of life in seniors.

RECOMMENDATIONS

By discussing the different aspects of immunosenescence and the strategies that can reduce it we recommend:

1. By easing stress cortisol levels decrease and sleeping patterns will be improved thus improving immune function.

2. Eating healthy nutrients, such as fruits, vegetables rich in beta-carotene, vitamins C and E, and zinc.
3. Probiotics are very important because they protect against infections and prevent several age-related diseases.
4. Regular physical activity promotes circulation and relaxes the body and mind. As walking, yoga classes are anti-immunesenescence and anti-inflammaging therapy that boost a senior’s immune system performance .
5. The elderly should receive regular booster vaccines with shortened vaccination intervals to maximize their immune response, especially vaccination against seasonal influenza, pneumococcal disease, and reactivation of varicella zoster virus (VZV), in addition regular booster shots against tetanus, diphtheria, pertussis, polio.
6. As elderly thirst less so they must stay hydrated because water keeps mucous membrane moist and lowers the chance of colds.

REFERENCES

1. Keenan CR, Allan RS. Epigenomic drivers of immune dysfunction in aging. *Aging Cell* 2019; 18(1):e12878.
2. Nikolich-Zugich J. The twilight of immunity: emerging concepts in aging of the immune system. *Nat Immunol* 2018; 19 : 10-9.
3. Weyand CM, Yang Z, Goronzy JJ. T-cell aging in rheumatoid arthritis. *Curr Opin Rheumatol* 2014; 26(1):93-100.
4. Agarwal S, Busse PJ. Innate and adaptive immunosenescence. *Ann Allergy Asthma Immunol* 2010; 104(3):183.
5. Pence DB, Yarbro RJ. Classical monocytes maintain *ex vivo* glycolytic metabolism and early but not later inflammatory responses in older adults. *Immun Ageing* 2019; 16 : 3.
6. Abd El-Kader SM, Al-Shreef FM. Impact of aerobic exercises on selected inflammatory markers and immune system response among patients with sickle cell anemia in asymptomatic steady state. *Afr Health Sci* 2018; 18(1):111-9.
7. Campos C, Pera A, Sanchez-Correa B, *et al.* Effect of age and CMV on NK cell subpopulations. *Exp Gerontol* 2014; 54 : 130-7.
8. Wagner A, Garner-Spitzer E, Jasinska J, *et al.* Age-related differences in humoral and cellular immune responses after primary immunisation: indications for stratified vaccination schedules. *Sci Rep* 2018; 8(1):9825.
9. Norman DC. Fever in the elderly. *Clin Infect Dis* 2000; 31 (1):148.
10. Morrison SJ, Spradling AC. Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell* 2008; 132 : 598-611.
11. de Haan G, Lazare SS. Aging of hematopoietic stem cells. *Blood* 2018; 131 : 479-548.
12. Weng NP. Aging of the immune system: How much can the adaptive immune system adapt? *Immunity* 2006; 24 : 495-9.
13. Geiger H, Denkinger M, Schirmbeck R. Hematopoietic stem cell aging. *Curr Opin Immunol* 2014; 29 : 86-92.
14. Ogawa T, Kitagawa M, Hirokawa K. Age-related changes of human bone marrow: a histometric estimation of proliferative cells, apoptotic cells, T cells, B cells and macrophages. *Mech Ageing Dev* 2000; 117 : 57-68.

15. Lee J, Suk Ran Y, Inpyo C, Haiyoung J. Causes and mechanisms of hematopoietic stem cell aging. *Int J Mol Sci* 2019; 20(6):1272.
16. Rundberg Nilsson A, Soneji S, Adolfsson S, Bryder D, Pronk CJ. Human and murine hematopoietic stem cell aging is associated with functional impairments and intrinsic megakaryocytic/erythroid bias. *PLoS One* 2016; 11 : e0158369.
17. Li T, Zhou ZW, Ju Z, Wang ZQ. DNA damage response in hematopoietic stem cell ageing. *Genom Proteom Bioinform* 2016; 14 : 147-54.
18. Lydyard P, Whelan A, Fanger M. *BIOS Instant Notes in Immunology*. Taylor & Francis, 2011.
19. McDonald DR, Levy O. Innate Immunity Clinical Immunology (Fifth Edition). *Principles and Practice* 2019;.
20. Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. *Annu Rev Immunol* 2011; 29(1):235-71.
21. Kinn PM, Holdren GO, Westermeyer BA, et al. Age-dependent variation in cytokines, chemokines, and biologic analytes rinsed from the surface of healthy human skin. *Sci Rep* 2015; 5 (1):10472.
22. Chilosì M, Facchetti F, Caliò A, et al. Oncogene-induced senescence distinguishes indolent from aggressive forms of pulmonary and non-pulmonary Langerhans cell histiocytosis. *Leuk Lymphoma* 2014; 55(11):2620-6.
23. Grewe M. Chronological ageing and photoageing of dendritic cells. *Clin Exp Dermatol* 2001; 26 : 608-12.
24. Kim SW, Mo JH, Kim JW, et al. Change of nasal function with aging in Korean. *Acta Otolaryngol* 2007; 558 : 90-4.
25. Makrantonaki E, Zouboulis CC. Molecular mechanisms of skin aging: state of the art. *Ann N Y Acad Sci* 2007; 1119 : 40-50.
26. Davalos AR, Coppe JP, Campisi J, Desprez PY. Senescent cells as a source of inflammatory factors for tumor progression. *Cancer Metastasis Rev* 2010; 29(2):273-83.
27. Shi C, Pamer EG. Monocyte recruitment during infection and inflammation. *Nat Rev Immunol* 2011; 11(11):762-74.
28. Metcalf TU, Wilkinson PA, Cameron MJ, et al. Human monocyte subsets are transcriptionally and functionally altered in aging in response to pattern recognition receptor agonists. *J Immunol* 2017; 199 : 1405-17.
29. Sebastian C, Herrero C, Serr AM, Lio Beras J, Blasco MA, Celada A. Telomere shortening and oxidative stress in aged macrophages results in impaired STAT5a phosphorylation. *J Immunol* 2009; 183(4):2356-64.
30. Seidler S, Zimmermann HW, Bartneck M, Trautwein C, Tacke F. Age-dependent alterations of monocyte subsets and monocyte-related chemokine pathways in healthy adults. *BMC Immunol* 2010; 21 : 11-30.
31. Ong SM, Hadadi E, Dang TM, et al. The pro-inflammatory phenotype of the human non-classical monocyte subset is attributed to senescence. *Cell Death Dis* 2018; 9 : 266.
32. Chen MM, Palmer JL, Plackett TP, Deburghgraeve CR, Kovacs EJ. Age-related differences in the neutrophil response to pulmonary *Pseudomonas* infection. *Exp Gerontol* 2014; 54 (1):42-6.
33. Alonso-Fernandez P, Puerto M, Maté I, Reibera JM, de la Fuente M. Neutrophils of centenarians show function levels similar to those of young adults. *J Am Geriatr Soc* 2008; 56 : 2244-51.
34. Hazeldine J, Harris P, Chapple IL, et al. Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals. *Aging Cell* 2014; 13(4): 690-8.
35. Brinkmann V, Zychlinsky A. Beneficial suicide: why neutrophils die to make NETs. *Nat Rev Microbiol* 2007; 5(8): 577-82.
36. Naccache PH, Lefebvre JS. A straight neutrophil path to healthy aging? *Blood* 2014; 123 : 154-6.
37. Sapely E, Greenwood H, Walton G, et al. Phosphoinositide 3-kinase inhibition restores neutrophil accuracy in the elderly: toward targeted treatments for immunosenescence. *Blood* 2014; 123 : 239-48.
38. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science* 2004; 303(5663): 1532-5.
39. Fuchs TA, Brill A, Wagner DD. Neutrophil extracellular trap (NET) impact on deep vein thrombosis. *Arterioscler Thromb Vasc Biol* 2012; 32 : 1777-83.
40. Qian F, Wang X, Zhang L, et al. Age-associated elevation in TLR5 leads to increased inflammatory responses in the elderly. *Aging Cell* 2012; 11 : 104-10.
41. Wessels I, Jansen J, Rink L, Uciechowski P. Immunosenescence of polymorphonuclear neutrophils. *Sci World J* 2010; 10 : 145-60.
42. Agrawal A, Gupta S. Impact of aging on dendritic cell functions in humans. *Ageing Res Rev* 2011; 10 : 336-45.
43. Gupta S. Role of dendritic cells in innate and adaptive immune response in human aging. *Exp Gerontol* 2014; 54(1): 47-52.
44. Del Prete A, Vermi W, Dander E, et al. Defective dendritic cell migration and activation of adaptive immunity in PI3K κ deficient mice. *EMBO J* 2004; 23 : 3505-15.
45. Agrawal A, Agrawal S, Cao JN, Su H, Osann K, Gupta S. Altered innate immune functioning of dendritic cells in elderly humans: a role of phosphoinositide 3-kinase-signaling pathway. *J Immunol* 2007; 178 : 6912-22.
46. Chouquet CA, Thacker RI, Shehata HM, et al. Loss of phagocytic and antigen cross-presenting capacity in aging dendritic cells is associated with mitochondrial dysfunction. *J Immunol* 2015; 195(6):2624-32.
47. Zaccia ER, Crespo MI, Acland RP, et al. Aging impairs the ability of conventional dendritic cells to cross-prime CD8+ T cells upon stimulation with a TLR7 ligand. *PLoS One* 2015; 10 (10):e0140672.
48. Liu WM, Nahar TE, Jacobi RH, et al. Impaired production of TNF-alpha by dendritic cells of older adults leads to a lower CD8+ T cell response against influenza. *Vaccine* 2012; 30 (9):1659-66.
49. Przemyska-Kosicka A, Childs CE, Maidens C, et al. Age-related changes in the natural killer cell response to seasonal influenza vaccination are not influenced by a probiotic: a randomised controlled trial. *Front Immunol* 2018; 9 : 591.
50. Weiskopf D, Weinberger B, Grubeck-Loebenstein B. The aging of the immune system. *Transpl Int* 2009; 22(11):1041-50.
51. Camous X, Pera A, Solana R, Larbi A. NK cells in healthy aging and age-associated diseases. *J Biomed Biotechnol* 2012; 2012 : 195956.
52. Almeida-Oliveira A, Smith-Carvalho M, Porto LC, et al. Age-related changes in natural killer cell receptors from childhood through old age. *Hum Immunol* 2011; 72 : 319-29.
53. Solana R, Campos C, Pera A, Tarazona R. Shaping of NK cell subsets by aging. *Curr Opin Immunol* 2014; 29 : 56-61.
54. Hazeldine J, Hampson P, Lord JM. Reduced release and binding of perforin at the immunological synapse underlies the age-related decline in natural killer cell cytotoxicity. *Aging Cell* 2012; 11 : 751-9.

55. Krishnaraj R. Senescence and cytokines modulate the NK cell expression. *Mech Ageing Dev* 1997; 96 : 89-101.

56. Pietilä M, Neuvonen M, Borodulin K, Korpela K, Sievänen T, Tyrvainen L. Relationships between exposure to urban green spaces, physical activity and self-rated health. *J Outdoor Recr Tou* 2015; 10(1):44-54.

57. Hazeldine J, Lord JM. The impact of ageing on natural killer cell function and potential consequences for health in older adults. *Ageing Res Rev* 2013; 12(4): 1069-78.

58. Chiu BC, Martin BE, Stolberg VR, Chensue SW. The host environment is responsible for aging related functional NK cell deficiency. *J Immunol* 2013; 191(9):4688-98.

59. Fiebich BL, Batista CRA, Saliba SW, Yousif NM, de Oliveira ACP. Role of Microglia TLRs in Neurodegeneration. *Front Cell Neurosci* 2018; 12 : 329.

60. Montoya-Ortiz G. Immunosenescence, aging and systemic lupus erythematosus. *Autoimmune Dis* 2013; 267078.

61. Kong KF, Delroux K, Wang X, et al. Dysregulation of TLR3 impairs the innate immune response to West Nile virus in the elderly. *J Virol* 2008; 82(15):7613-23.

62. Boche D, Perry VH, Nicoll JAR. Review: activation patterns of microglia and their identification in the human brain. *Neuropathol Appl Neurobiol* 2013; 39 : 3-18.

63. Calvo-Rodríguez M, de la Fuente C, García-Durillo M, García-Rodríguez C, Villalobos C, Núñez L. Aging and amyloid β oligomers enhance TLR4 expression, LPS-induced Ca^{2+} responses and neuron cell death in cultured rat hippocampal neurons. *J Neuroinflammation* 2017; 14(1):24.

64. Shaw AC, Joshi S, Greenwood H, Panda A, Lord MJ. Aging of the innate immune system. *Curr Opin Immunol* 2010; 22(4): 507-13.

65. Stervbo Bozzetti C, Baron U, Jürchott K, et al. Effects of aging on human leukocytes (part II): immunophenotyping of adaptive immune B and T cell subsets. *Age* 2015; 37(5):93.

66. Silverstein AM. Autoimmunity: a history of the early struggle for recognition. In : Mackay IR, Rose NR, (eds). *The Autoimmune Diseases (chapter 2)*. Academic Press, 2014 : Academic Press; 2014. p. .

67. Faragher R, Frasca D, Remarque E, et al. Better immunity in later life: a position paper. *Age* 2014; 36(3):9619.

68. Duggal AN, Pollock RD, Lazarus NR, Harridge S, Lord JM. Major features of immunosenescence, including reduced thymic output, are ameliorated by high levels of physical activity in adulthood. *Aging Cell* 2018; 7(2):e12750.

69. Griffith AV, Venables T, Shi J, et al. Metabolic damage and premature thymus aging caused by stromal catalase deficiency. *Cell reports* 2015; 12(7):1071-9.

70. Palmer DB. The effect of age on thymic function. *Frontiers in immunology* 2013; 4 : 316.

71. Strioga M, Pasukoniene V, Characiejus D. CD8 + CD28- and CD8 + CD57 + T cells and their role in health and disease. *Immunology* 2011; 134 : 17-32.

72. Kollar S, Berta L, Vasarhelyi ZE, et al. Impact of aging on calcium influx and potassium channel characteristics of T lymphocytes. *Oncotarget* 2015; 6(15):13750-6.

73. Deruy E, Nassour J, Martin N, et al. Level of macroautophagy drives senescent keratinocytes into cell death or neoplastic evasion. *Cell Death Dis* 2014; 5(1):e1577.

74. Jing H, Lee S. NF-kappaB in cellular senescence and cancer treatment. *Mol Cells* 2014; 37(3):189-95.

75. Ferrando-Martinez S, Ruiz-Mateos E, Hernandez A, et al. Age-related deregulation of naive T cell homeostasis in elderly humans. *Age* 2011; 33(2):197-207.

76. Li G, Yu M, Lee WW, et al. Decline in miR-181a expression with age impairs T cell receptor sensitivity by increasing DUSP6 activity. *Nat Med* 2012; 18(10):1518-24.

77. Mondal AM, Horikawa I, Pine SR, et al. p53 isoforms regulate aging- and tumor-associated replicative senescence in T lymphocytes. *J Clin Immunol* 2013; 123(12):5247-57.

78. Henson SM, Macaulay R, Riddell NE, Nunn CJ, Akbar AN. Blockade of PD-1 or p38 MAP kinase signaling enhances senescent human CD8(+) T-cell proliferation by distinct pathways. *Eur J Immunol* 2015; 45 : 1441-51.

79. Lanna A, Gomes DC, Muller-Durovic B, et al. A sestrin-dependent Erk-Jnk-p38 MAPK activation complex inhibits immunity during aging. *Nat Immunol* 2017; 18 : 354-63.

80. Lanna A, Henson SM, Escors D, Akbar AN. The kinase p38 activated by the metabolic regulator AMPK and scaffold TAB 1 drives the senescence of human T cells. *Nat Immunol* 2014; 15 : 965-72.

81. Comans-Bitter WM, de Groot R, van den Beemd R, et al. Immunophenotyping of blood lymphocytes in childhood. Reference values for lymphocyte subpopulations. *J Pediatr* 1997; 3 : 388-93.

82. Booth NJ, Akbar AN, Vukmanovic-Stejic M. Regulation of adaptive immunity in the elderly. In : Thiel A, ed. *Immunosenescence*. Basel: Springer Basel.

83. Maijo M, Clements SJ, Ivory K, Nicoletti C, Carding SR. Nutrition, diet and immunosenescence. *Mech Ageing Dev* 2014; 136-137 : 116-28.

84. Montecino-Rodriguez E, Berent-Maoz B, Dorshkind K. Causes, consequences and reversal of immune system aging. *J Clin Investig* 2013; 123(3):958-65.

85. Chou JP, Effros RB. T cell replicative senescence in human aging. *Curr Pharm Des* 2013; 19(9):1680-98.

86. Ovadya Y, Tomer Landsberger T, Hanna Leins H, et al. Impaired immune surveillance accelerates accumulation of senescent cells and aging. *Nat Commun* 2018; 9 : 5435.

87. Jagger A, Shimojima Y, Goronzy JJ, Weyand CM. Regulatory T cells and the immune aging process: a mini-review. *Gerontology* 2014; 60(2):130-7.

88. Hou PF, Zhu LJ, Chen XY, Qiu ZQ. Age-related changes in CD4 + CD25 + FOXP3+ regulatory T cells and their relationship with lung cancer. *PLoS One* 2017; 12 : 173.

89. Fessler J, Ficjan A, Duftner C, Dejaco C. The impact of aging on regulatory T-cells. *Front Immunol* 2013; 4 : 231.

90. Deng Y, Jing Y, Campbell AE, Gravenstein S. Age-related impaired type 1 T cell responses to influenza: reduced activation ex vivo, decreased expansion in CTL culture in vitro and blunted response to influenza vaccination in vivo in the elderly. *J Immunol* 2004; 172 : 3437-46.

91. Schmitt V, Rink L, Uciechowski P. The Th17/Treg balance is disturbed during aging. *Exp Gerontol* 2013; 48 : 1379-86.

92. Toda H, Araki K, Moritomo T, et al. *Molecular biology of the cell*; 4th edition., New York and London: Garland Science.

93. Fletcher JM, Vukmanovic-Stejic M, Dunne PJ, et al. Cytomegalovirus-specific CD4⁺ T cells in healthy carriers are continuously driven to replicative exhaustion. *J Immunol* 2005; 175(12):8218-25.

94. Alama I, Paweleca G. Aging, nutrition and immunity- their relationship and interaction. *Nutr Aging* 2012; 1 : 151-65.

95. Signer Robert AJ, Morrison Sean J. Mechanisms that regulate stem cell aging and life span. *Cell Stem Cell* 2013; 12 : 152-65.

96. Cancro MP, Hao Y, Scholz JL, *et al.* B cells and aging: molecules and mechanisms. *Trends Immunol* 2009; 30(7): 313-8.

97. Colonna-Romano G, Aquino A, Bulati M, *et al.* Memory B cell subpopulations in the aged. *Rejuvenation Res* 2006; 9 : 149-52.

98. Frasca D, Blomberg BB. Effects of aging on B cell function. *Curr Opin Immunol* 2009; 21 : 425-30.

99. Weksler ME. Changes in the B-cell repertoire with age. *Vaccine* 2000; 18(16):1624-8.

100. Duggal NA, Upton J, Phillips AC, Sapey E, Lord JM. An age-related numerical and functional deficit in CD19(+) CD24(hi) CD38(hi) B cells is associated with an increase in systemic autoimmunity. *Aging Cell* 2012; 12 : 873-81.

101. Gibson KL, Wu Y-C, Barnett Y, *et al.* B cell diversity decreases in old age and is correlated with poor health status". *Aging Cell* 2009; 8 : 18-25.

102. Pritz T, Lair J, Ban M, *et al.* Plasma cell numbers decrease in bone marrow of old patients. *Eur J Immunol* 2015; 45(3):738-46.

103. Weinberger B. Vaccines for the elderly: current use and future challenges. *Immun Aging* 2018; 15 : 13.

104. Romanyukha AA, Yashin AI. Age related changes in population of peripheral T cells: towards a model of immunosenescence. *Mech Ageing Dev* 2003; 124 : 433-44.

105. Duggal NA, Beswetherick A, Upton J, Hampson P, Phillips AC, Lord JM. Depressive symptoms in hip fracture patients are associated with reduced monocyte superoxide production. *Exp Gerontol* 2014; 54 : 27-34.

106. Garbe K, Bratke K, Wagner S, Virchow JC, Lommatsch M. Plasmacytoid dendritic cells and their Toll-like receptor 9 expression selectively decrease with age. *Hum Immunol* 2012; 73 : 493-7.

107. Agrawal S, Ganguly S, Tran A, Sundaram P, Agrawal A. Retinoic acid treated human dendritic cells induce T regulatory cells via the expression of CD141 and GARP which is impaired with age. *Ageing* 2016; 8 : 1223-35.

108. Minoda Y, Virshup I, Rojas IL, *et al.* Human CD141⁺ dendritic cell and CD1c⁺ dendritic cell undergo concordant early genetic programming after activation in humanized mice *in vivo* front. *Immunol* 2017; 8 : 1419.

109. Shehata HM, Hoebe K, Chouquet CA. The aged nonhematopoietic environment impairs natural killer cell maturation and function. *Aging Cell* 2015; 14(2):191-9.

110. Le Garff-Tavernier M, Bézat V, Decocq J, *et al.* Human NK cells display major phenotypic and functional changes over the life span. *Aging Cell* 2010; 9 : 527-35.

111. van Duin D, Mohanty S, Thomas V, *et al.* Age-associated defect in human TLR-1/2 function. *J Immunol* 2007; 178 : 970-5.

112. Shaw AC, Panda A, Joshi SR, Qian F, Allore HG, Montgomery RR. Dysregulation of human Toll-like receptor function in aging. *Ageing Res Rev* 2011; 10(3):346-53.

113. Sapin MR, Etingen LE. The human immune system. *Medicine* 1996.

114. Consolini R, Legitimo A, Calleri A. Distribution of age related thymulin titres in normal subjects through the course of life. *Clin Exp Immunol* 2000; 121 : 444-7.

115. Effros RB. Replicative senescence in the immune system: impact of the Hayflick limit on T-cell function in the elderly. *Am J Hum Genet* 1998; 62 : 1003-7.

116. Globerson A, Effros RB. Aging of lymphocytes and lymphocytes in the aged. *Immunol Today* 2000; 21 : 515-21.

117. Effros RB, Dagarag M, Spaulding C, Man J. The role of CD8+ T-cell replicative senescence in human aging. *Immunol Rev* 2005; 205(1):147-57.

118. Dorshkind K, Montecino-Rodriguez E, Signer RA. The ageing immune system: is it ever too old to become young again? *Nat Rev Immunol* 2009; 9 : 57-62.

119. Buffa S, Pellicano M, Bulati M, *et al.* A novel B cell population revealed by a CD38/CD24 gating strategy: CD38(-)CD24(-) B cells in centenarian offspring and elderly people. *Age* 2013; 35 (5):2009-24.

120. Jacobs TL, Epel ES, Lin J, *et al.* Intensive meditation training, immune cell telomerase activity and psychological mediators. *Psychoneurol Endocr* 2011; 36(5):664-81.

121. Morgan N, Irwin MR, Chung M, Wang C. The effects of mind-body therapies on the immune system: meta-analysis. *PLoS One* 2014; 9(7):e100903.

122. Fan Y, Tang YY, Ma Y, Posner MI. Mucosal immunity modulated by integrative meditation in a dose-dependent fashion. *J Altern Complement Med* 2010; 16(2):151-5.

123. Falkenberg RI, Eising C, Peters ML. Yoga and immune system functioning: a systematic review of randomized controlled trials. *J Behav Med* 2018; 41(4):467-82.

124. Infante JR, Peran F, Rayo JI, *et al.* Levels of immune cells in transcendental meditation practitioners. *Int J Yoga* 2014; 7 (2):147-51.

125. Carroll JE, Cole SW, Seeman TE, *et al.* Partial sleep deprivation activates the DNA damage response (DDR) and the senescence-associated secretory phenotype (SASP) in aged adult humans. *Brain Behav Immun* 2016; 51 : 223-9.

126. Hampson P, Rossi A, Arora T, Lord JM, Taheri S. Sleep and immunity in older age. In : Bosch JA, Phillips AC, Lord JM, (eds). *Immunosenescence: psychosocial and behavioral determinants*. New York, NY: Springer New York.

127. Besedovsky L, Lange T, Born J. Sleep and immune function. *Pflugers Arch* 2012; 463(1):121-37.

128. Li T, Ping Wang P, Wang SC, Wang YF. Approaches mediating oxytocin regulation of the immune system. *Front Immunol* 2016; 7 : 693.

129. Ponsonby A-L, Lucas RM, Van der Mei AF. UVR, vitamin D and three autoimmune diseases–multiple sclerosis, type 1 diabetes, rheumatoid arthritis. *Photochem Photobiol* 2005; 81 : 1267-75.

130. Grant WB. Hypothesis–ultraviolet-B irradiance and vitamin D reduce the risk of viral infections and thus their sequelae, including autoimmune diseases and some cancers. *Photochem Photobiol* 2008; 84 : 356-65.

131. Phan TX, Jaruga B, Pingle SC, Bandyopadhyay BC, Ahern GP. Intrinsic photosensitivity enhances motility of T lymphocytes. *Sci Rep* 2016; 6 : 39479.

132. Brenner IK, Castellani JW, Gabaree C, *et al.* Immune changes in humans during cold exposure: effects of prior heating and exercise. *J Appl Physiol* 1999; 87 : 699-710.

133. Shevchuk NA, Radoja S. Possible stimulation of anti-tumor immunity using repeated cold stress: a hypothesis. *Infect Agent Cancer* 2007; 2 : 20.

134. Burton D, Stolzing A. Cellular senescence: immunosurveillance and future immunotherapy. *Ageing Res Rev* 2018; 43 : 17-25.

135. Choi IY, Lee C, Longo VD. Nutrition and fasting mimicking diets in the prevention and treatment of autoimmune diseases and immunosenescence. *Mol Cell Endocrinol* 2017; 455 : 4-12.

136. Cheng CW, Adams GB, Perin L, *et al.* Prolonged fasting reduces IGF-1/PKA to promote hematopoietic stem cell-based

regeneration and reverse immunosuppression. *Cell Stem Cell* 2014; 14(6):810-23.

137. Rangan P, Choi I, Wei M, et al. Fasting-mimicking diet modulates microbiota and promotes intestinal regeneration to reduce inflammatory bowel disease pathology. *Cell Rep* 2019; 26(10):2704-2719.e6.
138. Lee C, Longo V. Dietary restriction with and without caloric restriction for healthy aging. *Mol Cell Endocrinol* 2017; 5 (455):4-12.
139. Steven S, Taylor R. Restoring normoglycaemia by use of a very low calorie diet in long- and short-duration Type 2 diabetes. *Diabet Med* 2015; 32(9):1149-55.
140. Yang H, Youm YH, Dixit VD. Inhibition of thymic adipogenesis by caloric restriction is coupled with reduction in age-related thymic involution. *J Immunol* 2009; 183(5):3040-52.
141. Messaoudi I, Warner J, Fischer M, et al. Delay of T cell senescence by caloric restriction in aged long-lived nonhuman primates. *Proc Natl Acad Sci U S A* 2006; 103(51):19448-53.
142. Spaulding CC, Walford RL, Effros RB. Calorie restriction inhibits the age-related dysregulation of the cytokines TNF- and IL-6 in C3B10RF1 mice. *Mech Ageing Dev* 1997; 93 : 87-94.
143. Jolly CA. Dietary restriction and immune function. *J Nutr* 2004; 134 : 1853-6.
144. Tang D, Tao S, Chen Z, Koliesnik LO, Calmes PG, Hoerr V. Dietary restriction improves repopulation but impairs lymphoid differentiation capacity of hematopoietic stem cells in early aging. *J Exp Med* 2016; 213 : 535-53.
145. Qiu X, Brown K, Hirschey MD, Verdin E, Chen D. Calorie restriction reduces oxidative stress by SIRT3-mediated SOD2 activation. *Cell Metab* 2010; 12(6):662-7.
146. Oldways. *What is the Mediterranean Diet?* 2016. <https://oldwayspt.org/traditional-diets/mediterranean-diet/what-mediterranean-diet>
147. Casas R, Sacanella E, Urpí-Sardà M, et al. The effects of the Mediterranean diet on biomarkers of vascular wall inflammation and plaque vulnerability in subjects with high risk for cardiovascular disease. A randomized trial. *PLoS One* 2014; 9 (6):e100084.
148. Clements SJ, Maijo M, Ivory K, Nicoletti C, Carding SR. Age-associated decline in dendritic cell function and the impact of mediterranean diet intervention in elderly subjects. *Front Nutr* 2017; 4 : 65.
149. Fantuzzi G. Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* 2005; 115(5):911-9.
150. Baylis D, Bartlett DB, Patel HP, Roberts HC. Understanding how we age: insights into inflammmaging. *Longevity Healthspan* 2013; 2(1):8.
151. Bartlett BD, Huffman MK. *Lifestyle interventions to improve*. Switzerland: Springer International Publishing.
152. Gleeson M, Bishop NC, Stensel DJ, et al. The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. *Nat Rev Immunol* 2011; 11 : 607-15.
153. Bartlett D, Fox O, McNulty CL, et al. Habitual physical activity is associated with the maintenance of neutrophil migratory dynamics in healthy older adults.. *Brain Behav Immun* 2016; 56 : 12-20.
154. Zheng Q, Cui G, Chen J, et al. Regular exercise enhances the immune response against microbial antigens through up-regulation of toll-like receptor signaling pathways. *Cell Physiol Biochem* 2015; 37(2):735-46.
155. Takahashi M, Miyashita M, Kawanishi N, et al. Low-volume exercise training attenuates oxidative stress and neutrophils activation in older adults. *Eur J Appl Physiol* 2013; 113(5): 1117-26.
156. Turner JE, Brum PC. Does regular exercise counter T cell immunosenescence reducing the risk of developing cancer and promoting successful treatment of malignancies? *Oxid Med Cell Longev* 2017; 2017 : 4234765.
157. van der Geest KSM, Wang Q, Eijsvogel TMH, et al. Changes in peripheral immune cell numbers and functions in octogenarian walkers—an acute exercise study. *Immun Ageing* 2017; 14 : 5.
158. Duggal NA. Reversing the immune ageing clock: lifestyle modifications and pharmacological interventions. *Biogerontology* 2018; 19(6):481-96.
159. Maggini S, Wintergerst ES, Beveridge S, Hornig DH. Selected vitamins and trace elements support immune function by strengthening epithelial barriers and cellular and humoral immune responses. *Br J Nutr* 2007; 98(Suppl 1):S29.
160. Zhang LJ, Chen SX, Guerrero-Juarez CF, et al. Age-related loss of innate immune antimicrobial function of dermal fat is mediated by transforming growth factor beta. *Immunity* 2019; 50(1):121-136.e5.
161. Shao Y, He T, Fisher GJ, Voorhees JJ, Quan T. Molecular basis of retinol anti-ageing properties in naturally aged human skin *in vivo*. *Int J Cosmet Sci* 2017; 39(1):56-65.
162. Raverdeau M, Mills KH. Modulation of T cell and innate immune responses by retinoic acid. *J Immunol* 2014; 192 (7):2953-8.
163. Erdman J, MacDonald I, Zeisel S. *Present knowledge in nutrition*, 10th ed, Wiley-Blackwell, 2018.
164. Porter K, Hoey L, Hughes CF, Ward M, McNulty H. Causes, consequences and public health implications of low B-vitamin status in ageing. *Nutrients* 2016; 8(11):725.
165. Laird EJ, O'Halloran AM, Carey D, O'Connor D, Kenny RA, Molloy AM. Voluntary fortification is ineffective to maintain the vitamin B12 and folate status of older Irish adults: evidence from the Irish Longitudinal Study on Ageing (TILDA). *Br J Nutr* 2018; 120(1):111-20.
166. Green R. Vitamin B₁₂ deficiency from the perspective of a practicing hematologist. *Blood* 2017; 129 : 2603-11.
167. Michels AJ, Leonard SW, Uesugi SL, Bobe G, Frei B, Traber MG. Daily consumption of oregon hazelnuts affects α-tocopherol status in healthy older adults: a pre-post intervention study. *J Nutr* 2018; 148(12):1924.
168. Moriguchi S, Muraga M. Vitamin E and immunity. *Vitam Horm* 2000; 59 : 305-36.
169. Wu D, Meydani SN. Age-associated changes in immune function: impact of vitamin E intervention and the underlying mechanisms. *Endocr Metab Immune Disord Drug Targets* 2014; 14(4):283-9.
170. Adolfsson O, Huber BT, Meydani SN. Vitamin E-enhanced IL-2 production in old mice: naive but not memory T cells show increased cell division cycling and IL-2-producing capacity. *J Immunol* 2001; 167 : 3809-17.
171. Crétel E, Veen I, Pierres A, Bongrand P, Gavazzi G. Immunosénescence et infections, mythe ou réalité? *Med Mal Infect* 2010; 40(6):307-18.
172. Mizwicki MT, Menegaz D, Zhang J, et al. Genomic and nongenomic signaling induced by 1α,25(OH)2-vitamin D3 promotes the recovery of amyloid-β phagocytosis by Alzheimer's disease macrophages. *J Alzheimers Dis* 2012; 29(1): 51-62.
173. Schmitt EB, Nahas-Neto J, Bueloni-Dias F, Poloni PF, Orsatti CL, Nahas EAP. Vitamin D deficiency is associated with

metabolic syndrome in postmenopausal women. *Maturitas* 2018; 107 : 97.

174. De Carvalho Goncalves CMR, Ribeiro SML. Aging, low grade systemic inflammation and vitamin D: a mini review. *European journal of clinical nutrition* 2017; 71 : 434-40.

175. Uwitonze AM, Razzaque MS. Role of magnesium in vitamin D activation and function. *J Am Osteopath Assoc* 2018; 118 (3):181.

176. Vanherwegen AS, Gysemans C, Mathieu C. Regulation of immune function by vitamin D and its use in diseases of immunity. *Endocrinol Metab Clin* 2017; 46 : 1061-94.

177. Hemila H, Chalker E. Vitamin C for preventing and treating the common cold. *Cochrane Database Syst Rev* 2013; 1 : CD000980.

178. Sorice A, Guerriero E, Capone F, Colonna G, Castello G, Costantini S. Ascorbic acid: its role in immune system and chronic inflammation diseases. *Mini Rev Med Chem* 2014; 14 (5):444-52.

179. Prasad AS. Zinc: role in immunity, oxidative stress and chronic inflammation. *Curr Opin Clin Nutr Metab Care* 2009; 12 : 646-52.

180. Prasad AS, Beck FW, Bao B, et al. Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress. *Am J Clin Nutr* 2007; 85(3):837-44.

181. Bao B, Prasad AS, Beck FW, et al. Zinc decreases C-reactive protein, lipid peroxidation, and inflammatory cytokines in elderly subjects: a potential implication of zinc as an athero -protective agent. *Am J Clin Nutr* 2010; 91(6): 1634-41.

182. Meydani SN, Barnett JB, Dallal GE, et al. Serum zinc and pneumonia in nursing home elderly. *Am J Clin Nutr* 2007; 86 (4):1167-73.

183. Mocchegiani E, Muzzioli M, Giacconi R, et al. Metallothioneins/PARP-1/IL-6 interplay on natural killer cell activity in elderly: parallelism with nonagenarians and old infected humans. Effect of zinc supply. *Mech Ageing Dev* 2003; 124 : 459-68.

184. Uciechowski P, Kahmann L, Plümäkers B, et al. TH1 and TH2 cell polarization increases with aging and is modulated by zinc supplementation. *Exp Gerontol* 2008; 43 : 493-8.

185. Boukaïba N, Flament C, Acher S, et al. A physiological amount of zinc supplementation: effects on nutritional, lipid, and thymic status in an elderly population. *Am J Clin Nutr* 1993; 57 : 566-72.

186. Duchateau J, Delepesse G, Vrijens R, Collet H. Beneficial effects of oral zinc supplementation on the immune response of old people. *Am J Med* 1981; 70 : 1001-4.

187. Mocchegiani E, Costarelli L, Giacconi R, Piacenza F, Basso A, Malavolta M. Micronutrient (Zn, Cu, Fe)- gene interactions in ageing and inflammatory age-related diseases: implications for treatments. *Ageing Res Rev* 2012; 11(2):297-319.

188. Dao MC, Meydani SN. Iron biology, immunology, aging and obesity: four fields connected by the small peptide hormone hepcidin. *Adv Nutr* 2013; 4(6):602-17.

189. Fairweather-Tait SJ, Wawer AA, Gillings R, Jennings A, Myint PK. Iron status in the elderly. *Mech Ageing Dev* 2014; 136-137 : 22-8.

190. Strindhall J, Ernerudh J, Mörner A, et al. Humoral response to influenza vaccination in relation to pre-vaccination antibody titres, vaccination history, cytomegalovirus serostatus and CD4/CD8 ratio. *Infect Dis* 2016; 48(6):436-42.

191. Murasko DM, Bernstein ED, Gardner EM, et al. Role of humoral and cell-mediated immunity in protection from influenza disease after immunization of healthy elderly. *Exp Gerontol* 2002; 37 : 427-39.

192. Siegrist CA, Aspinall R. B-cell responses to vaccination at the extremes of age. *Nat Rev Immunol* 2009; 9(3):185.

193. Ciabattini A, Nardini C, Santoro F, Garagnani P, Franceschig C, Medaglini D. Vaccination in the elderly: the challenge of immune changes with aging. *Semin Immunol* 2018; 40 : 83-94.

194. Osterholm MT, Kelley NS, Sommer A, Belongia EA. Efficacy and effectiveness of influenza vaccines: a systematic review and meta-analysis. *Lancet Infect Dis* 2012; 12(1): 1236-44.

195. Van Deursen JM. The role of senescent cells in ageing. *Nature* 2014; 509(7501):439-46.

196. Maruyama T, Taguchi O, Niederman MS, et al. Efficacy of 23-valent pneumococcal vaccine in preventing pneumonia and improving survival in nursing home residents: double blind, randomised and placebo controlled trial. *BMJ* 2010; 340 : c1004.

197. Vila-Corcoles A, Salsench E, Rodriguez-Blanco T, et al. Clinical effectiveness of 23-valent pneumococcal polysaccharide vaccine against pneumonia in middle-aged and older adults: a matched case-control study. *Vaccine* 2009; 27 : 1504-10.

198. Izurieta HS, Werneck M, Kelman J, et al. Effectiveness and duration of protection provided by the live-attenuated herpes zoster vaccine in the medicare population ages 65 years and older. *Clin Infect Dis* 2017; 64 : 785-93.

199. Levin MJ, Kroehl ME, Johnson MJ, et al. Th1 memory differentiates recombinant from live herpes zoster vaccines. *J Clin Invest* 2018; 128(10):4429-40.

200. Levin MJ, Oxman MN, Zhang JH, et al. Varicella-zoster virus-specific immune responses in elderly recipients of a herpes zoster vaccine. *J Infect Dis* 2008; 197 : 825-35.

201. Lal H, Cunningham AL, Godeaux O, et al. Efficacy of an adjuvanted herpes zoster subunit vaccine in older adults. *N Engl J Med* 2015; 372 : 2087-96.

202. Coccia M, Collignon C, Hervé C, et al. Cellular and molecular synergy in AS01-adjuvanted vaccines results in an early IFN γ response promoting vaccine immunogenicity. *NPJ Vaccin* 2017; 2 : 25.

203. Didierlaurent AM, Laupèze B, Di Pasquale A, et al. Adjuvant system AS01: helping to overcome the challenges of modern vaccines. *Expert Rev Vaccines* 2017; 16 : 55-63.

204. Weinberger B, Grubeck-Loebenstein B. Vaccination in the Elderly. In : Thiel A, ed. *Immunosenescence*. Basel: Springer Basel.

205. Cunningham AL, Lal H, Kovac M, et al. Efficacy of the Herpes Zoster subunit vaccine in adults 70 years of age or older. *N Engl J Med* 2016; 375 : 1019-32.

206. Silva SL, Albuquerque AS, Matoso P, et al. IL-7-induced proliferation of human naive CD4 T-cells relies on continued thymic activity. *Front Immunol* 2017; 8 : 20.

207. Pellegrini M, Calzascia T, Toe JG, et al. IL-7 engages multiple mechanisms to overcome chronic viral infection and limit organ pathology. *Cell* 2011; 144 : 601-13.

208. Lang PO, Govind S, Aspinall R. Reversing T cell immunosenescence: why, who, and how. *Age* 2013; 35(3):609-20.

209. Ma LJ, Acero LF, Zal T, Schluns KS. Trans-presentation of IL-15 by intestinal epithelial cells drives development of CD8 $\alpha\alpha$ IELs. *J Immunol* 2009; 183(2):1044-54.

210. Chiu BC, Martin BE, Stolberg VR, Chensue SW. The host environment is responsible for aging related functional NK cell deficiency. *J Immunol* 2013; 191(9):4688-98.

211. Gangemi S, Basile G, Monti D, et al. Age-related modifications in circulating IL-15 levels in humans. *Mediators Inflamm* 2005; 4 : 245-7.

212. Flores I, Cayuela ML, Blasco MA. Effects of telomerase and telomere length on epidermal stem cell behavior. *Science* 2005; 309(5738):1253-6.

213. Mitchell WA, Pink RC, Lapenna A, Aspinall R. Immunosenescence and the 3Rs: Restoration, Replacement and Reprogramming. In : Thiel A, ed. *Immunosenescence*. Basel: Springer Basel.

214. Berent-Maoz B, Montecino-Rodriguez E, Signer RAJ, Kenneth Dorshkind K. Fibroblast growth factor-7 partially reverses murine thymocyte progenitor aging by repression of *Ink4a*. *Blood* 2012; 119(24):5715-21.

215. Buford TW, Willoughby DS. Impact of DHEA(S) and cortisol on immune function in aging: a brief review. *Appl Physiol Nutr Metab* 2008; 33(3):429-33.

216. Kroll J. Dehydroepiandrosterone, molecular chaperones and the epigenetics of primate longevity. *Rejuvenation Res* 2015; 18 (4):341-6.

217. Prall SP, Muehlenbein MP. Dehydroepiandrosterone and multiple measures of functional immunity in young adults. *Am J Hum Biol* 2015; 27(6):877-80.

218. Khorram O, Vu L, Yen SS. Activation of immune function by dehydroepiandrosterone (DHEA) in age-advanced men. *J Gerontol A Biol Sci Med Sci* 1997; 52(1):M1-7.

219. Rondanelli M, Giacosa A, Faliva MA, Perna S, Allieri F, Castellazzi AM. Review on microbiota and effectiveness of probiotics use in older. *World J Clin Cases* 2015; 3(2): 156-62.

220. Toward RE, Walton GE, Gibson GR. Immunosenescence and the gut microbiota: the role of probiotics and prebiotics. *Nutr Aging* 2012; 1(3-4):167-80.

221. Toward R, Montandon S, Walton G, Gibson GR. Effect of prebiotics on the human gut microbiota of elderly persons. *Gut Microbes* 2012; 3(1):57-60.

222. Fu YR, Yi ZJ, Pei JL, Guan S. Effects of *Bifidobacterium bifidum* on adaptive immune senescence in aging mice. *Microbiol Immunol* 2010; 54 : 578-83.

223. Meydani SN, Ha WK. Immunologic effects of yogurt. *Am J Clin Nutr* 2000; 71(4):861-72.

224. Nagai T, Makino S, Ikegami S, Itoh H, Yamada H. Effects of oral administration of yogurt fermented with *Lactobacillus delbrueckii* ssp. *bulgaricus* OLL1073R-1 and its exopolysaccharides against influenza virus infection in mice. *Int Immunopharmacol* 2011; 11 : 2246-50.

225. Makino S, Ikegami S, Kano H, et al. Immunomodulatory effects of polysaccharides produced by *Lactobacillus delbrueckii* ssp. *bulgaricus* OLL1073R-1. *J Dairy Sci* 2006; 89(2):873-81.

226. Dalton N, Goldman GH, Gern RM. Beta-glucans: medicinal activities, characterization, biosynthesis and new horizons. *Appl Microbiol Biotechnol* 2015; 99(19):7893-906.

227. Karumuthil-Meletih S, Gudi R, Johnson BM, Perez N, Vasu C. Fungal beta-glucan, a Dectin-1 ligand, promotes protection from type 1 diabetes by inducing regulatory innate immune response. *Journal Immunol* 2014; 193(7): 3308-21.

228. Jin X, Ruiz Beguerie J, Sze D M, Chan G C. *Ganoderma lucidum* (Reishi mushroom) for cancer treatment. *Cochrane Database Syst Rev* 2012; 6 : CD007731.

229. Lee SS, Wei YH, Chen CF, Wang SY, Chen KY. Antitumor effects of *Ganoderma lucidum*. *J Chin Med* 1995; 6 : 1-12.

230. Ji Z, Tang Q, Zhang J, Yang Y, Jia W, Pan Y. Immunomodulation of RAW264.7 macrophages by GLIS, a proteopolysaccharide from *Ganoderma lucidum*. *J Ethnopharmacol* 2007; 112 : 445-50.

231. Mao T, Van De Water J, Keen CL, Stern JS, Hackman R, Gershwin ME. Two mushrooms, *Grifola frondosa* and *Ganoderma lucidum*, can stimulate cytokine gene expression and proliferation in human T lymphocytes. *Int J Med Mushrooms* 2007; 142 : 13-22.

232. Sun LX, Lin ZB, Li XJ, et al. Promoting effects of *Ganoderma lucidum* polysaccharides on B16F10 cells to activate lymphocytes. *Basic Clin Pharmacol Toxicol* 2011; 108 : 149-54.

233. Bao XF, Wang XS, Dong Q, Fang JN, Li XY. Structural features of immunologically active polysaccharides from *Ganoderma lucidum*. *Phytochem* 2002; 59 : 175-81.

234. Zhang K, Ma X, He W, et al. Extracts of *Cistanche deserticola* can antagonize immunosenescence and extend life span in senescence-accelerated mouse prone 8 (SAM-P8) Mice. Evidence-based complementary and alternative medicine. *eCAM* 2014; 601383.

235. Yonei Y, Kitano T, Ogura M, et al. Effects of health food containing *Cistanche deserticola* extract on qol and safety in elderly: an open pilot study of 12-week oral treatment. *Anti Aging Med* 2011; 8(2):7-14.

236. Zhang L, Shao WF, Yuan LF, Tu PF, Ma ZZ. Decreasing pro-inflammatory cytokine and reversing the immunosenescence with extracts of Pu-erh tea in senescence accelerated mouse (SAM). *Food chemistry* 2012; 135(4):2222-8.

237. Chu SL, Fu H, Yang JX, et al. A randomized double-blind placebo-controlled study of Pu'er tea extract on the regulation of metabolic syndrome. *Chin J Integr Med* 2011; 17(7):492-8.

238. Ebrahimi M, Mohammad Hassan Z, Mostafaie A, Zare Mehrjardi N, Ghazanfari T. Purified protein fraction of garlic extract modulates cellular immune response against breast transplanted tumors in BALB/c mice model. *Cell J Spring* 2013; 15(1):65-75.

239. Ried K. Garlic Lowers blood pressure in hypertensive individuals, regulates serum cholesterol, and stimulates immunity: an updated meta-analysis and review. *J Nutr* 2016; 146(2):389s-96s.

240. Percival SS. Aged garlic extract modifies human immunity. *J Nutr* 2016; 146(2):433s-6s.

241. Zhu Y, Tchkonia T, Pirtskhalava T, et al. The Achilles' heel of senescent cells: from transcription to senolytic drugs. *Aging Cell* 2015; 14 : 644-58.

242. Cámara M, Maríade, Sánchez-Mata C, et al. Lycopene: a review of chemical and biological activity related to beneficial health effects studies in natural products. *Chemistry* 2013; 11 : 383-426.

243. Riso P, Visioli F, Grande S, et al. Effect of a tomato-based drink on markers of inflammation, immunomodulation, and oxidative stress. *J Agric Food Chem* 2006; 54(7):2563-6.

244. Briviba K, Kulling SE, Möseneder J, Watzl B, Rechkemmer G, Bub A. Effects of supplementing a low-carotenoid diet with a tomato extract for 2 weeks on endogenous levels of DNA single strand breaks and immune functions in healthy non-smokers and smokers. *Carcinogenesis* 2004; 25 (12):2373-8.

245. van Breda SG, Wilms LC, Gaj S, et al. The exposome concept in a human nutrigenomics study: evaluating the impact of exposure to a complex mixture of phytochemicals using transcriptomics signatures. *Mutagenesis* 2015; 30(6):723-31.

246. Pan P, Huang YW, Oshima K, et al. An immunological perspective for preventing cancer with berries. *J Berry Res* 2018; 8(3):163-75.

247. Deng Y, Wang F, Hughes T, Yu J. FOXOs in cancer immunity: knowns and unknowns. *Semin Cancer Biol* 2018; 50 : 53-64.

248. Carr AC, Maggini S. Vitamin C and immune function. *Nutrients* 2017; 9(11):E1211.

249. Daniel M, Tollefsbol TO. Epigenetic linkage of aging, cancer and nutrition. *J Exp Biol* 2015; 218(1):59-70.

250. Wang J, Fang X, Ge L, et al. Antitumor, antioxidant and anti-inflammatory activities of kaempferol and its corresponding glycosides and the enzymatic preparation of kaempferol. *PLoS One* 2018; 13(5):e0197563.

251. Das SK, Roberts SB, Bhapkar MV, et al. Body-composition changes in the comprehensive assessment of long-term effects of reducing intake of energy (CALERIE)-2 study: a 2-year randomized controlled trial of calorie restriction in nonobese humans. *Am J Clin Nutr* 2017; 105 : 913-27.

252. Ravussin E, Redman LM, Rochon J, et al. A 2-year randomized controlled trial of human caloric restriction: feasibility and effects on predictors of health span and longevity. *J Gerontol A Biol Sci Med Sci* 2015; 70 : 1097-104.

253. Ostan R, Béné MC, Spazzafumo L, et al. Impact of diet and nutraceutical supplementation on inflammation in elderly people. Results from the RISTOMED study, an open-label randomized control trial. *Clin Nutr* 2016; 5 : 812-8.

254. Moro-García MA, Alonso-Arias R, Baltadjieva M, et al. Oral supplementation with *Lactobacillus delbrueckii* subsp. *bulgaricus* 8481 enhances systemic immunity in elderly subjects. *Age* 2013; 35 : 1311-26.

255. Guillemard E, Tondu F, Lacoin F, Schrezenmeir J. Consumption of a fermented dairy product containing the probiotic *Lactobacillus casei* DN-114001 reduces the duration of respiratory infections in the elderly in a randomised controlled trial. *Br J Nutr* 2010; 103 : 58-68.

256. Shelbaya S, Seddik S, Ahmed A, Roshdy N, Abbas M. Assessment of vitamin D status in different samples of an elderly Egyptian population. *Egypt J Obes Diabetes Endocrinol* 2017; 3 : 53-8.

257. El-Sabbagh NM, Shahin EM, Abo El Makarem NH, et al. Study of the C-reactive protein and tumor necrosis factor alpha levels in the elderly before and after resistance exercise training.. *Egypt J Obes Diabet Endocr* 2015; 1 : 7-13.

258. Kohut ML, Lee W, Martin A, et al. The exercise-induced enhancement of influenza immunity is mediated in part by improvements in psychosocial factors in older adults. *Brain Behav Immun* 2005; 19 : 357-66.