

Swainsonine Inhibits Invasion and the EMT Process in Esophageal Carcinoma Cells by Targeting Twist1

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Esophageal cancer is a common gastrointestinal cancer, with a very high mortality rate in patients with metastasis. Swainsonine, a cytotoxic fungal alkaloid, has been shown to inhibit cell growth in esophageal cancer. In the present study, we explored the effects of swainsonine on cell invasion and metastasis in esophageal cancer cells. Human esophageal carcinoma cells were treated with different doses of swainsonine, and then cell viability, invasion, and apoptosis were measured. The mRNA and protein expressions of Twist1, apoptosis- and EMT-related factors, and PI3K/AKT pathway factors were detected by qRT-PCR and Western blot. Swainsonine had no effect on esophageal cancer cell viability and apoptosis, but it significantly decreased cell invasion in a dose-dependent manner. Swainsonine increased the expression of E-cadherin but decreased the expression of N-cadherin, vimentin, ZEB1, and snail in a dose-dependent manner, thereby inhibiting EMT. Last, we found that swainsonine inhibits cell invasion and EMT in the esophageal carcinoma cells by down-regulation of Twist1 and deactivation of the PI3K/AKT signaling pathway.

Key words: Swainsonine; Esophageal carcinoma; Cell invasion; Metastasis; Epithelial–mesenchymal transition (EMT); Twist1

INTRODUCTION

Esophageal cancer is a common gastrointestinal cancer and is most prevalent in Asia (including China and Central Asia) and in East and South Africa^{1,2}. In 2012, about 455,800 new cases of esophageal cancer were recorded worldwide, about 400,200 people died from esophageal cancer, and more than 80% of these deaths occurred in developing countries^{3,4}. Esophageal cancer is generally three to four times more common among males than females⁵. Esophageal cancer shows no signs or symptoms in the early stage; it is usually diagnosed at an advanced stage, with a 5-year survival rate of 12%^{6–8}. Dysphagia and weight loss are the most common presenting features at advanced stage⁹. Treatment options include surgery, radiotherapy, chemotherapy, targeted therapy, and endoscopic interventions¹⁰. Despite the available treatment options, patients usually die due to relapse or metastasis¹¹.

Swainsonine is a cytotoxic fungal alkaloid, produced by fungal symbionts of plants (endophytes), plant pathogens, or insect pathogens¹². Swainsonine primarily inhibits α -mannosidase II and lysosome α -mannosidase activities¹³. Swainsonine takes part in a wide range of biological functions, including weight loss, decreased libido, altered

behavior, depression, abortion, infertility, birth defects, and death¹². Swainsonine disrupts the endomembrane system of cells by inhibiting α -mannosidase II^{14–16} and is therefore under consideration as a chemotherapeutic treatment option for cancer^{17,18}. Santos et al. showed that the administration of swainsonine in combination with cisplatin increases the sensitivity of Ehrlich ascites carcinoma cells to cisplatin¹⁷. Swainsonine has also been reported to inhibit cell growth of lung cancer cells¹⁵ and esophageal squamous carcinoma cells¹⁹ via a mitochondria-mediated, caspase-dependent apoptotic pathway both in vitro and in vivo. In the present study, we further explored the effects of swainsonine on esophagus cancer, with more focus on cell invasion and metastasis, as the survival rate is much less (4%) in patients with metastasis²⁰.

MATERIALS AND METHODS

Cell Culture and Treatment

Human esophageal carcinoma cell lines EC9706 and 293T (American Type Culture Collection, Manassas, VA, USA) were grown in Roswell Park Memorial Institute (RPMI)-1640 medium, supplemented with 1× antibiotic–antimycotic mixture and 10% fetal bovine serum (FBS; Hyclone Laboratories, Logan, UT, USA). This culture

was then maintained at 37°C in an atmosphere of 5% CO₂–95% air. Transforming growth factor-β1 (TGF-β1; 10 ng/ml) was used to induce epithelial–mesenchymal transition (EMT) in the cells. Swainsonine (Fig. 1) was purchased from Sigma-Aldrich (S6640; St. Louis, MO, USA) and was dissolved in dimethyl sulfoxide (DMSO). Swainsonine was used to treat cells at 10, 20, 50, and 100 µg/ml concentrations in the following experiments.

CCK-8 Assay

Cell proliferation was assessed by cell counting kit-8 (CCK-8; Dojindo Molecular Technologies, Gaithersburg, MD, USA). Cells were seeded in a 96-well plate with 5,000 cells/well. After stimulation, the CCK-8 solution was added to the culture medium, and the cultures were incubated for 1 h at 37°C in humidified 95% air and 5% CO₂. The absorbance was measured at 450 nm using a Microplate Reader (Bio-Rad, Hercules, CA, USA).

Cell Invasion Assay

Cell invasion was measured using 24-well Millicell Hanging Cell Culture Inserts with 8-µm polyethylene terephthalate (PET) membranes (Millipore, Bedford, MA, USA). Briefly, after the cells were treated for indicated conditions, 5.0×10⁴ cells in 200 µl of serum-free Dulbecco's modified Eagle's medium (DMEM) were plated onto BD BioCoat™ Matrigel™ Invasion Chambers (8-µm pore size polycarbonate filters; BD Biosciences, San Jose, CA, USA), while complete medium containing 10% FBS was added to the lower chamber. After processing the invasion chambers for 48 h (37°C, 5% CO₂) in accordance with the manufacturer's protocol, the noninvading cells were removed with a cotton swab; the invading cells were fixed in 100% methanol and then stained with crystal violet solution and counted microscopically. The data were presented as average number of cells attached to the bottom surface from five randomly chosen fields.

Apoptosis Assay

Cell apoptosis was measured using propidium iodide (PI) and fluorescein isothiocyanate (FITC)-conjugated annexin V staining. Briefly, cells were washed in phosphate-buffered saline (PBS). These cells were then

washed twice in PBS and stained in PI/FITC-annexin V in the presence of 50 µg/ml RNase A (Sigma-Aldrich), and then incubated for 1 h at room temperature in the dark. Flow cytometry analysis was done using a FACScan (Beckman Coulter, Fullerton, CA, USA). The data were analyzed using FlowJo software.

Quantitative Reverse Transcription Polymerase Chain Reaction (qRT-PCR)

Total RNA was extracted from cells using TRIzol reagent (Life Technologies Corporation, Carlsbad, CA, USA) according to the manufacturer's instructions. One Step SYBR® PrimeScript® PLUS Kit (TaKaRa Biotechnology, Dalian, P.R. China) was used for qRT-PCR analysis to measure the expression levels of Twist1 in the cells.

Western Blot

The proteins used for Western blotting were extracted using RIPA lysis buffer (Beyotime Biotechnology, Shanghai, P.R. China) supplemented with protease inhibitors (Roche, Guangzhou, P.R. China). The proteins were quantified using BCA™ Protein Assay Kit (Pierce, Appleton, WI, USA). Western blot system was established using a Bio-Rad Bis-Tris Gel system according to the manufacturer's instructions. Primary antibodies were prepared in 5% blocking buffer at a dilution of 1:1,000. Primary antibodies were incubated with the membrane at 4°C overnight, followed by washing and incubation with secondary antibody tagged by horseradish peroxidase for 1 h at room temperature. After rinsing, the polyvinylidene difluoride blot membrane and antibodies were transferred into the Bio-Rad ChemiDoc™ XRS system, and then 200 µl of Immobilon Western Chemiluminescent HRP Substrate (Millipore) was added to cover the membrane surface. The signals were captured, and the intensity of the bands was quantified using Image Lab™ Software (Bio-Rad, Shanghai, P.R. China). Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used as a loading control.

Statistical Analysis

All experiments were repeated three times. The results of multiple experiments are presented as mean±standard deviation. Statistical analyses were performed using SPSS 19.0 statistical software. Values of *p* were calculated using a one-way analysis of variance. A value of *p*<0.05 was considered to indicate a statistically significant result.

RESULTS

Effects of Swainsonine on Cell Viability, Apoptosis, and Invasion

Human esophageal carcinoma cells were treated with different concentrations of swainsonine (10, 20, 50, and

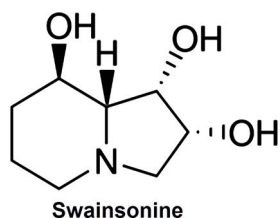


Figure 1. Stereochemical structure of swainsonine.

100 µg/ml); nontreated cells served as control. Cell viability was measured using the CCK-8 assay, apoptosis using flow cytometry, and invasion using Transwell invasion assay. Results showed that swainsonine had no effect on cell viability (Fig. 2A) and apoptosis (Fig. 2B),

but it significantly decreased cell invasion at 20, 50, and 100 µg/ml concentrations compared with that of the control cells ($p < 0.05$) (Fig. 2C). We measured cell viability and apoptosis using Western blot. Results showed that swainsonine had no effect on the expressions of

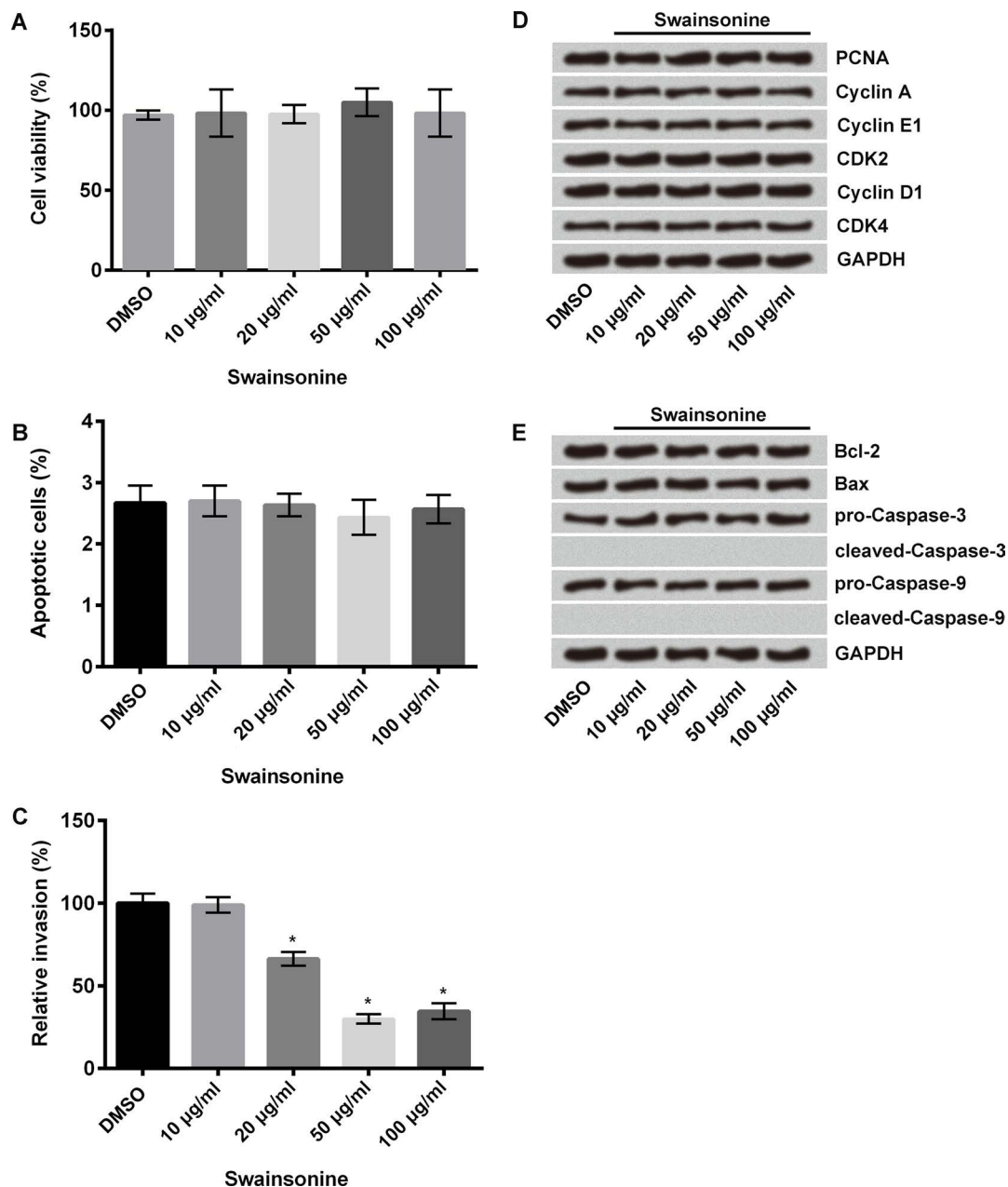


Figure 2. Effects of swainsonine on cell viability, apoptosis, and invasion. Esophageal carcinoma cells were treated with swainsonine at different concentrations (10, 20, 50, and 100 µg/ml); nontreated cells served as control. (A) Cell viability was measured using cell counting kit-8 (CCK-8) assay. (B) Cell apoptosis was measured using flow cytometry. (C) Cell invasion was measured using Transwell invasion assay. (D) Western blot was used to measure the expressions of viability-associated proteins [proliferating cell nuclear antigen (PCNA), cyclin A, cyclin E1, cyclin-dependent kinase 2 (CDK2), cyclin D1, and CDK4]. (E) Western blot was used to measure the expressions of apoptosis-related proteins [B-cell lymphoma 2 (Bcl-2), Bcl-2 X-associated protein (Bax), cleaved caspase 3, and cleaved caspase 9]. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used as a loading control. * $p < 0.05$ compared to control.

viability-associated proteins [proliferating cell nuclear antigen (PCNA), cyclin A, cyclin E1, cyclin-dependent kinase 2 (CDK2), cyclin D1, and CDK4] (Fig. 2D) as well as on the expressions of apoptosis-related proteins [B-cell lymphoma 2 (Bcl-2), Bcl-2 X-associated protein (Bax), cleaved caspase 3, and cleaved caspase 9] (Fig. 2E). These results indicate that swainsonine inhibits invasion of esophageal carcinoma cells but does not affect cell viability and apoptosis.

Effects of Swainsonine on EMT

Because swainsonine inhibited cell invasion, we measured its effect on EMT using Western blot. Cells were treated with TGF- β 1 (10 ng/ml) to induce EMT in the cells. Swainsonine was used at different concentrations (10, 20, 50, and 100 μ g/ml); nontreated cells served as control. As shown in Figure 3, swainsonine increased the expression of E-cadherin but decreased the expression of N-cadherin, vimentin, zinc finger E-box-binding homeobox 1 (ZEB1), and snail in a dose-dependent manner compared to the expressions of the same proteins in the control TGF- β 1-treated cells. These results indicate that swainsonine inhibits EMT in esophageal carcinoma cells.

Effects of Swainsonine on Twist1

We then measured the relative mRNA expression of Twist1 in esophageal carcinoma cells using qRT-PCR. Cells were treated with swainsonine at different concentrations (10, 20, 50, and 100 μ g/ml); nontreated cells

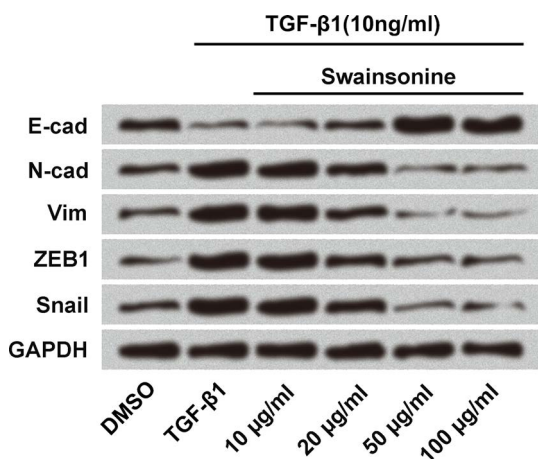


Figure 3. Effects of swainsonine on epithelial–mesenchymal transition (EMT). Esophageal carcinoma cells were treated with transforming growth factor- β 1 (TGF- β 1; 10 ng/ml) to induce EMT in the cells. Swainsonine was used at different concentrations (10, 20, 50, and 100 μ g/ml); nontreated cells served as control. Western blot was used to measure the expression of E-cadherin (E-cad), N-cadherin (N-cad), vimentin (Vim), zinc finger E-box-binding homeobox 1 (ZEB1), and snail in the cells.

served as control. As shown in Figure 4A, swainsonine significantly decreased the mRNA expression of Twist1 in the cells at 50 and 100 μ g/ml concentrations compared with the control (all $p < 0.05$). Similar results were found in the Western blot analysis (Fig. 4B).

Swainsonine Inhibits Cell Invasion and EMT by Downregulation of Twist1 and Deactivation of the PI3K/AKT Signaling Pathway

Esophageal carcinoma cells were transfected with pEX or pEX-Twist1; untransfected cells served as control. We measured mRNA and protein expression of Twist1 in these cells using qRT-PCR and Western blot, respectively. Results showed that pEX-Twist1 significantly increased the mRNA expression ($p < 0.01$) (Fig. 5A) and protein expression (Fig. 5B) of Twist1 in the cells.

For further experiments, cells were treated with TGF- β 1, TGF- β 1 + swainsonine (50 μ g/ml), TGF- β 1 + pEX + swainsonine (50 μ g/ml), or TGF- β 1 + pEX-Twist1 + swainsonine (50 μ g/ml). Nontreated cells served as control. We then measured cell invasion using Transwell invasion

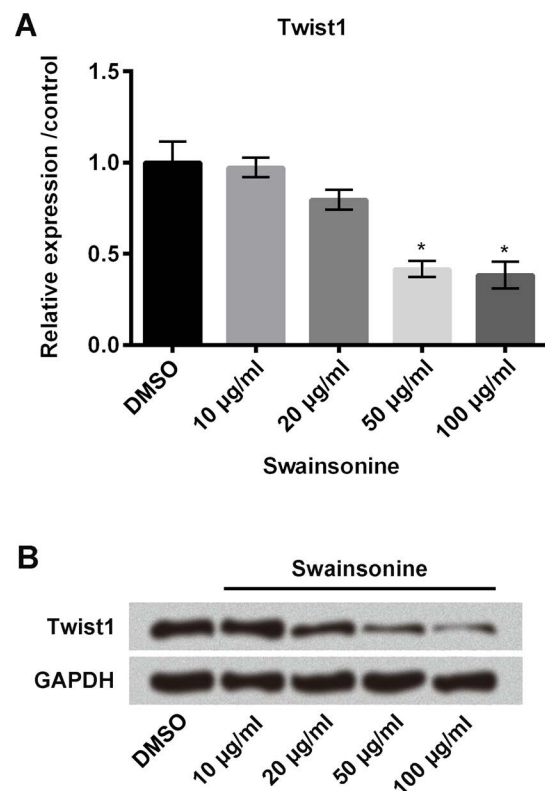


Figure 4. Effects of swainsonine on Twist1. Esophageal carcinoma cells were treated with swainsonine at different concentrations (10, 20, 50, and 100 μ g/ml); nontreated cells served as control. The mRNA and protein expressions of Twist1 were measured using quantitative reverse transcription polymerase chain reaction (qRT-PCR) (A) and Western blot (B), respectively. * $p < 0.05$ compared to control.

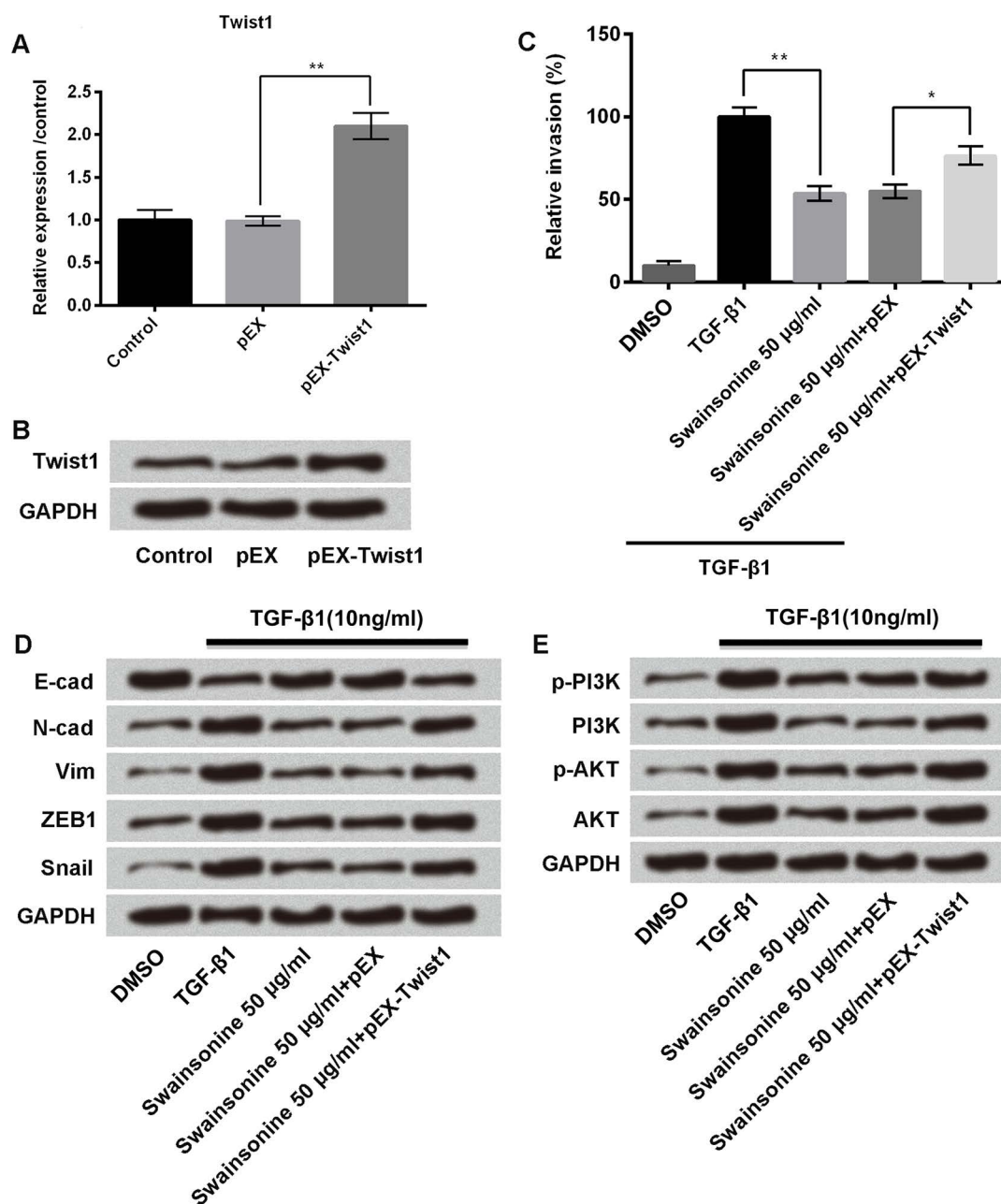


Figure 5. Swainsonine inhibits cell invasion and EMT by downregulation of Twist1 and deactivation of the phosphoinositide 3-kinase (PI3K)/AKT signaling pathway. Esophageal carcinoma cells were transfected with pEX or pEX-Twist1; untransfected cells served as control. The mRNA and protein expressions of Twist1 were measured using qRT-PCR (A) and Western blot (B), respectively. Then cells were treated with TGF-β1, TGF-β1 + swainsonine (50 μg/ml), TGF-β1 + pEX + swainsonine (50 μg/ml), or TGF-β1 + pEX-Twist1 + swainsonine (50 μg/ml), and the following experiments were carried out: (C) Transwell invasion assay, to measure cell invasion; and Western blot, to measure the expression of (D) E-cadherin, N-cadherin, vimentin, ZEB1, and snail, and (E) PI3K and AKT. **p*<0.05, ***p*<0.01 as indicated.

assay. As shown in Figure 5C, TGF-β1 increased cell invasion compared to the control group of cells, but the addition of swainsonine 50 μg/ml decreased the cell invasion (*p*<0.01); transfection of cells with pEX-Twist1 again increased the cell invasion (*p*<0.05) compared to

cells treated with swainsonine 50 μg/ml and transfected with pEX. These results indicate that swainsonine inhibits cell invasion by downregulation of Twist1. Similar findings were observed for the EMT-associated proteins (E-cadherin, N-cadherin, vimentin, ZEB1, and snail):

swainsonine inhibited EMT in the cells by downregulation of Twist1 (Fig. 5D). We also measured the expression of the phosphoinositide 3-kinase (PI3K)/AKT signaling pathway proteins in these cells using Western blot. As shown in Figure 5E, TGF- β 1 increased the expression of PI3K and AKT, but the addition of swainsonine decreased these expressions, whereas the addition of pEX-Twist1 again increased these expressions.

These findings together indicate that swainsonine inhibits cell invasion and EMT in esophageal carcinoma cells by downregulation of Twist1 and deactivation of the PI3K/AKT signaling pathway.

DISCUSSION

In *in vitro* and *in vivo* studies, swainsonine has shown inhibitory and antimetastatic effects on several human and mouse cancer cell lines^{21–25}. These studies provided the rationale for exploring swainsonine as an antitumor agent. In the present study, we evaluated the effects of swainsonine on human esophageal carcinoma cell lines EC9706 and 293T *in vitro* and found that swainsonine inhibits invasion of these cells at 20, 50, and 100 μ g/ml concentrations, with maximal effect at 50 μ g/ml. Similar effects were reported by Seftor et al., who showed that swainsonine, in combination with deoxymannojirimycin, inhibits human melanoma cell invasion *in vitro*²⁶. Additionally, in a phase I study in patients with advanced malignancies, swainsonine reduced tumor cell invasion²¹. Apart from its effect on cell invasion, swainsonine has also been shown to decrease cancer cell growth and induce apoptosis via the mitochondrial pathway in lung cancer and esophageal cancer cells^{19,27}. However, in our study, swainsonine had no effect on cell viability and apoptosis. This could be due to a difference in the types of cells used in these studies.

Cell invasion is followed by cell metastasis. Clinically detectable metastasis is the end product of a complex series of cellular–biological events. During metastatic progression, tumor cells exit the primary site of growth, translocate systemically, and adapt to survive and grow in distant tissues²⁸. Metastasis is an important step in tumor progression and is a main cause of mortality in cancer patients²⁹. After metastasis of esophageal cells to organs or lymph nodes away from the tumor, the 5-year survival rate is only 4%²⁰.

EMT is a crucial process during which epithelial cells acquire mesenchymal properties and show reduced intercellular adhesion and increased motility. Growing evidence suggests that EMT plays an important role during tumor progression and malignant transformation, thereby providing invasive and metastatic properties to the embryonic cancer cells³⁰. The main characteristic of EMT is loss of epithelial surface markers, most notably E-cadherin, and the acquisition of mesenchymal markers, including

N-cadherin and vimentin. E-cadherin is downregulated by EMT, which is mediated by its transcriptional repression through the binding of EMT transcription factors, such as snail, Slug, and Twist, present in the E-cadherin promoter³¹. In cancer, E-cadherin plays a tumor suppressor role, inhibiting invasion and metastasis³⁰. In the present study, swainsonine increased the expression of E-cadherin but decreased the expression of N-cadherin, vimentin, ZEB1, and snail, indicating that swainsonine inhibited EMT in esophageal carcinoma cells.

EMT-related transcription factor, Twist-1, promotes the early steps of metastasis, which include local invasion and subsequent dissemination to distant tissues³². Evidence suggests that Twist1 is overexpressed in several types of cancer, including esophageal cancer³³. Therefore, we measured the effect of swainsonine on the expression of Twist1 and found that swainsonine significantly decreased the expression of Twist1 in esophageal carcinoma cells. In further experiments, we showed that swainsonine inhibits cell invasion and EMT in esophageal carcinoma cells by downregulation of Twist1 and deactivation of the PI3K/AKT signaling pathway. The PI3K/AKT signal pathway is an important pathway for cell survival, proliferation, and migration³⁴. It has been reported that the use of PI3K/AKT inhibitors in combination with chemotherapy may be a potential therapeutic strategy in treating esophageal cancer³⁵.

To the best of our knowledge, the present study is the first to explore the effects of swainsonine on cell invasion and metastasis in esophageal carcinoma cells. In summary, swainsonine decreased esophageal carcinoma cell invasion, but did not affect cell viability and apoptosis; swainsonine decreased the EMT process in the cancer cells by increasing the expression of E-cadherin and decreasing the expression of N-cadherin, vimentin, ZEB1, and snail; swainsonine also decreased the expression of Twist1 and deactivated the PI3K/AKT signaling pathway. Our findings indicate the potential inhibitory role of swainsonine in esophageal carcinoma metastasis.

ACKNOWLEDGMENT: The authors declare no conflicts of interest.

REFERENCES

1. Balmadrid B, Hwang JH. Endoscopic resection of gastric and esophageal cancer. *Gastroenterol Rep.* 2015;3(4): 330–8.
2. Sadjadi A, Marjani H, Semnani S, Nasserimoghaddam S. Esophageal cancer in Iran: A review. *Middle East J Cancer* 2010;1(1):5–14.
3. Napier KJ, Scheerer M, Misra S. Esophageal cancer: A review of epidemiology, pathogenesis, staging workup and treatment modalities. *World J Gastrointest Oncol.* 2014; 6(5):112–20.
4. Brower JV, Chen S, Bassetti MF, Yu M, Harari PM, Ritter MA, Baschnagel AM. Radiation dose escalation in

- esophageal cancer revisited: Contemporary analysis of the National Cancer Data Base, 2004-2012. *Int J Radiat Oncol Biol Phys.* 2016;96(5):985-93.
5. Xie Z, Chen G, Zhang X, Li D, Huang J, Yang C, Zhang P, Qin Y, Duan Y, Gong B. Salivary microRNAs as promising biomarkers for detection of esophageal cancer. *PLoS One* 2013;8(4):e57502.
 6. Islami F, Kamangar F, Aghcheli K, Fahimi S, Semnani S, Taghavi N, Marjani HA, Merat S, Nasserimoghaddam S, Pourshams A. Epidemiologic features of upper gastrointestinal tract cancers in Northeastern Iran. *Br J Cancer* 2004;90(7):1402-6.
 7. Tran GD, Sun XD, Abnet CC, Fan JH, Dawsey SM, Dong ZW, Mark SD, Qiao YL, Taylor PR. Prospective study of risk factors for esophageal and gastric cancers in the Linxian general population trial cohort in China. *Int J Cancer* 2005;113(3):456-63.
 8. De Angelis R, Sant M, Coleman MP, Francisci S, Baili P, Pierannunzio D, Trama A, Visser O, Brenner H, Ardanaz E, Bielska-Lasota M, Engholm G, Nennecke A, Siesling S, Berrino F, Capocaccia R, EURO CARE-5 Working Group. Cancer survival in Europe 1999-2007 by country and age: Results of EURO CARE-5—A population-based study. *Lancet Oncol.* 2014;15(1):23-34.
 9. Short MW, Burgers KG, Fry VT. Esophageal cancer. *Am Fam Physician* 2017;95(1):22-8.
 10. Sohda M, Kuwano H. Current status and future prospects for esophageal cancer treatment. *Ann Thorac Cardiovasc Surg.* 2017;23(1):1-11.
 11. Jiao Y, Shen Y, Yan H, Liu Y, Tan H, Li J. Short-term clinical effect of conformal radiotherapy combined with tegafur gimeracil oteracil potassium in treating recurrent esophageal cancer. *Pakistan J Med Sci.* 2016;32(5):1141-5.
 12. Cook D, Donzelli BG, Creamer R, Baucom DL, Gardner DR, Pan J, Moore N, Jaromczyk JW, Schardl CL. Swainsonine biosynthesis genes in diverse symbiotic and pathogenic fungi. *G3 (Bethesda)* 2017; 7(6):1791-7.
 13. Lu H, Quan H, Ren Z, Wang S, Xue R, Zhao B. The genome of *Undifilum oxytropis* provides insights into swainsonine biosynthesis and locoisim. *Scientific Rep.* 2016;6:30760.
 14. Tulsiani DR, Harris TM, Touster O. Swainsonine inhibits the biosynthesis of complex glycoproteins by inhibition of Golgi mannosidase II. *J Biol Chem.* 1982;257(14):7936-9.
 15. Winchester B, Al DS, Carpenter NC, Cenci dBI, Choi SS, Fairbanks AJ, Fleet GW. The structural basis of the inhibition of human alpha-mannosidases by azafuranose analogues of mannose. *Biochem J.* 1993;290(Pt 3)(3):743-9.
 16. Dorling PR, Huxtable CR, Colegate SM. Inhibition of lysosomal alpha-mannosidase by swainsonine, an indolizidine alkaloid isolated from *Swainsona canescens*. *Biochem J.* 1980;191(2):649-51.
 17. Santos FM, Latorre AO, Hueza IM, Sanches DS, Lippi LL, Gardner DR, Spinosa HS. Increased antitumor efficacy by the combined administration of swainsonine and cisplatin in vivo. *Phytomedicine* 2011;18(12):1096-101.
 18. Li Z, Huang Y, Dong F, Li W, Ding L, Yu G, Xu D, Yang Y, Xu X, Tong D. Swainsonine promotes apoptosis in human oesophageal squamous cell carcinoma cells in vitro and in vivo through activation of mitochondrial pathway. *J Biosci.* 2012;37(1):1005-16.
 19. Li Z, Huang Y, Dong F, Li W, Ding L, Yu G, Xu D, Yang Y, Xu X, Tong D. Swainsonine promotes apoptosis in human oesophageal squamous cell carcinoma cells in vitro and in vivo through activation of mitochondrial pathway. *J Biosci.* 2012;37(6):1005-16.
 20. American Cancer Society. Global cancer facts & figures 3rd edition. Available from <https://www.cancer.org/content/dam/cancer-org/research/cancer-facts-and-statistics/global-cancer-facts-and-figures/global-cancer-facts-and-figures-3rd-edition.pdf>
 21. Goss PE, Baptiste J, Fernandes B, Baker M, Dennis JW. A phase I study of swainsonine in patients with advanced malignancies. *Cancer Res.* 1994;54(6):1450-7.
 22. Dennis JW, Koch K, Yousefi S, Vanderelst I. Growth inhibition of human melanoma tumor xenografts in athymic nude mice by swainsonine. *Cancer Res.* 1990;50(6):1867-72.
 23. Sun JY, Zhu MZ, Wang SW, Miao S, Xie YH, Wang JB. Inhibition of the growth of human gastric carcinoma in vivo and in vitro by swainsonine. *Phytomedicine* 2007;14(5):353-9.
 24. Sun JY, Yang H, Miao S, Li JP, Wang SW, Zhu MZ, Xie YH, Wang JB, Liu Z, Yang Q. Suppressive effects of swainsonine on C6 glioma cell in vitro and in vivo. *Phytomedicine* 2009;16(11):1070-4.
 25. Yagita M, Saksela E. Swainsonine, an inhibitor of glycoprotein processing, enhances cytotoxicity of large granular lymphocytes. *Scand J Immunol.* 1990;31(3):275-82.
 26. Seftor RE, Seftor EA, Grimes WJ, Liotta LA, Stetlerstevenson WG, Welch DR, Hendrix MJ. Human melanoma cell invasion is inhibited in vitro by swainsonine and deoxymannojirimycin with a concomitant decrease in collagenase IV expression. *Melanoma Res.* 1991;1(1):43-54.
 27. Li Z, Xu X, Huang Y, Ding L, Wang Z, Yu G, Xu D, Li W, Tong D. Swainsonine activates mitochondria-mediated apoptotic pathway in human lung cancer A549 cells and retards the growth of lung cancer xenografts. *Int J Biol Sci.* 2012;8(3):394-405.
 28. Valastyan S, Weinberg RA. Tumor metastasis: Molecular insights and evolving paradigms. *Cell* 2011;147(2):275-92.
 29. Hou CH, Lin FL, Hou SM, Liu JF. Cyr61 promotes epithelial-mesenchymal transition and tumor metastasis of osteosarcoma by Raf-1/MEK/ERK/Elk-1/TWIST-1 signaling pathway. *Mol Cancer* 2014;13(1):236.
 30. Larue L, Bellacosa A. Epithelial-mesenchymal transition in development and cancer: Role of phosphatidylinositol 3' kinase/AKT pathways. *Oncogene* 2005;24(50):7443-54.
 31. Serranogomez SJ, Maziveyi M, Alahari SK. Regulation of epithelial-mesenchymal transition through epigenetic and post-translational modifications. *Mol Cancer* 2016; 15(1):18.
 32. Scheel C, Weinberg RA. Cancer stem cells and epithelial-mesenchymal transition: Concepts and molecular links. *Semin Cancer Biol.* 2012;22(5-6):396-403.
 33. Ren H, Du P, Ge Z, Jin Y, Ding D, Liu X, Zou Q. TWIST1 and BMI1 in cancer metastasis and chemoresistance. *J Cancer* 2016;7(9):1074-80.
 34. Chen CH, Sung CS, Huang SY, Feng CW, Hung HC, Yang SN, Chen NF, Tai MH, Wen ZH, Chen WF. The role of the PI3K/Akt/mTOR pathway in glial scar formation following spinal cord injury. *Exp Neurol.* 2016;278:27-41.
 35. Li B, Li J, Xu WW, Guan XY, Qin YR, Zhang LY, Law S, Tsao SW, Cheung AL. Suppression of esophageal tumor growth and chemoresistance by directly targeting the PI3K/AKT pathway. *Oncotarget* 2014;5(22):11576-87.