

Extraintestinal manifestations of inflammatory bowel disease, nitroxidative stress and dysbiosis: What is the link between them?

AMYLLY SANUELLY DA PAZ MARTINS¹; SAMARA BOMFIM GOMES CAMPOS²; MARÍLIA OLIVEIRA FONSECA GOULART^{1,2,3}; FABIANA ANDRÉA MOURA^{4,*}

¹ Programa de Pós-Graduação da Rede Nordeste de Biotecnologia (RENORBIO), Universidade Federal de Alagoas (UFAL), Maceió, AL 57072-970, Brazil

² Programa de Pós-Graduação em Ciências da Saúde (PPGCS), Universidade Federal de Alagoas (UFAL), Maceió, AL 57072-970, Brazil

³ Programa de Pós-Graduação em Química e Biotecnologia (PPGQB/UFAL), Campus A. C. Simões, Avenida Lourival Melo Mota, s/n, Tabuleiro dos Martins, Maceió, AL 57072-970, Brazil

⁴ Programa de Pós-Graduação em Nutrição (PPGNU/UFAL), Programa de Pós-Graduação em Ciências Médicas (PPGCM/UFAL), Universidade Federal de Alagoas (UFAL), Maceió, AL 57072-970, Brazil

Key words: Oxidative stress, Microbiota, Bacterial translocation, Ulcerative colitis, Crohn disease

Abstract: Inflammatory bowel disease (IBD), which includes Crohn's disease and ulcerative colitis, has a not yet completely defined aetiology and is characterized by a progressive chronic inflammation that involves nitroxidative stress and dysbiosis. Extraintestinal manifestations can occur and affect several organs, including the liver and bile ducts, joints, skin, eyes, and less frequently, the heart, brain, and kidneys, increasing the risk of morbidity and mortality. These repercussions may be associated with the activity or severity of IBD. The present review proposes to report and analyse the participation of dysbiosis and nitroxidative stress in the genesis of extraintestinal manifestations, aiming to contribute to a better understanding of the disease and to focus on the development of individualized preventive and therapeutic strategies.

Abbreviations

4-HNE:	4-hydroxy-2-nonenal
AP-1:	activating protein-1
HLA-B27:	B27-human leukocyte antigen
BBB:	blood-brain barrier
BDNF:	brain-derived neurotrophic factor
BMD:	bone mineral density
CAT:	catalase
CCL25:	C-C motif chemokine ligand 25
CNS:	central nervous system
CCR6:	chemokine receptors 6
D161:	cluster of differentiation 161
CRP:	C-reactive protein
CD:	crohn disease
CYP27A1:	cytochrome P450 family 27 subfamily A member 1
CYP7A1:	cytochrome P450 family 7 subfamily A member 1
EIMs:	extraintestinal manifestations

GPx:	glutathione peroxidase
HM:	hepatobiliary manifestations
HCy:	homocysteine
HLA-DR1:	human leukocyte antigen-DR1
IgA:	immunoglobulin A
iNOS:	inducible nitric oxide synthase
IBD:	inflammatory bowel disease
ICAM-1:	intercellular Adhesion Molecule-1
IFN-γ:	interferon gamma
IL:	interleukin
LRR:	leucine rich repeats
LPS:	lipopolysaccharide
MDA:	malondialdehyde
MAPK:	mitogen activated protein kinase
MCP-1:	monocyte chemoattractant protein-1
MAcCAM-1:	mucosal vascular addressin cell adhesion molecule-1
MAIT:	mucosal associated invariant T cells
MS:	multiple sclerosis
NAFLD:	non-alcoholic fatty liver disease
NFκB:	nuclear factor kappa B

*Address correspondence to: Fabiana Andréa Moura, fabiana.moura@fanut.ufal.br

Received: 17 September 2020; Accepted: 20 November 2020



NLRP3:	nucleotide-binding domain, leucine-rich-repeat-containing receptor 3
NOD:	nucleotide-binding oligomerization domain
OmpC:	outer membrane protein C
oxLDL:	oxidized low density lipoprotein
PAMP:	pathogen-associated molecular patterns
PD:	parkinson disease
PPARs:	peroxisome Proliferator-Activated Receptors
PRR:	pattern recognition receptors
ONOO-:	peroxynitrite
PSC:	primary sclerosing cholangitis
RONS:	reactive Oxygen and Nitrogen Species
ROS:	reactive Oxygen Species
SCFA:	short chain fatty acids
STAT3:	signal transducer and activator of transcription 3
SpA:	spondyloarthropathies
SREBP:	sterol Regulatory Element Binding Protein
TLR4:	toll like receptor 4
TRAF3IP2:	tumor necrosis factor alfa receptor-associated factor interacting protein 2
TMA:	trimethylamine
TMAO:	trimethylamine-N-oxide
TNF- α :	tumor necrosis factor alfa
UC:	ulcerative colitis
VCAM-1:	vascular adhesion molecule-1
VAP-1:	vascular adhesion protein-1
VEGF:	vascular endothelial growth factor.

Introduction

Inflammatory bowel disease (IBD) is a chronic and recurrent disease that includes Crohn's disease (CD) and ulcerative colitis (UC). The IBD aetiology is not fully elucidated, however, it is known that there is an intimate interaction between genetic, immunological, and environmental factors (diet, smoking, circadian cycle), the intestinal microbiota and nitroxidative stress (Manichanh *et al.*, 2012; Moura *et al.*, 2016; Garber and Regueiro, 2019; Feuerstein and Cheifetz, 2014).

Intestinal microbiota comprises more than 150.000 species of commensal microorganisms that inhabit the gastrointestinal tract and perform beneficial functions to the host, such as the synthesis of substances important for energy metabolism, defense against luminal pathogens, and modulation of the immune response (Qin *et al.*, 2010; Nishida *et al.*, 2018).

The participation of the microbiota in the modulation of innate and adaptive immunity of the mucosa is fundamental to maintain the integrity of the epithelial barrier and occurs through the interaction between the pathogen-associated molecular patterns (PAMPs) and the pattern recognition receptors (PRR), such as Toll-like receptors (TLR), present in immune cells, as well as by the action of their metabolites, especially short-chain fatty acids (SCFA), which act to induce immunological tolerance by stimulating, mainly, the polarization of regulatory T lymphocytes.

In addition, they secrete or promote the secretion of antimicrobial factors, such as defensins and immunoglobulin

A (IgA) (Kamada and Núñez, 2013; Hart *et al.*, 2005; Nishida *et al.*, 2018).

Imbalance in microbial diversity and density, known as dysbiosis, can alter the interaction between the host-microbiota-immune system and has been associated with the appearance of many inflammatory and autoimmune disorders, including IBD. Some studies have reported that IBD patients have changes in microbial composition, when compared to healthy individuals, with a decrease in SCFA producing commensal bacteria, such as *Faecalibacterium prausnitzii*, and an increase in mucolytic, sulfate-producing, and pathogenic bacteria (Fujimoto *et al.*, 2013; Takahashi *et al.*, 2016; Nishino *et al.*, 2018; Nishida *et al.*, 2018).

It has been described that the increase in intestinal permeability and the unregulated immune response causes a continuous inflammatory process (with neutrophilic infiltration) and a redox imbalance, generating nitroxidative stress, production of pro-inflammatory cytokines, and consequent impairment of the intestinal barrier, characterized by the destruction of tight junctions and oxidative damage caused by lipid peroxidation (Fig. 1) (Moura *et al.*, 2015). This scenario allows pathogenic bacteria and their products, such as lipopolysaccharide (LPS), not only to enter the sterile submucosa and activate immune cells through the binding and recognition of their PAMPs in the respective PRRs but also favour microbial translocation, via current blood, to other organs and tissues, characterizing extraintestinal manifestations (EIM), with an impact on the functional status of the patients and their quality of life (Fig. 2) (Knutson *et al.*, 2013; Hussein *et al.*, 2008).

EIM can be identified in 25–40% of patients with IBD. The variation between remission and activity of symptoms is linked to morbidity and mortality. The main described manifestations include those that affect the liver and bile ducts, the skin, joints, eyes, and blood vessels. The heart, brain, and kidney are also affected (Annese, 2019; Garber and Regueiro, 2019). In addition, a common link regarding the microbial composition/nitroxidative stress of IBD and some diseases involving extraintestinal organs has been reported. This review aims to elucidate the participation of nitroxidative stress and dysbiosis in EIM genesis.

Materials and Methods

A narrative review of the literature was carried out on the association of inflammation, oxidative and nitrosative stress, and dysbiosis in the onset of EIM of IBD using the PubMed database. Systematic reviews, *in vivo* and *in vitro* studies (rats and mice) were included, with a total of 200 papers. Studies carried out on dogs, rabbits, pigs, or monkeys were not included in this review. The following keywords were used: oxidative stress, intestinal microbiota, dysbiosis, ulcerative colitis, Crohn's disease, inflammatory bowel disease, liver, hepatic, joint, skin, eyes, ophthalmological, thromboembolism, cardiovascular diseases, heart, brain, cerebral, neurologic, renal, kidney, lipopolysaccharide, endotoxin.

Systematic description

After an extensive review, the results are displayed in the following topics: hepatobiliary; osteoarticular, dermatological

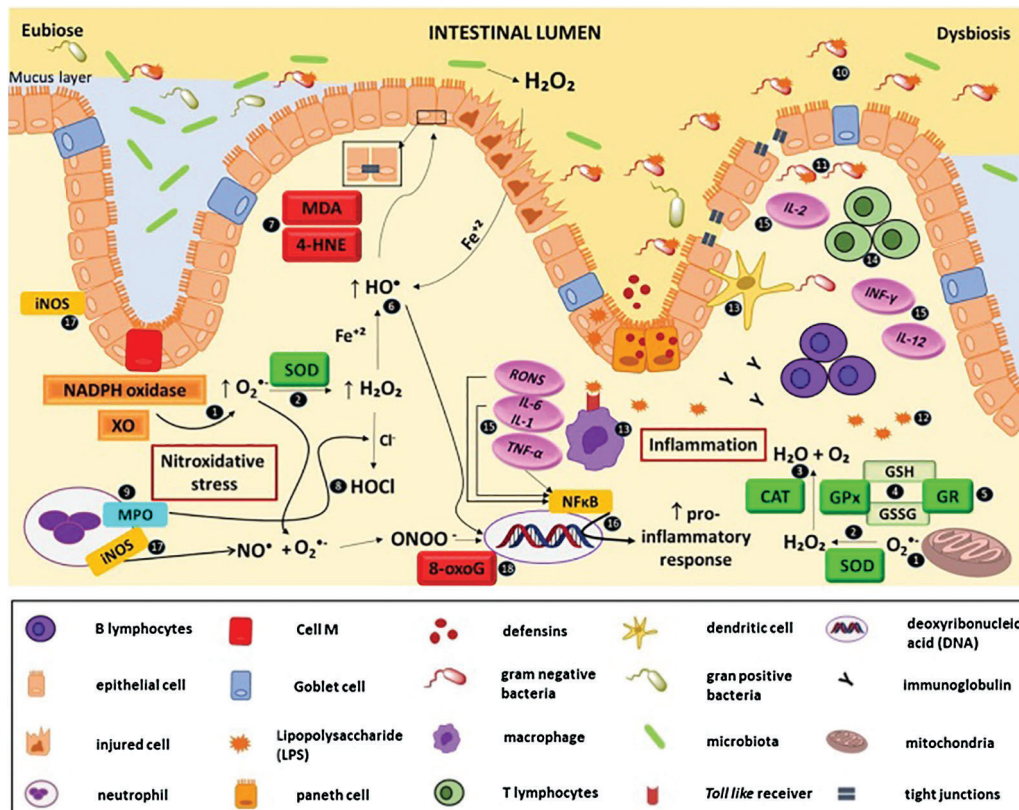


FIGURE 1. Interaction between dysbiosis, nitroxidative stress and inflammation in inflammatory bowel disease.

Legend: Representation of the interaction between dysbiosis, nitroxidative stress and inflammation involved in the pathophysiology of inflammatory bowel disease (IBD). The activation of xanthine oxidase (XO), NADPH (nicotinamide adenine dinucleotide phosphate) oxidase enzyme complex, and mitochondrial stimuli generate significant amounts of the superoxide radical anion ($O_2^{\bullet-}$) (1). In the cytosol, this reactive species, through superoxide dismutase (SOD), is quickly converted into hydrogen peroxide (H_2O_2) (2), which in turn is decomposed into H_2O and O_2 , through the activity of catalase (CAT) and glutathione peroxidase (GPx) –formed by oxidation of GSH (glutathione reduced) to GSSG (glutathione oxidized) (4). GSSG is converted back to GSH by glutathione reductase (GR) (5). The generated redox imbalance causes a decrease in these antioxidant defences. The presence of transition metals, such as ferrous iron (Fe^{2+}), converts H_2O_2 into the hydroxyl radical (HO^{\bullet}) (6) that indiscriminately oxidizes cell membrane proteins and phospholipids (lipid peroxidation), causing epithelial barrier dysfunction. The lipid peroxidation generates toxic aldehydes, such as malondialdehyde (MDA) and 4-hydroxynonenal (4-HNE) (7), which are also the result of the action of hypochlorous acid (HOCl) (8) formed from the reaction between H_2O_2 and chloride (Cl^-), through the activity of myeloperoxidase (MPO), secreted from neutrophil granules (9). All those alterations, associated with dysbiosis (10), make the environment conducive to bacterial translocation (11), which consists of the entry of bacteria, mostly gram-negative, and their endotoxins, such as lipopolysaccharide (LPS), in the sterile sub-mucous (12). These microbes interact with immune cells (dendritic cells and macrophages) (13), through the link between pathogen-associated molecular patterns (PAMPs) and pattern recognition receptors (PRRs), respectively, and activate T lymphocytes (14), which promote the secretion of proinflammatory cytokines (15) that contribute to the maintenance of the inflammatory arsenal. These inflammatory cytokines and RONS activate the nuclear factor kappa B (NfκB) (16), which in turn stimulates, even more, the proinflammatory response. Reactive nitrogen species also participate, where the nitric oxide (NO) derived from inducible nitric oxide synthase (iNOS) (17), can react with $O_2^{\bullet-}$ and generate peroxynitrite ($ONOO^-$), which in turn, acts causing deoxyribonucleic acid (DNA) fragmentation, increasing metabolites such as 8-oxoguanine (8-oxoG) (19). Additionally, nitroxidative stress causes recruitment of inflammatory cells that generate additional damage and injury into intestinal microenvironment.

and ophthalmological; thromboembolic; cardiovascular; neurological; and renal manifestations.

Hepatobiliary manifestations

Hepatobiliary manifestations (HM) comprise one of the most common repercussions in IBD, with a prevalence of 3–50%, especially in those with UC. In addition, up to 5% of adults with IBD will develop some liver disease (Mendes *et al.*, 2007; Restellini *et al.*, 2017; Silva *et al.*, 2019; Venkatesh *et al.*, 2011). Primary sclerosing cholangitis (PSC) and non-alcoholic fatty liver disease (NAFLD) are the most common forms and can occur at any time during the course of intestinal disease or before its diagnosis (Annese, 2019; Fousekis *et al.*, 2018).

PSC is a progressive chronic cholestatic disease, characterized by inflammation, stenosis, fibrosis, and obstruction of the intra- and extrahepatic ducts, which can progress with complications such as cirrhosis, liver failure, and portal hypertension, in addition to an increased risk of cholangiocarcinoma and colorectal cancer (Eaton and Talwalkar, 2013; Maggs and Chapman, 2008; Tsaitas *et al.*, 2014). Epidemiological studies demonstrate that 60–80% of patients with PSC have IBD, especially UC (approximately 75%), and up to 8% of patients with IBD have PSC (Hirschfield *et al.*, 2013; Mendes *et al.*, 2007; Tsaitas *et al.*, 2014).

NAFLD, in turn, characterized by excess fat deposited in the liver, is responsible for up to 40% of the hepatic repercussions of

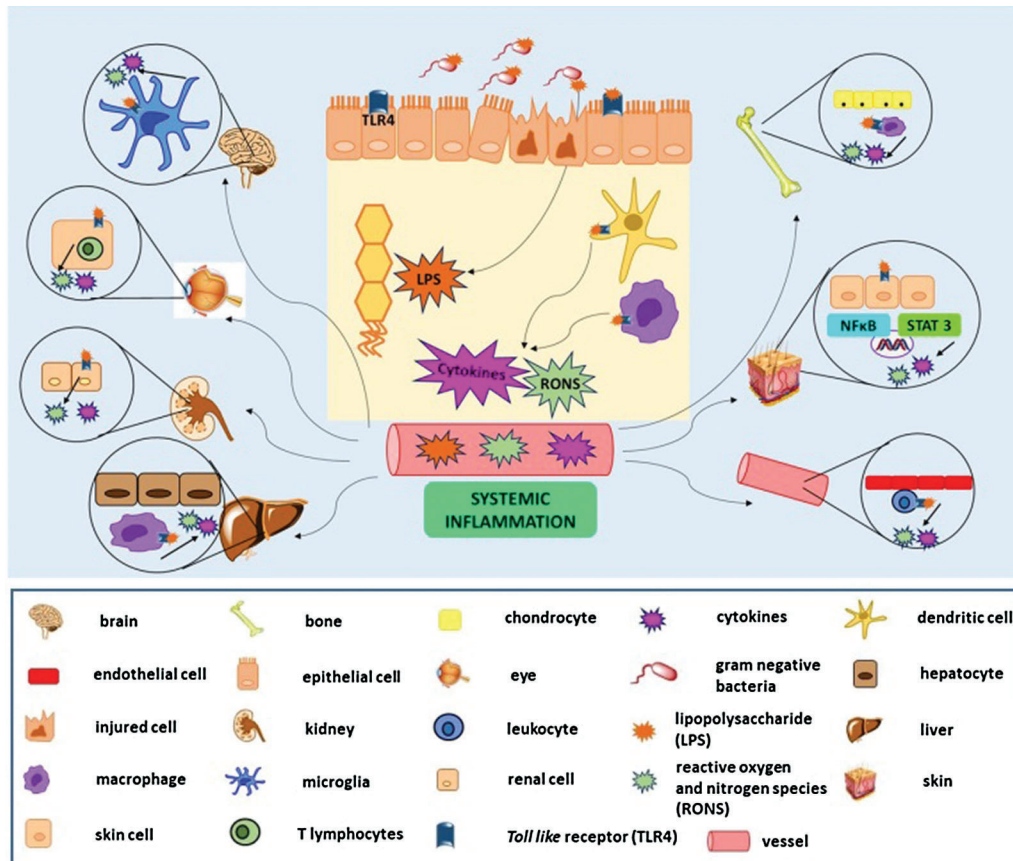


FIGURE 2. Microbiota translocation in inflammatory bowel disease promotes systematic inflammation and affects several tissues, causing local production of reactive oxygen and nitrogen species (RONS) and cytokines.

Legend: Bacterial translocation and nitrosidative stress contribute significantly to unregulated and exacerbated inflammatory response. The interaction between endotoxins, such as LPS, and their pattern recognition receptors, Toll like receptors, in particular TLR 4, in immune cells, causes secretion of pro-inflammatory cytokines and RONS, which cause harmful damage to all intestinal barriers and allow the spread of these toxic products through the bloodstream, which can be deposited and activate the immune response in extraintestinal organs. In these organs, LPS finds its TLR on the surface of innate immunity cells, causing the secretion of molecules with inflammatory profile and RONS, with consequent recruitment of other cells of the immune system, which intensifies the inflammatory cascade, generating local damage and contributing to the appearance of diseases. NFκB: nuclear factor kappa B; STAT 3: signal transducers and activators of transcription 3.

the non-alcoholic subtype in patients with IBD (Magri *et al.*, 2019; Vernon *et al.*, 2011). The prevalence is higher when compared to the general population, ranging between 1.5% and 55% (Chao *et al.*, 2016; Palumbo *et al.*, 2019).

The link between HM and IBD is still not well understood; however, multiple factors are involved, including genetic predisposition (HLA-B8, HLA-DRB1* 0301, HLA-DRB3* 0101, HLA-DRB1* 0401, REL, IL2 and CARD9) and hepatotoxicity induced by the use of drugs (corticosteroids, aminosalicylates, methotrexate, thiopurine, and antitumour necrosis factor-alpha) (Chapman *et al.*, 2010; Eaton and Talwalkar, 2013; Janse *et al.*, 2011; Karlsen *et al.*, 2010). However, the role of intestinal dysbiosis, inflammation, and nitrosidative stress also stands out, resulting from immune-mediated processes (Fousekis *et al.*, 2018; Navaneethan, 2014; Restellini *et al.*, 2017).

Bacterial translocation, evidenced by the significant increase in the levels of endotoxins and LPS in the portal vein in humans and animal models, implies the participation of the intestinal microbiota in the pathogenesis of HM (Nakamoto *et al.*, 2019).

Changes in the intestinal microbiota and dysfunction of the epithelial barrier play a crucial role in PSCs concomitant

with IBD, as evidenced in humans. Intestinal dysbiosis—generally called dysbiosis—is an inherent characteristic of IBD, participating in the perpetuation and maintenance of chronic intestinal inflammation (Fujimoto *et al.*, 2013; Nishida *et al.*, 2018).

In humans, cohort studies have reported changes in microbial diversity and composition and have identified distinct phenotypes among patients with PSC and/or IBD when compared to healthy controls. It is noteworthy the significant decrease in the strain *Faecalibacterium prausnitzii*, present in both PSC and patients with IBD (Bajer *et al.*, 2017; Quevrain *et al.*, 2016; Sabino *et al.*, 2016). This dysbiosis may be associated with an unregulated mucosal immune response and altered permeability that directs a local and extraintestinal inflammatory response through bacterial translocation. It has been suggested that in the liver, endotoxins, especially LPS, bind to Toll-like receptor 4 (TLR 4) and activate dendritic cells and macrophages, which are involved in the secretion of pro-inflammatory cytokines, such as tumour necrosis factor-alpha (TNF-α) and reactive oxygen and nitrogen species (RONS); the expression of adhesion molecules, such as intercellular adhesion molecule 1 (ICAM-1) and vascular

adhesion molecule 1 (VCAM-1); and in the initiation and progression of the fibrotic process characteristic of PSC (Reyes-Gordillo *et al.*, 2017).

In an animal model of PSC, dysbiosis was associated with changes in the enterohepatic circulation of bile acids, damage to the mucus layer, reduced expression of tight junction proteins, and increased bacterial translocation (Liao *et al.*, 2019).

Furthermore, a review showed that activation of the inflammasome nucleotide-binding domain, leucine-rich containing family, pyrin domain-containing-3 (NLRP3) in the intestine-liver axis by bacterial products contributed to the progression of liver damage through secretion of interleukin (IL)-1 β and IL-18 via caspase-1 activity (Liao *et al.*, 2019), which is related to the amplification of hepatocellular damage (Zmora *et al.*, 2017).

Intestinal dysbiosis has also been recognized as a predisposing factor for NAFLD. The binding of bacterial DNA to TLR9 in Kupffer cells has been reported in animal studies to stimulate the secretion of IL-1 β , which amplifies steatosis and liver fibrosis, in a MyD88-dependent mechanism (Miura *et al.*, 2010; Muruve *et al.*, 2008; Purchiaroni *et al.*, 2013).

The involvement of *Klebsiella pneumoniae* in NAFLD, a gram-negative bacterium associated with lung infections, has also been investigated. Recurrent infections by this resistant pathogenic bacterium stimulate an influx of pro-inflammatory cytokines in the colon and ileum, and the increased colonization of *K. pneumoniae* induced UC in an animal model, where it was able to increase the expression of cyclooxygenase-2 (COX-2), IL-6, IL-1 β and TNF- α and the levels of nitric oxide (NO *) and reduce tight junction proteins (Kaur *et al.*, 2018; Lee and Kim, 2011; Zhou *et al.*, 2009). Additionally, a study in mice demonstrated that these animals developed liver steatosis after faecal microbiota transplant containing *K. pneumoniae* isolated from a patient with NAFLD. Lastly, a cohort study reported a strong association between this bacterium and the severity of NAFLD in 61% of individuals. This result suggested that dysbiosis causes this condition due to excessive alcohol production, as this bacterium is associated with ethanol production in the faeces of patients with NAFLD when compared to healthy patients, and, consequently, ROS production and mitochondrial dysfunction (Chen *et al.*, 2020; Yuan *et al.*, 2019).

In dysbiosis, redox imbalance plays a crucial role in the damage, apoptosis/necrosis, and formation of toxic aldehydes (lipid peroxidation) (Leung and Nieto, 2013; Shearn *et al.*, 2018).

Significant periportal inflammation in the PSC marked by neutrophilic infiltrate and activation of NADPH oxidase is related to the generation of elevated levels of lipid peroxidation products and derivatives, such as malondialdehyde (MDA), acrolein, and 4-hydroxy-2-nonenal (4 HNE), which are important markers in the liver associated with protein and deoxyribonucleic acid (DNA) damage. Toxic aldehydes can cause post-translational changes in certain proteins, causing protein carbonylation, which determines the toxicity and degree of inflammation/fibrosis in the liver (Osna *et al.*, 2016; Shearn *et al.*, 2018; Shearn *et al.*, 2019).

In an animal model of PSC, an increase in periportal oxidative stress depending on the stage of cholestasis was demonstrated (Shearn *et al.*, 2019). Corroborating these findings, human studies carried out in patients with PSC concomitant with IBD showed an increase in periportal oxidative stress, with the elevated presence of toxic aldehydes, inflammation (\uparrow lymphocytes, \uparrow Kupffer cells and \uparrow MPO), and unregulated antioxidant response (Shearn *et al.*, 2018).

Additionally, the increase in intestinal permeability observed in IBD patients allows lymphocytes, activated in the intestine, to enter the enterohepatic circulation and cause inflammation in the liver. This promotes the recruitment of adhesion molecules and chemokines, such as MAdCAM-1 (mucosal vascular addressin cell adhesion molecule-1) and CCL25 (C-C motif chemokine ligand 25), which are chemotactic for other inflammatory cells, such as macrophages and dendritic cells (Adams and Eksteen, 2006; Eksteen *et al.*, 2004; Grant *et al.*, 2002).

In NAFLD, increased β -oxidation in peroxisomes and microsomes generates hydrogen peroxide (H $_2$ O $_2$) not coupled to phosphorylation and cytochromes P450E1 and P450A, respectively (Bellanti *et al.*, 2017; Robertson *et al.*, 2001). The imminent redox imbalance and the elevation of lipids, associated with glutathione reductase (GR) depletion, can lead to the formation of intermediate lipids and cause stress of the endoplasmic reticulum, which in turn are related to inflammation and apoptosis (Bellanti *et al.*, 2017; Higa and Chevet, 2012; Mari *et al.*, 2006; Pagliassotti, 2012).

In humans, it was seen that the accumulation of lipids in the liver, especially cholesterol, alters the cellular redox status by prominently activating the oxidative pathways in the mitochondria, causing a significant increase in the formation of ROS that surpasses the neutralizing capacity of endogenous antioxidant defenses, leading to mitochondrial dysfunction (Muriel, 2009; Serviddio *et al.*, 2011; Sunny *et al.*, 2011).

Moreover, lipid peroxidation products, due to overproduction of ROS, influence the progression of NAFLD by complex inflammatory mechanisms, including inhibition of the peroxisome proliferator-activated receptors (PPARs) and TLR 7 pathways and activation of nuclear factor kappa B (NF κ B) and activating protein 1 (AP-1) (Green and Wahli (1994)). These factors stimulate the production of pro-inflammatory cytokines, especially TNF- α and IL-1 β , which increase the expression of sterol regulatory element-binding protein (SREBP), responsible for the transcription of genes that encode enzymes involved in the synthesis of lipids and the later appearance of steatosis (Bellanti *et al.*, 2017; Chen *et al.*, 2008; Kohjima *et al.*, 2007).

Therefore, the involvement of dysbiosis/inflammation/nitroxidative stress as mediators of hepatobiliary lesions triggered by IBD seems to be increasingly consistent and should be considered in clinical practice.

Osteoarticular, dermatological and ophthalmological manifestations

These manifestations occur even before the diagnosis of UC, correlating or not with disease activity (Olpin *et al.*, 2017). Genetic susceptibility has been proposed to explain the link between then and IBD, such as TNF- α and variations in TNF receptor-associated factor interacting protein 2 (TRAF3IP2) that occur in erythema nodosum (Ciccacci *et al.*, 2013;

Orchard *et al.*, 2002; Suh *et al.*, 2019; Timani and Mutasim, 2008). In this context, in some of these diseases, genetic alterations related to microbiota disruption have been described, including ankylosing spondylitis and uveitis, which were identified as the presence of B27 human leukocyte antigen (HLA-B27) that predisposes individuals to dysbiosis (Ciccia *et al.*, 2016; Costello *et al.*, 2015; Hermann *et al.*, 1993; Lin *et al.*, 2014a; Pimentel-Santos *et al.*, 2013; Sheth *et al.*, 2015; Specia and Dubuquoy, 2017).

However, genetic factors alone are unable to elucidate the pathophysiological pathways, suggesting the involvement of environmental factors. It has been proposed that intestinal lumen antigens can form circulating immune complexes and could be deposited in these organs, causing an inflammatory response and RONS production, which are responsible for the characteristic lesions of the disease (Lin *et al.*, 2014a; Lin *et al.*, 2014b; Yang *et al.*, 2016).

Osteoarticular manifestations: Spondyloarthropathies (SpA), including ankylosing spondylitis and peripheral arthritis, are found in 10–39% of patients with IBD, especially in patients with CD (Fantini *et al.*, 2009; Gionchetti *et al.*, 2015; Karreman *et al.*, 2017). Some mechanisms are described to explain this association. One of these findings suggests that lymphocytes and macrophages activated in the Payer plate and in the mesenteric lymph nodes start to express and stimulate cell and vascular adhesion molecules ($\alpha 4\beta 7$ and $\alpha E\beta 7$ integrins, cadherin E, vascular adhesion protein 1 [VAP-1], ICAM-1) that promote the adherence of these immune cells in the synovial endothelium and inside the joints, releasing cytokines directed to synovial fibroblasts. These findings were corroborated in samples from the affected part of the intestine of patients with CD and CU, where it was demonstrated that the leukocyte populations of the inflamed intestine were avidly bound to synovial vessels, evidenced by the expression of adhesion molecules and ligands (ICAM-1 and P-selectin) (Jacques and Elewaut, 2008; Salmi and Jalkanen, 2001).

Another link described in some studies suggests that the mechanisms of IBD-associated SpA involve a hypothesis of “intestine-synovia-joint axis”, including changes in the intestinal microbiota and activation of T cells in the intestine for liquid synovial joint in genetically predisposed individuals (Arvikar and Fisher, 2011; Fragoulis *et al.*, 2019; Brakenhoff *et al.*, 2010).

In this context, germ-free and transgenic animals for the HLA-B27 gene (related to joint disease) did not develop inflammation or lesions characteristic of colitis and spondyloarthritis (Gilis *et al.*, 2018; Scher *et al.*, 2016; Taurog *et al.*, 1994).

Additionally, it has been reported in patients with IBD and SPA overexpression of E-cadherin, the major component of adhering junctions, responsible for maintaining intestinal homeostasis and barrier function, indicating the participation of microbiota/endotoxins in these conditions (Demetter *et al.*, 2000; Demetter *et al.*, 2005).

These changes in microbiota and intestinal permeability allow the translocation of bacteria and endotoxins, such as LPS, to the synovial fluid and joints, causing local inflammation (Asquith *et al.*, 2014). Dysbiosis induces the M1 phenotype in macrophages, which are responsible for

increasing the production of pro-inflammatory cytokines (Yang *et al.*, 2016). Ciccia *et al.* indicated in intestinal biopsy samples from patients with SpA that infiltrating monocytes were responsible for the increased IL-23 expression, and overexpression of IL-23 is an important characteristic of subclinical gut inflammation in SpA. Further, to confirm this theory of “intestine-synovia-joint axis”, a reduction in the number of *Faecalibacterium prausnitzii* species, which are beneficial species for intestinal health and the immune system, has been described in the faeces of patients with SpA and IBD (Tito *et al.*, 2017).

Together, these findings (in experimental and human studies) suggest the involvement of microbiota homeostasis in the immune response and molecular mimicry in patients with SpA and IBD.

Osteoporosis is another osteoarticular disease common in patients with IBD and has been identified in 18–42% of these, increasing the fracture risk, especially in older patients (>60 years). However, it is not clear if IBD patients have an increased risk of reduced bone mineral density (BMD) (van Hogezaand and Hamdy, 2006).

Osteoporosis in IBD is multifactorial, but medications used in IBD treatment seems to be the main cause of BMD in these patients. Chronic steroid therapy may decrease BMD, damage bone tissue structure, increase the risk of fractures (van Staa *et al.*, 2005), and, by the other side, decrease intestinal calcium absorption and calcium kidney reabsorption, leading to an increase in the parathormone level, responsible for the stimulation of osteoclasts and increasing bone loss (van Staa, 2006).

The link between osteoporosis/IBD/oxidative stress/microbiota is not evident. However, in a recent review of Ratajczak *et al.* (Ratajczak *et al.*, 2020), the authors did an association between vitamin C deficiency and risk of osteoporosis in IBD patients. According to then, as an antioxidant, vitamin C: (1) Decreases the level of ROS, which increase bone resorption, throughout the activation of (NF- κ B) which is a crucial mediator of TNF- α and osteoclastogenesis; (2) Creates a redox state and modulates gut microbiota increasing Firmicutes and a decreased Bacteroides level. All dates together suggest an important role of antioxidant status in bone health to IBD patients. However, the exact mechanism is unclear and must be studied posteriorly.

Dermatological manifestations: Erythema nodosum and pyoderma gangrenosum are the most frequent EIM on the skin of IBD (Greuter and Vavricka, 2019; Vavricka *et al.*, 2011). Nodosum erythema is the most common symptom, more present in CD than in UC, especially in women (Farhi *et al.*, 2008; Vavricka *et al.*, 2011; Vavricka *et al.*, 2015). Its relationship with IBD is more associated with genetic factors (Suh *et al.*, 2019; Timani and Mutasim, 2008).

Gangrenous pyoderma, in turn, mainly affects patients with UC to the detriment of those with CD and can appear before or after the diagnosis of IBD (Ahn *et al.*, 2018; Annese, 2019). The pathophysiological link is still unclear, but an unregulated inflammatory response is proposed, involving neutrophils and T cells against antigens common to the organs. Data from immunohistochemistry and analysis of proteins in necrotic tissue in patients with gangrenous pyoderma have demonstrated overexpression of

the TNF- α , NF- κ B, and signal transducer and activator of transcription 3 (STAT3) pathways (Vavricka *et al.*, 2015).

Psoriasis, a chronic inflammatory skin disease, has also been associated with IBD and is more prevalent in patients with CD than in those with UC (Fu *et al.*, 2018; Suh *et al.*, 2019). A recent meta-analysis also showed that psoriasis was significantly related to IBD (Fu *et al.*, 2018), being more prevalent in these patients than in the general population.

Despite the genetic link (IL23R and IL12B), it has been proposed that the intestinal microbiota plays an important role in this context (Capon *et al.*, 2007; Cho, 2008; Cottone *et al.*, 2019; Ellinghaus *et al.*, 2012). The decrease in the genus *Faecali bacterium prausnitzii* (a beneficial bacterium for the maintenance of intestinal homeostasis, as already mentioned in this review), observed in both patients with CD and psoriasis, demonstrates the relevance of an interaction between the microbiome and an adequate immune response (Eppinga *et al.*, 2016).

Ophthalmological manifestations: Ophthalmological manifestations may be present in 4–12% of patients with IBD; however, the reported prevalence rate can reach up to 29%, especially in CD. The most common conditions reported are anterior uveitis and episcleritis, present in 0.3–10% of patients with IBD (Greuter and Vavricka, 2019; Harbord *et al.*, 2016; Larsen *et al.*, 2010; Taleban *et al.*, 2016).

The hypothesis of the microbiota-intestine-eye has been proposed and investigated. In animal models of autoimmune uveitis, it has been shown that dysbiosis induces the activation of retinal-specific autoreactive T cells and intraocular inflammation (Horai and Caspi, 2019; Horai *et al.*, 2015; Nakamura *et al.*, 2016). In addition, there was an increase in intestinal permeability and antimicrobial peptide expression concomitant with the effector T cell response in the initial stage of uveitis (Janowitz *et al.*, 2019).

In addition, nitroxidative stress, identified through MDA elevation levels in the aqueous humour and decreased antioxidant defense (superoxide dismutase [SOD], catalase [CAT] and glutathione peroxidase [GPx]), has been confirmed in an endotoxin-induced anterior uveitis model (Rahman and Biswas, 2004).

Recent cohorts, including individuals with autoimmune anterior uveitis, identified changes in the composition of the intestinal microbiota and a faecal metabolic phenotype that significantly differed when compared to that of healthy controls (Huang *et al.*, 2018; Chakravarthy *et al.*, 2018). However, the scarcity of studies in humans that have evaluated the intestine in eye diseases hinders the effective association.

Despite these molecular findings suggesting the close link between microbiota, nitroxidative stress, intestinal inflammation and osteoarticular, dermatological and ophthalmological manifestations in patients with IBD, these crosslinks are not yet well understood, and additional studies, especially in humans, are still needed.

Thromboembolic manifestations

The prothrombotic state is considered a characteristic of patients with IBD, especially during the symptomatic phase (Nguyen *et al.*, 2014). The most common manifestations are deep venous thromboembolism and pulmonary embolism (van Assche *et al.*, 2013). Chronic inflammation (monocyte

chemoattractant protein-1 [MCP-1], IL-6 and IL-8) activates hemostasis and causes hypercoagulation and, consequently, abnormalities in the microvascular tissue characteristic of endothelial dysfunction (Danese *et al.*, 2007; Esmon, 2005; Levi *et al.*, 2012; Papa *et al.*, 2008; Roifman *et al.*, 2009; Zegos *et al.*, 2014).

Several factors cause lesions in the gut microvascular endothelium, among which the most important described are the bacterial endotoxins present in the gut lumen, pro-inflammatory cytokines, and hypoxia (Danese, 2007; Joseph *et al.*, 2002; Levi *et al.*, 2012; Makrides, 1998; Stadnicki, 2012; Zegos *et al.*, 2014).

Once activated, the vascular endothelium starts to express a large amount of cell adhesion molecules (ICAM-1, VCAM-1, and PCAM-1) and adhesins (selectins and integrins) that allow the recruitment and transmigration of leukocytes through the vessel, platelet activation, and aggregation (Davis *et al.*, 2003; Panes and Granger, 1998; Schuermann *et al.*, 1993). This scenario promotes the coagulation cascade, with an evident increase in the factors involved in this process (factors V, VIII, von Willebrand, and fibrinogen), thus enabling the formation of thrombi. Vascular and tissue damage spread through a vicious cycle characterized by an increase in the production of cytokines, especially TNF- α , IL-6 and vascular endothelial growth factor (VEGF), RONS and chemokines (Danese, 2007; Hudson *et al.*, 1992; Jorens *et al.*, 1990; Scaldaferrri *et al.*, 2011; Zegos *et al.*, 2014). Increased expression of VEGF and its receptor was seen in samples from patients with IBD when compared to healthy individuals (Scaldaferrri *et al.*, 2009).

Furthermore, vascular dysfunction in IBD was associated with an imbalance in RONS levels. Inflammatory and immunological stimuli, bacteria and LPS have been shown to activate inducible nitric oxide synthase (iNOS), which induces the production of large amounts of NO^* —a potent vasodilator and anti-aggregation agent that causes indirect harmful effects through the generation of other species, such as nitroxyl anion (NO^-) and peroxynitrite (OONO^-), which is able to cause lipoperoxidation and damage to DNA molecules (Beckman and Koppenol, 1996; Kolios *et al.*, 2004). Studies with colonic biopsies of patients with CD and UC, showed overexpression of iNOS in the inflamed epithelium and nitrotyrosine, suggesting that this finding is associated with the formation of OONO^- and the nitration of cellular proteins (Singer *et al.*, 1996; Dijkstra *et al.*, 1998).

In addition, literature reviews show that in the endothelial cells of the chronically inflamed microvasculature, there is a decrease in the production of NO^* , mainly due to the selective inhibition of iNOS and the non-selective inhibition of the constitutive forms of nitric oxide synthase (NOS), and an increase in superoxide anion radical ($\text{O}_2^{\bullet-}$), which was proportional to the increase in recruitment and leukocyte adhesion (Binion *et al.*, 1998; Binion *et al.*, 2000; Hatoum *et al.*, 2003).

The increase in the serum levels of homocysteine (HCy), a sulfur-containing amino acid resulting from the demethylation of methionine, is also associated with an elevated risk of venous thromboembolism by inducing platelet activation in the endothelium and increasing prothrombotic components (Zhang *et al.*, 2014). In a dysbiotic environment, the bacteria

that synthesize methionine contribute to excessive HCy formation (Kurilshikov *et al.*, 2019).

Intestinal dysbiosis and bacterial translocation have also been linked to endothelial dysfunction. A review described that, in the bloodstream, microbial endotoxins present in the outer membrane of gram-negative bacteria, especially LPS, bind to TLR4 in immune cells, forming a complex that binds to the MD-2 protein and CD14. This complex formation results in the production of pro-inflammatory cytokines and other mediators involved in endothelial damage, the procoagulant state, the recruitment and transformation of macrophages into foam cells, and the initiation of atherosclerotic plaque (Szeto *et al.*, 2018a; Szeto *et al.*, 2018b).

Therefore, the activation of the endothelium associated with chronic bowel inflammation and dysbiosis may be critically implicated in triggering hypercoagulability and subsequent onset of thromboembolic events.

Cardiovascular manifestations

Patients with IBD have a high risk of coronary artery disease, in part, attributed to a greater susceptibility to the occurrence of thromboembolic events and nutrient malabsorption, especially selenium (Benstoem *et al.*, 2015; Castro Aguilar-Tablada *et al.*, 2016; Hansson, 2005; Wu *et al.*, 2017). On the other hand, it has been suggested that the risk of atherosclerosis and other cardiovascular diseases is associated with endothelial dysfunction caused by systemic inflammation, dysbiosis, and nitroxidative stress (Aniwan *et al.*, 2018; Bigeh *et al.*, 2019; Horowitz *et al.*, 2007; Wu *et al.*, 2017).

Systemic inflammation in IBD leads to RONS generation and nitroxidative stress (Wu *et al.*, 2017; Zanolini *et al.*, 2015). A study in patients with CD observed high levels of pro-inflammatory molecules such as TNF- α , IL-1, IL-6 and C-reactive protein (CRP) contribute to endothelial dysfunction, characterized by vascular smooth muscle cell hyperplasia and decreased *NO production, resulting in a reduction in vessel relaxation. This induces the infiltration of neutrophils into the blood vessels, causing changes in the smooth muscle cell phenotype, an increase in matrix metalloproteinase production, and a decrease in elastin and collagen fibres due to the activation of collagenases and elastases, thus leading to the formation of rigid fragments that induce atherosclerotic processes (Schinzari *et al.*, 2008). Moreover, TNF- α signaling pathways induce the expression of osteoblast markers (osteocalcin and osteopontin) in endothelial cells, which leads to an increase in calcification and reduced vascular elasticity, resulting in arterial stiffening, coronary artery disease, and heart failure (Floege and Ketteler, 2004; Wu *et al.*, 2017; Zanolini *et al.*, 2015; Zieman *et al.*, 2005).

Intestinal dysbiosis, in turn, associated with increased permeability, allows bacterial translocation. Has been demonstrated in *in vitro* assays (Howell *et al.*, 2011; Maziere *et al.*, 1999) and an animal model (Wiesner *et al.*, 2010) that high levels of LPS and bacterial products directly induce the atherosclerotic process through the formation of oxidized low-density lipoprotein (oxLDL), a key atherosclerotic lesion component, activation of macrophages and adhesion molecules and, consequently, the formation of foam cells

and stimulation of pro-inflammatory cytokine expression. The latter, in association with other cellular mediators such as VEGF, CRP, and platelets, cause endothelial dysfunction and eventual atherosclerosis.

A recent cohort study exploring the relationship of intestinal microbiota with plasma metabolites, cardiovascular metabolic risk score, and cardiometabolic phenotypes demonstrated that L-methionine-producing bacteria were associated with atherosclerosis in obese individuals. It is suggested that methionine causes this effect through its direct conversion to HCy, contributing to serum elevation, which is a known cardiovascular risk factor that will be addressed later (Kurilshikov *et al.*, 2019).

The presence of bacterial DNA in human plasma has been addressed in some reviews and associated with endotoxin levels to trigger systemic inflammation and increase the instability of atherosclerotic plaques. It binds to TLR-9, stimulating inflammatory intracellular signaling pathways such as mitogen-activated protein kinase (MAPK), NF- κ B, PI3-kinase, and Jun N-terminal kinase (El Kebir *et al.*, 2008; Szeto *et al.*, 2018a).

The intestinal microbiota is also related to the trimethylamine/N-trimethylamide (TMA/TMAO) pathway (Hansen *et al.*, 2015; Wang and Zhao, 2018). *Proteus mirabilis* is a gram-negative bacterium, a component of the fecal microbiota, but in a dysbiotic environment, it is found in higher numbers. These species, from the carbon extraction of some compounds such as choline, phosphatidylcholine, glycerol phosphocholine, carnitine, betaine, and γ -butyrobetaine, produce the metabolite TMA by TMA lyases (Tang and Hazen, 2014; Wang *et al.*, 2015; Wang and Zhao, 2018). TMA is oxidized to TMAO in the liver by flavin monooxygenase and transported to the systemic circulation (Bennett *et al.*, 2013; Koeth *et al.*, 2013; Tang and Hazen, 2014; Wang and Zhao, 2018) and activates smooth muscle cells of the vessels, endothelial MAPK and the NF- κ B pathway, leading to the expression of pro-inflammatory cytokines and leukocyte adhesion in addition to inducing transformation of macrophages to foam cells to activate NOD (nucleotide-binding oligomerization domain), LRR (leucine-rich repeats) and NLRP3 (nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3) (Chen *et al.*, 2017; Seldin *et al.*, 2016; Wang and Zhao, 2018; Wei *et al.*, 2017). Additionally, TMAO reduces the expression of the genes CYP7A1 (cytochrome P450 family 7 subfamily A member 1) and CYP27A1 (cytochrome P450 family 27 subfamily A member 1), affecting the metabolism of cholesterol and bile salts and the release of calcium in the platelet endoplasmic reticulum, facilitating the formation of thrombi (Koeth *et al.*, 2013; Zhu *et al.*, 2016).

Furthermore, hyperhomocysteinaemia is an independent cardiovascular risk factor and has been observed in both patients with UC and CD at concentrations 4–5 times higher than those in healthy individuals (Drzewoski *et al.*, 2006; Oussalah *et al.*, 2011; Tyagi *et al.*, 2005; Wu *et al.*, 2017).

High levels of HCy cause nitroxidative stress and endothelial dysfunction through self-oxidation catalysed by cationic metals, which results in the formation of O₂⁻ and, when reacting with *NO, forms (ONOO⁻), which reduces

the bioavailability of *NO (Sen *et al.*, 2010; Tyagi *et al.*, 2005). Studies in cell culture (Liu *et al.*, 2013; Zhu *et al.*, 2016) and animals (Sen *et al.*, 2010) have shown that high levels of HCy induce RONS formation, causing an increase in the concentrations of H₂O₂ and MDA and mitochondrial damage by decreasing the expression of cytochrome c oxidase III/ATPase 6/8 and heat shock protein 60 and impairing the antioxidant defence system (GPx, hemeoxygenase and SOD), thus leading to redox imbalance (Liu *et al.*, 2013; Tyagi *et al.*, 2005). It also activates the NFκB pathway, adhesion molecules (ICAM-1, VCAM-1) and MCP-1. These, in turn, interact with inflammatory cells of the endothelium, leading to the atherosclerotic process (Silverman *et al.*, 2002; Wang *et al.*, 2002).

Neurological manifestations

Malabsorption, nutritional deficiencies (vitamins B1, B12, D and E, folic acid and nicotinamide), infections, thromboembolism, adverse drug effects (metronidazole, sulfasalazine, steroids, cyclosporine), and immunological abnormalities are related to changes in the bowel-brain axis (Casella *et al.*, 2014; Moris, 2014). The pathophysiology of neurological manifestations of IBD and their prevalence are not yet fully understood. Neurologic and neuromuscular complications in IBD (peripheral neuropathies, multiple sclerosis, cerebrovascular diseases, and others including psychiatric disorders) have been estimated from 0.25 to 35.7% due to the different forms of diagnosis (Elsehty and Bertorini, 1997; Gondim *et al.*, 2005; Lossos *et al.*, 1995).

A link has been proposed between chronic intestinal inflammation and neuropathy onset. The deregulated immune response involving mainly T cells towards the autoantigens that crossed the blood-brain barrier (BBB) induces central nervous system activation of astrocytes and microglia, which stimulates the production of pro-inflammatory cytokines, RONS, glutamatergic excitotoxicity, and autoantibody generation against the myelin sheath, causing axonal injury and neuronal dysfunction (Casella *et al.*, 2014; Moris, 2014; Singh *et al.*, 2013). According to Nemati *et al.* (2019), the development of neurological disease in IBD may be coincidental (chance association) or the consequence of the primary disease and this differentiation is necessary because they may require different treatments. Furthermore, investigations in this context are not yet routine in clinical practice.

An intimate link between the microbiota-gut-brain axis has been proposed. The microbiota influences several brain activities, including the modulation of neuroimmune responses and maintenance of the integrity of the epithelial barrier and the BBB, through the expression of occludin and claudin, demonstrated in an animal model (Braniste *et al.*, 2014). Changes in the microbiota and bowel permeability allow the spread of bacteria, toxins such as LPS, and other metabolites (TMAO, for example) towards the BBB. This causes an unregulated immune response, with activation of the microglia that alters its secretory profile and stimulates the production of pro-inflammatory chemokines and cytokines (MCP-1, interferon-gamma [IFN-γ], IL-6, IL-8, and TNF-α), as well as increases RONS (Block *et al.*, 2007; Del Rio *et al.*, 2017; Shemer *et al.*, 2015).

Multiple sclerosis (MS), characterized by demyelination and neuroinflammation, has been closely related to IBD.

Studies indicate that first-degree relatives of patients with MS are at risk for CD development (risk of 1.4), while IBD patients have a risk of 1.7 of developing MS (Gupta *et al.*, 2005; Nielsen *et al.*, 2008; Noseworthy *et al.*, 2000). Changes in the microbiota in patients with MS have been identified in this process, such as an increase in antibodies against intestinal microbial components (Banati *et al.*, 2013). Furthermore, in cell culture, the activation of mucosal-associated invariant T cells (MAIT), seen in IBD, also plays a role in MS, as they can interact with microbial components and stimulate the innate and adaptive response, including CD8+ T cells and the production of pro-inflammatory cytokines, such as IL-17. Moreover, these cells express high levels of CD161 (cluster of differentiation 161), CCR6 (chemokine receptor 6), and IL-18 (Moreira *et al.*, 2017).

Neurodegenerative diseases, such as Parkinson's disease (PD), have also been associated with intestinal dysbiosis and increased intestinal permeability, seen in IBD. Animal studies have shown that the increase in LPS in UC causes an overactivation of microglia, TNF-α secretion and iNOS activation. The identification of α-synuclein (Parkinson's disease-PD-marker protein) in the enteric nervous system and later in the neurons of the central nervous system (CNS) reinforced the link (de La Serre *et al.*, 2015; Hoban *et al.*, 2013; Qin *et al.*, 2013; Villaran *et al.*, 2010).

In humans, this relationship has also been confirmed. According to Villumsen *et al.* (2019), patients with IBD have up to 22% risk of developing PD when compared to healthy individuals.

The reduced integrity of the BBB in model animals and PD individuals has been reported, and it is possible that concomitant epithelial barrier dysfunction increases microbiota communication with CNS cells (Gray and Woulfe, 2015; Zhao *et al.*, 2007).

On the other hand, studies have shown that patients with IBD have a 2–3 times higher risk of developing anxiety and depression than the general population, affecting 30% and 25%, respectively, of these patients, especially in the active phase (Mikocka-Walus *et al.*, 2016; Walker *et al.*, 2008).

The intimate connection and communication of the microbiota-gut-brain axis is proposed to explain this prevalence. The hypothalamic-pituitary-adrenal axis perceives and responds to the stimulus of stress and inflammation, leading to a decrease in the effector response of the vagus nerve and an increase in intestinal permeability. This favors bacterial translocation that activates the mucosal immune response, as well as other cells of the innate immune system in the brain, increasing microglia activity, pro-inflammatory cytokine levels and nitroxidative stress, which generate changes in brain functions and damage to neuroplasticity (Abautret-Daly *et al.*, 2018; Bailey *et al.*, 2006; Maes *et al.*, 2012; Santos *et al.*, 1999).

In cell culture and animal model assays, dysbiosis impairs its appropriate interaction with the host and influences neural activities in brain areas related to stress and behavior, with changes in brain-derived neurotrophic factor (BDNF) in the hypothalamus and amygdala, in addition to the decrease in SCFA-producing bacteria, which, in turn, are immunomodulatory metabolites. The participation of the microbiota seems to be related to the

gamma-aminobutyric acid (GABA), serotonin and dopamine pathways, which are implicated in depressive and anxiety disorders (Bernstein, 2017; Foster and McVey Neufeld, 2013; Lyte et al., 2011).

Renal manifestations

The link between IBD and renal manifestations, although less frequent, has been demonstrated in both CD and UC. The reported diseases are nephrolithiasis, glomerulonephritis, and tubule-interstitial nephritis (Ambruzs et al., 2014; Corica and Romano, 2016).

Nephrolithiasis affects 12–28% of patients with IBD, and few studies report a 9–18% higher risk than in the general population. The frequency is higher in CD and patients submitted to surgical procedures such as colectomy with ileostomy and intestinal resection (Bianchi et al., 2018; Gkentzis et al., 2016; McConnell et al., 2002; Parks et al., 2003). In this context, a small cohort of 83 patients demonstrated an incidence of 24% IgA glomerulonephritis and 19% tubule-interstitial nephritis, especially in CD (Ambruzs et al., 2014).

Some links between renal manifestations and IBD have been described that seem to have an important role, including genetics by identifying the HLA-DR1 (human leukocyte antigen-DR1) and HLA-DR1/DQw5 genes present in both IgA glomerulonephritis and CD; adverse effects of drugs such as aminosalicylates and its derivatives, especially in tubule-interstitial nephritis; and a reduction in anti-lithogenic factors (citrate and magnesium) in nephrolithiasis, mainly due to chronic diarrhea (Ambruzs et al., 2014; Ganji-Arjenaki et al., 2017; Hueppelshaeuser et al., 2012; Kane, 2006; Oikonomou et al., 2011; Takemura et al., 2002; Worcester, 2002). However, these factors cannot fully explain the renal manifestations in patients with IBD.

In relation to nephrolithiasis, intestinal dysbiosis seems to be involved in the formation of calcium oxalate stones in the kidneys. It has been observed that individuals with IBD, who have a smaller population of *Oxalobacter formigenes* in their intestinal microbiota, have a high prevalence of nephrolithiasis (Kumar et al., 2004). *O. formigenes* is a gram-negative, anaerobic, commensal bacterium from the gastrointestinal tract that acts on the regulation of oxalate homeostasis; it degrades oxalate through the enzymes oxalyl-coenzyme A decarboxylase and formyl-coenzyme transferase and interacts with the intestinal mucosa, stimulating the secretion of endogenous oxalate into the lumen, a transport mechanism through the epithelium, thus contributing to the excretion of this compound (Arvans et al., 2017; Liu et al., 2017; Siener et al., 2013; Siva et al., 2009; Stewart et al., 2004).

It has been observed, especially in patients with CD, that decolonization of TGI by *O. formigenes* leads to a reduction in the intestinal catabolism of oxalate and the subsequent appearance of hyperoxaluria, the main mechanism involved in the formation of kidney stones. In addition, an increase in urinary oxalate was observed in these patients when compared to healthy controls. It occurs more frequently in individuals who have undergone ileum resection and colon-jejunal anastomoses (Hueppelshaeuser et al., 2012; Kane, 2006; Kumar et al., 2004; Oikonomou et al., 2011; Siva et al., 2009). The causes that lead to the decolonization of this

bacterium have not yet been elucidated. However, it has been proposed that the possible reasons are precisely associated with changes in the composition of the intestinal microbiota, typical of IBD, which is the result of an inflammatory response exacerbated against these commensals, which results in a break in immunological tolerance (Kumar et al., 2004).

Nevertheless, other commensal bacteria, such as *Lactobacillus* and *Bifidobacterium*, also secrete enzymes capable of degrading oxalate in a medium that contains glucose and lactose (Campieri et al., 2001). The role of oxalate-degrading bacteria in the treatment of kidney stones has become of great interest in the scientific community, aiming at their use as a probiotic. Animal and human research using *O. formigenes* or their enzymes have shown promising results; however, data security is still required through well-controlled and larger-scale clinical trials. Other studies investigating the species *Lactobacillus* and *Bifidobacterium* have also shown a reduction in oxalate excretion, but the results are still controversial. The importance of additional standardized studies that determine the direct relationship between the decolonization of these bacteria and the presence of hyperoxaluria and consequent kidney stones and their influence as a risk factor for the identification of possible probiotics in the treatment of this condition is emphasized. It is said that functional and molecular approaches are needed to choose the best species that also have the ability to effectively colonize the intestine (Abratt and Reid, 2010; Mehta et al., 2016; Sadaf et al., 2017).

Oxalate generates toxic responses that result in the activation of phospholipase A2, which culminates in the production of arachidonic acid and various lysophospholipids. These factors, in turn, lead to mitochondrial dysfunction, an increase of RONS production, and the induction of changes in the expression of genes involved in the synthesis of molecules that inhibit the formation of calcium oxalate stones and in the activation of caspases, which are involved in apoptotic cell death (Cao et al., 2000; Cao et al., 2004; Jonassen et al., 2005; Oikonomou et al., 2011). In addition, the presence of TLRs, especially TLR4, expressed in renal epithelial cells and the identification of significant amounts of endotoxins in kidney stones suggest that this interaction generates activation of inflammatory pathways (Anders et al., 2004a; Anders et al., 2004b; McAleer et al., 2003; Oikonomou et al., 2011).

In IgA nephropathy, the intimate relationship of the microbiota-intestine-kidney axis has been implicated. The breakdown of the intestinal barrier and the increase in pro-inflammatory cytokines leads to bacterial translocation, facilitating the diffusion of microbial endotoxins and DNA through the bloodstream, which become established in the renal glomeruli. It was proposed that the deposited immune complexes react with luminal antigens and that there is a loss of exclusion and antigenic tolerance, sustained immune response and deregulation in the production and transport of IgA. Evidence shows that patients with CD present high levels of IgG and IgA due to an inflammatory response of the mucosa to *K. pneumoniae* (Anders et al., 2004a; Corica and Romano, 2016; Forshaw et al., 2005; O'Mahony et al., 1992; Takemura et al., 2002).

Finally, in tubule-interstitial nephritis, a deregulated immune and inflammatory response, the presence of

autoantibodies and immunocomplexes against epitopes common to the intestine and kidney and molecular mimicry are likely pathophysiological mechanisms (Ambruzs *et al.*, 2014; Corica and Romano, 2016; Fraser *et al.*, 2001; Mahmud *et al.*, 2002; Oikonomou *et al.*, 2011; Poulou *et al.*, 2006).

The diseases addressed may progress to renal failure and end-stage renal disease if the inflammation persists and/or is not contained. A few epidemiological studies have reported the incidence of renal failure between 2 and 15.9% in patients with IBD, and there is an elevated risk for end-stage renal disease in CD that is five times higher than that in CU and healthy controls, suggesting that IBD, especially CD, is an independent risk factor (Park *et al.*, 2018; Primas *et al.*, 2013). Some mechanisms have been proposed to explain the increased risk in patients with CD, including systemic inflammation resulting from the unregulated immune response in the intestine and the transmural character that can exacerbate renal damage, as has been reported with the increase in serum levels of IL-6 and CRP, in addition to autoimmunity (Cioffi *et al.*, 2015; Fried *et al.*, 2004). Therefore, it is essential to monitor renal function in patients with IBD to identify early signs of renal damage and guide the choice of appropriate and effective therapy.

Study limitations

There are few prospective studies in humans that assess the microbiota and nitroxidative stress in EIM of IBD. In some studies, associated comorbidities, such as diabetes mellitus and arterial hypertension, were not mentioned as a possible confounding factor. Thus, in this review, we propose a crosslink between redox imbalance and dysbiosis, using mainly *in vitro* and experimental reports, as well as physiological and biochemical established routes.

Prospects and future direction

Evidence suggests that chronic continuous nitroxidative stress and alteration of the intestinal microbiota may be strongly associated with the appearance of EIM inherent to patients with IBD (Fig. 3). Despite the involvement of genetic factors, it is necessary that other variables act to stimulate and/or to facilitate the expression of this phenotype, and the chronic inflammatory environment associated with increased intestinal permeability becomes decisive in determining these manifestations of the disease.

However, the studies carried out in humans, herein described, demonstrate the association and the presence of dysbiosis in the hepatic (PSC), osteoarticular, and dermatological manifestations (psoriasis) and as a contributing factor for the formation of kidney stones. It is associated with the nitroxidative stress and lipid peroxidation in triggering NAFLD and endothelial dysfunction described in the pathophysiology of thromboembolic events. Nevertheless, according to the evidence presented, it is possible that an imbalance in the intestinal microbiota and nitroxidative stress are also implicated in cardiac, ophthalmological, and neurological outcomes, including psychological disorders. However, there are still gaps to explore in regard to the mechanisms and pathways that culminate in such conditions.

The absence of standardized clinical studies and the scarcity of large, prospective studies make it impossible to establish a cause-and-effect link, hindering a clinical intervention in this dimension. In view of this, well-designed clinical studies are necessary to further investigate the influence of nitroxidative stress and dysbiosis, as well as other factors mentioned in this work, such as the current pharmacological therapy used to trigger EIM. The understanding of the intrinsic pathophysiological pathways is urgently required for the development of effective

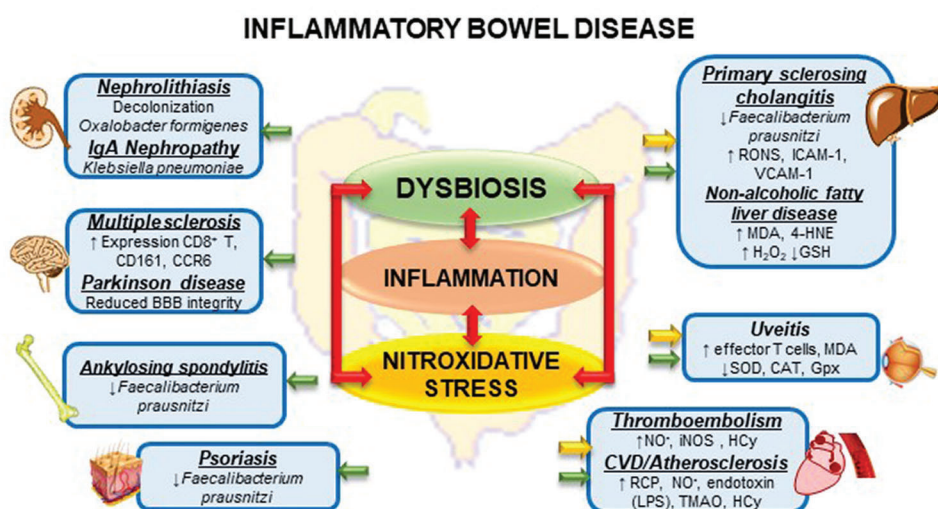


FIGURE 3. Interactions between dysbiosis, inflammation and nitroxidative stress and Extraintestinal Manifestation in inflammatory Bowel Disease. Legend: Green arrows: dysbiosis influence; Yellow arrows: nitroxidative stress influence; BBB: blood-brain barrier; CAT: catalase; CCR6: chemokine receptor 6; CD161: cluster of differentiations 161- lecithin-like receptor; CD8+ T: CD8+ T lymphocytes; CVD: cardiovascular disease; Hcy: homocysteine; GPx: glutathione peroxidase; GSH: reduced glutathione; ICAM-1: intercellular adhesion molecule 1; IgA: immunoglobulin A; LPS: lipopolysaccharide; MDA: malondialdehyde; *NO: nitric oxide; RCP: reactive C protein; RONS: reactive oxygen and nitrogen species; SOD: superoxide dismutase; TMAO: N-trimethylamine-N-oxide; VCAM-1: vascular cell adhesion molecule 1; 4-HNE: 4-hydroxynonenal; iNOS: inducible nitric oxide synthase.

preventive and therapeutic strategies aimed at manipulating the intestinal microbiota and controlling inflammation/nitroxidative stress.

Units: Units of measurement should be used concisely according to the International System of Units (SI). All units should be converted to SI units whenever possible.

Statistical Analysis: Appropriate statistical treatment of the data is essential. When statistical analysis is performed, the name of the statistical test used, the number for each analysis, the comparisons of interest, the alpha level and the actual *p*-value for each test should be provided.

Author Contribution: Amylly Sanuely da Paz Martins (amyllymartins@gmail.com): Analysis and interpretation of data, writing, preparation of figures, and graphical abstract. Samara Bomfim Gomes Campos (bomfim_samara@hotmail.com): Acquisition of data, library search. Marília Oliveira Fonseca Goulart (mofg@qui.ufal.br): Critical evaluation of the content and contribution in conclusions and perspectives. Fabiana Andrea Moura (fabianamoura_al@hotmail.com): Supervised all the steps, conception and design, figures and critical revision.

Funding Statement: This work was supported by grants from the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq–Brazil) [435704/2018-4] and Fundação de Amparo à Pesquisa do Estado de Alagoas (FAPEAL)/PPSUS/Ministério da Saúde (MS) [60030-000876/2016].

Conflicts of Interest: The authors declare that they have no conflicts of interest to report regarding the present study.

References

- Abautret-Daly A, Dempsey E, Parra-Blanco A, Medina C, Harkin A (2018). Gut–brain actions underlying comorbid anxiety and depression associated with inflammatory bowel disease. *Acta Neuropsychiatrica* **30**: 275–296. DOI 10.1017/neu.2017.3.
- Abratt VR, Reid SJ (2010). Oxalate-degrading bacteria of the human gut as probiotics in the management of kidney stone disease. *Advances in Applied Microbiology* **72**: 63–87.
- Adams DH, Eksteen B (2006). Aberrant homing of mucosal T cells and extra-intestinal manifestations of inflammatory bowel disease. *Nature Reviews Immunology* **6**: 244–251. DOI 10.1038/nri1784.
- Ahn C, Negus D, Huang W (2018). Pyoderma gangrenosum: A review of pathogenesis and treatment. *Expert Review of Clinical Immunology* **14**: 225–233. DOI 10.1080/1744666X.2018.1438269.
- Ambruzs JM, Walker PD, Larsen CP (2014). The histopathologic spectrum of kidney biopsies in patients with inflammatory bowel disease. *Clinical Journal of the American Society of Nephrology* **9**: 265–270. DOI 10.2215/CJN.04660513.
- Anders HJ, Banas B, Schlöndorff D (2004a). Signaling danger: Toll-like receptors and their potential roles in kidney disease. *Journal of the American Society of Nephrology* **15**: 854–867. DOI 10.1097/01.ASN.0000121781.89599.16.
- Anders HJ, Vielhauer V, Eis V, Linde Y, Kretzler M, Perez de Lema G, Schlöndorff D (2004b). Activation of toll-like receptor-9 induces progression of renal disease in MRL-Fas(lpr) mice. *FASEB Journal* **18**: 534–536. DOI 10.1096/fj.03-0646fje.
- Aniwan S, Pardi DS, Tremaine WJ, Loftus EV Jr (2018). Increased risk of acute myocardial infarction and heart failure in patients with inflammatory bowel diseases. *Clinical Gastroenterology and Hepatology* **16**: 1607–1615.e1. DOI 10.1016/j.cgh.2018.04.031.
- Annese V (2019). A review of extraintestinal manifestations and complications of inflammatory bowel disease. *Saudi Journal of Medicine and Medical Sciences* **7**: 66–73. DOI 10.4103/sjms.sjms_81_18.
- Arvans D, Jung YC, Antonopoulos D, Koval J, Granja I, Bashir M, Hassan H (2017). *Oxalobacter formigenes*-derived bioactive factors stimulate oxalate transport by intestinal epithelial cells. *Journal of the American Society of Nephrology* **28**: 876–887. DOI 10.1681/ASN.2016020132.
- Arvikar SL, Fisher MC (2011). Inflammatory bowel disease associated arthropathy. *Current Reviews in Musculoskeletal Medicine* **4**: 123–131. DOI 10.1007/s12178-011-9085-8.
- Asquith M, Elewaut D, Lin P, Rosenbaum JT (2014). The role of the gut and microbes in the pathogenesis of spondyloarthritis. *Best Practice & Research in Clinical Rheumatology* **28**: 687–702. DOI 10.1016/j.berh.2014.10.018.
- Bailey MT, Engler H, Sheridan JF (2006). Stress induces the translocation of cutaneous and gastrointestinal microflora to secondary lymphoid organs of C57BL/6 mice. *Journal of Neuroimmunology* **171**: 29–37. DOI 10.1016/j.jneuroim.2005.09.008.
- Bajer L, Kverka M, Kostovcik M, Macinga P, Dvorak J, Stehlikova Z, Drastich P (2017). Distinct gut microbiota profiles in patients with primary sclerosing cholangitis and ulcerative colitis. *World Journal of Gastroenterology* **23**: 4548–4558. DOI 10.3748/wjg.v23.i25.4548.
- Banati M, Csecsei P, Koszegi E, Nielsen HH, Suto G, Bors L, Illes Z (2013). Antibody response against gastrointestinal antigens in demyelinating diseases of the central nervous system. *European Journal of Neurology* **20**: 1492–1495.
- Beckman JS, Koppenol WH (1996). Nitric oxide, superoxide, and peroxynitrite: The good, the bad, and ugly. *American Journal of Physiology-Cell Physiology* **271**: C1424–C1437. DOI 10.1152/ajpcell.1996.271.5.C1424.
- Bellanti F, Villani R, Facciorusso A, Vendemiale G, Serviddio G (2017). Lipid oxidation products in the pathogenesis of non-alcoholic steatohepatitis. *Free Radical Biology and Medicine* **111**: 173–185. DOI 10.1016/j.freeradbiomed.2017.01.023.
- Bennett BJ, de Aguiar Vallim TQ, Wang Z, Shih DM, Meng Y, Gregory J, Lusis AJ (2013). Trimethylamine-N-oxide, a metabolite associated with atherosclerosis, exhibits complex genetic and dietary regulation. *Cell Metabolism* **17**: 49–60. DOI 10.1016/j.cmet.2012.12.011.
- Benstoem C, Goetzenich A, Kraemer S, Borosch S, Manzanares W, Hardy G, Stoppe C (2015). Selenium and its supplementation in cardiovascular disease—what do we know? *Nutrients* **7**: 3094–3118. DOI 10.3390/nu7053094.
- Bernstein CN (2017). The brain-gut axis and stress in inflammatory bowel disease. *Gastroenterology Clinics of North America* **46**: 839–846. DOI 10.1016/j.gtc.2017.08.006.
- Bianchi L, Gaiani F, Vincenzi F, Kayali S, Di Mario F, Leandro G, Ruberto C (2018). Hemolytic uremic syndrome: Differential diagnosis with the onset of inflammatory bowel diseases. *Acta Biomedica* **89**: 153–157.
- Bigeh A, Sanchez A, Maestas C, Gulati M (2019). Inflammatory bowel disease and the risk for cardiovascular disease: Does all inflammation lead to heart disease? *Trends in*

- Cardiovascular Medicine* **30**: 463–469. DOI 10.1016/j.tcm.2019.10.001.
- Binion DG, Fu S, Ramanujam KS, Chai YC, Dweik RA, Drazba JA, Wilson KT (1998). iNOS expression in human intestinal microvascular endothelial cells inhibits leukocyte adhesion. *American Journal of Physiology* **275**: G592–G603.
- Binion DG, Rafiee P, Ramanujam KS, Fu S, Fisher PJ, Rivera MT, Wilson KT (2000). Deficient iNOS in inflammatory bowel disease intestinal microvascular endothelial cells results in increased leukocyte adhesion. *Free Radical Biology and Medicine* **29**: 881–888. DOI 10.1016/S0891-5849(00)00391-9.
- Block ML, Zecca L, Hong JS (2007). Microglia-mediated neurotoxicity: Uncovering the molecular mechanisms. *Nature Reviews Neuroscience* **8**: 57–69. DOI 10.1038/nrn2038.
- Brakenhoff LK, van der Heijde DM, Hommes DW, Huizinga TW, Fidder HH (2010). The joint–gut axis in inflammatory bowel diseases. *Journal of Crohn's and Colitis* **4**: 257–268. DOI 10.1016/j.crohns.2009.11.005.
- Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Toth M, Korecka A, Bakocevic N, Guan Ng L, Kundu G, Gulyás B, Hallidin C, Hultenby K, Nilsson H, Hebert H, Volpe BT, Diamond B, Pettersson S (2014). The gut microbiota influences blood-brain barrier permeability in mice. *Science Translational Medicine* **6**: 263ra158. DOI 10.1126/scitranslmed.3009759.
- Campieri C, Campieri M, Bertuzzi V, Swennen E, Matteuzzi D, Stefoni S, Pirovano F, Centi C, Ulisse S, Famularo G, De Simone C (2001). Reduction of oxaluria after an oral course of lactic acid bacteria at high concentration. *Kidney International* **60**: 1097–1105. DOI 10.1046/j.1523-1755.2001.0600031097.x.
- Cao LC, Honeyman T, Jonassen J, Scheid C (2000). Oxalate-induced ceramide accumulation in Madin-Darby canine kidney and LLC-PK1 cells. *Kidney International* **57**: 2403–2411. DOI 10.1046/j.1523-1755.2000.00099.x.
- Cao LC, Honeyman TW, Cooney R, Kennington L, Scheid CR, Jonassen JA (2004). Mitochondrial dysfunction is a primary event in renal cell oxalate toxicity. *Kidney International* **66**: 1890–1900. DOI 10.1111/j.1523-1755.2004.00963.x.
- Capon F, Di Meglio P, Szaub J, Prescott NJ, Dunster C, Baumber L, Nestle FO (2007). Sequence variants in the genes for the interleukin-23 receptor (IL23R) and its ligand (IL12B) confer protection against psoriasis. *Human Genetics* **122**: 201–206. DOI 10.1007/s00439-007-0397-0.
- Casella G, Tontini GE, Bassotti G, Pastorelli L, Villanacci V, Spina L, Vecchi M (2014). Neurological disorders and inflammatory bowel diseases. *World Journal of Gastroenterology* **20**: 8764–8782.
- Castro Aguilar-Tablada T, Navarro-Alarcon M, Quesada Granados J, Samaniego Sanchez C, Rufian-Henares JA, Noguera-Lopez F (2016). Ulcerative colitis and crohn's disease are associated with decreased serum selenium concentrations and increased cardiovascular risk. *Nutrients* **8**: 12. DOI 10.3390/nu8120780.
- Chakravarthy KS, Jayasudha R, Sai Prashanthi G, Ali MH, Sharma S, Tyagi M, Shivaji S (2018). Dysbiosis in the gut bacterial microbiome of patients with uveitis, an inflammatory disease of the eye. *Indian Journal of Microbiology* **58**: 457–469. DOI 10.1007/s12088-018-0746-9.
- Chao CY, Battat R, Al Khoury A, Restellini S, Sebastiani G, Bessissow T (2016). Co-existence of non-alcoholic fatty liver disease and inflammatory bowel disease: A review article. *World Journal of Gastroenterology* **22**: 7727–7734. DOI 10.3748/wjg.v22.i34.7727.
- Chapman R, Fevery J, Kallou A, Nagorney DM, Boberg KM, Shneider B (2010). Diagnosis and management of primary sclerosing cholangitis. *American Association for the Study of Liver* **51**: 660–678.
- Chen ML, Zhu XH, Ran L, Lang HD, Yi L, Mi MT (2017). Trimethylamine-N-oxide induces vascular inflammation by activating the NLRP3 inflammasome through the SIRT3-SOD2-mtROS signaling pathway. *Journal of the American Heart Association* **6**: 9.
- Chen S, Li Y, Li S, Yu C (2008). A Val227Ala substitution in the peroxisome proliferator activated receptor alpha (PPAR alpha) gene associated with non-alcoholic fatty liver disease and decreased waist circumference and waist-to-hip ratio. *Journal of Gastroenterology and Hepatology* **23**: 1415–1418. DOI 10.1111/j.1440-1746.2008.05523.x.
- Chen X, Zhang Z, Li H, Zhao J, Wei X, Lin W, Zhao X, Jiang A, Yuan J (2020). Endogenous ethanol produced by intestinal bacteria induces mitochondrial dysfunction in non-alcoholic fatty liver disease. *Journal of Gastroenterology and Hepatology* **35**: 2009–2019. DOI 10.1111/jgh.15027.
- Cho JH (2008). The genetics and immunopathogenesis of inflammatory bowel disease. *Nature Reviews Immunology* **8**: 458–466. DOI 10.1038/nri2340.
- Ciccacci C, Biancone L, Di Fusco D, Ranieri M, Condino G, Giardina E, Onali S, Lepre T, Pallone F, Novelli G, Borgiani P (2013). TRAF3IP2 gene is associated with cutaneous extraintestinal manifestations in inflammatory bowel disease. *Journal of Crohn's and Colitis* **7**: 44–52. DOI 10.1016/j.crohns.2012.02.020.
- Ciccica F, Ferrante A, Triolo G (2016). Intestinal dysbiosis and innate immune responses in axial spondyloarthritis. *Current Opinion in Rheumatology* **28**: 352–358. DOI 10.1097/BOR.0000000000000296.
- Cioffi M, Rosa AD, Serao R, Picone I, Vietri MT (2015). Laboratory markers in ulcerative colitis: Current insights and future advances. *World Journal of Gastrointestinal Pathophysiology* **6**: 13–22. DOI 10.4291/wjgp.v6.i1.13.
- Corica D, Romano C (2016). Renal involvement in inflammatory bowel diseases. *Journal of Crohn's and Colitis* **10**: 226–235. DOI 10.1093/ecco-jcc/jjv138.
- Costello ME, Ciccica F, Willner D, Warrington N, Robinson PC, Gardiner B, Marshall M, Kenna TJ, Triolo G, Brown MA (2015). Brief report: Intestinal dysbiosis in ankylosing spondylitis. *Arthritis & Rheumatology* **67**: 686–691. DOI 10.1002/art.38967.
- Cottone M, Sapienza C, Macaluso FS, Cannizzaro M (2019). Psoriasis and inflammatory bowel disease. *Digestive Diseases* **37**: 451–457. DOI 10.1159/000500116.
- Danese S (2007). Inflammation and the mucosal microcirculation in inflammatory bowel disease: The ebb and flow. *Current Opinion in Gastroenterology* **23**: 384–389. DOI 10.1097/MOG.0b013e32810c8de3.
- Danese S, Papa A, Saibeni S, Repici A, Malesci A, Vecchi M (2007). Inflammation and coagulation in inflammatory bowel disease: The clot thickens. *American Journal of Gastroenterology* **102**: 174–186. DOI 10.1111/j.1572-0241.2006.00943.x.
- Davis C, Fischer J, Ley K, Sarembock IJ (2003). The role of inflammation in vascular injury and repair. *Journal of Thrombosis and Haemostasis* **1**: 1699–1709. DOI 10.1046/j.1538-7836.2003.00292.x.
- de La Serre CB, de Lartigue G, Raybould HE (2015). Chronic exposure to low dose bacterial lipopolysaccharide inhibits leptin signaling in vagal afferent neurons. *Physiology & Behavior* **139**: 188–194. DOI 10.1016/j.physbeh.2014.10.032.

- Del Rio D, Zimetti F, Caffarra P, Tassotti M, Bernini F, Brighenti F, Zini A, Zanotti I (2017). The gut microbial metabolite trimethylamine-N-oxide is present in human cerebrospinal fluid. *Nutrients* **9**: 10. DOI 10.3390/nu9101053.
- Demetter P, Baeten D, De Keyser F, De Vos M, Van Damme N, Verbruggen G, Vermeulen S, Mareel M, Elewaut G, Mielants H, Veys EM, Cuvelier CA (2000). Subclinical gut inflammation in spondyloarthritis patients is associated with upregulation of the E cadherin/catenin complex. *Annals of the Rheumatic Diseases* **59**: 211–216. DOI 10.1136/ard.59.3.211.
- Demetter P, De Vos M, Van Huysse JA, Baeten D, Ferdinande L, Peeters H, Mielants H, Veys EM, De Keyser F, Cuvelier CA (2005). Colon mucosa of patients both with spondyloarthritis and Crohn's disease is enriched with macrophages expressing the scavenger receptor CD163. *Annals of the Rheumatic Diseases* **64**: 321–324. DOI 10.1136/ard.2003.018382.
- Dijkstra G, Moshage H, van Dullemen HM, de Jager-Krikken A, Tiebosch AT, Kleibeuker GH, Jansen PL, van Goor H (1998). Expression of nitric oxide synthases and formation of nitrotyrosine and reactive oxygen species in inflammatory bowel disease. *Journal of Pathology* **186**: 416–421.
- Drzewoski J, Gasiorowska A, Malecka-Panas E, Bald E, Czupryniak L (2006). Plasma total homocysteine in the active stage of ulcerative colitis. *Journal of Gastroenterology and Hepatology* **21**: 739–743. DOI 10.1111/j.1440-1746.2006.04255.x.
- Eaton JE, Talwalkar JA (2013). Primary sclerosing cholangitis: Current and future management strategies. *Current Hepatitis Reports* **12**: 28–36. DOI 10.1007/s11901-012-0155-1.
- Eksteen B, Grant AJ, Miles A, Curbishley SM, Lalor PF, Hübscher SG, Briskin M, Salmon M, Adams DH (2004). Hepatic endothelial CCL25 mediates the recruitment of CCR9+ gut-homing lymphocytes to the liver in primary sclerosing cholangitis. *Journal of Experimental Medicine* **200**: 1511–1517. DOI 10.1084/jem.20041035.
- El Kebir D, Jozsef L, Filep JG (2008). Neutrophil recognition of bacterial DNA and Toll-like receptor 9-dependent and -independent regulation of neutrophil function. *Archivum Immunologiae et Therapiae Experimentalis* **56**: 41–53. DOI 10.1007/s00005-008-0008-3.
- Ellinghaus D, Ellinghaus E, Nair RP, Stuart PE, Esko T, Metspalu A, Debrus S, Raelson JV, Tejasvi T, Belouchi M, West SL, Barker JN, Köks S, Kingo KC, Balschun T, Palmieri O, Annese V, Gieger C, Wichmann HE, Kabesch M, Trembath RC, Mathew CG, Abecasis GR, Weidinger S, Nikolaus S, Schreiber S, Elder JT, Weichenthal M, Nothnagel M, Franke A (2012). Combined analysis of genome-wide association studies for Crohn disease and psoriasis identifies seven shared susceptibility loci. *American Journal of Human Genetics* **90**: 636–647. DOI 10.1016/j.ajhg.2012.02.020.
- Elsehty A, Bertorini TE (1997). Neurologic and neuropsychiatric complications of Crohn's disease. *Southern Medical Journal* **90**: 606–610. DOI 10.1097/00007611-199706000-00005.
- Eppinga H, Sperna Weiland CJ, Thio HB, van der Woude CJ, Nijsten TE, Peppelenbosch MP, Konstantinov SR (2016). Similar depletion of protective *Faecalibacterium prausnitzii* in psoriasis and inflammatory bowel disease, but not in hidradenitis suppurativa. *Journal of Crohn's and Colitis* **10**: 1067–1075. DOI 10.1093/ecco-jcc/jjw070.
- Esmon CT (2005). The interactions between inflammation and coagulation. *British Journal of Haematology* **131**: 417–430. DOI 10.1111/j.1365-2141.2005.05753.x.
- Fantini MC, Pallone F, Monteleone G (2009). Common immunologic mechanisms in inflammatory bowel disease and spondylarthropathies. *World Journal of Gastroenterology* **15**: 2472–2478. DOI 10.3748/wjg.15.2472.
- Farhi D, Cosnes J, Zizi N, Chosidow O, Seksik P, Beaugerie L, Aractingi S, Khosrotehrani K (2008). Significance of erythema nodosum and pyoderma gangrenosum in inflammatory bowel diseases: A cohort study of 2402 patients. *Medicine* **87**: 281–293. DOI 10.1097/MD.0b013e318187cc9c.
- Feuerstein JD, Cheifetz AS (2014). Ulcerative colitis: Epidemiology, diagnosis, and management. *Mayo Clinic Proceedings* **89**: 1553–1563. DOI 10.1016/j.mayocp.2014.07.002.
- Floege J, Ketteler M (2004). Vascular calcification in patients with end-stage renal disease. *Nephrology Dialysis Transplantation* **19**: V59–V66. DOI 10.1093/ndt/gfh1058.
- Forshaw MJ, Guirguis O, Hennigan TW (2005). IgA nephropathy in association with Crohn's disease. *International Journal of Colorectal Disease* **20**: 463–465. DOI 10.1007/s00384-004-0696-z.
- Foster JA, McVey Neufeld KA (2013). Gut-brain axis: How the microbiome influences anxiety and depression. *Trends in Neuroscience* **36**: 305–312. DOI 10.1016/j.tins.2013.01.005.
- Fousekis FS, Theopistos VI, Katsanos KH, Tsianos EV, Christodoulou DK (2018). Hepatobiliary manifestations and complications in inflammatory bowel disease: A review. *Gastroenterology Research* **11**: 83–94. DOI 10.14740/gr990w.
- Fragoulis GE, Liava C, Daoussis D, Akriviadis E, Garyfallos A, Dimitroulas T (2019). Inflammatory bowel diseases and spondyloarthropathies: From pathogenesis to treatment. *World Journal of Gastroenterology* **25**: 2162–2176. DOI 10.3748/wjg.v25.i18.2162.
- Fraser JS, Muller AF, Smith DJ, Newman DJ, Lamb EJ (2001). Renal tubular injury is present in acute inflammatory bowel disease prior to the introduction of drug therapy. *Alimentary Pharmacology & Therapeutics* **15**: 1131–1137. DOI 10.1046/j.1365-2036.2001.01041.x.
- Fried L, Solomon C, Shlipak M, Seliger S, Stehman-Breen C, Bleyer AJ, Chaves P, Furberg C, Kuller L, Newman A (2004). Inflammatory and prothrombotic markers and the progression of renal disease in elderly individuals. *Journal of the American Society of Nephrology* **15**: 3184–3191. DOI 10.1097/01.ASN.0000146422.45434.35.
- Fu Y, Lee CH, Chi CC (2018). Association of psoriasis with inflammatory bowel disease: A systematic review and meta-analysis. *JAMA Dermatology* **154**: 1417–1423. DOI 10.1001/jamadermatol.2018.3631.
- Fujimoto T, Imaeda H, Takahashi K, Kasumi E, Bamba S, Fujiyama Y, Andoh A (2013). Decreased abundance of *Faecalibacterium prausnitzii* in the gut microbiota of Crohn's disease. *Journal of Gastroenterology and Hepatology* **28**: 613–619. DOI 10.1111/jgh.12073.
- Ganji-Arjenaki M, Nasri H, Rafieian-Kopaei M (2017). Nephrolithiasis as a common urinary system manifestation of inflammatory bowel diseases; a clinical review and meta-analysis. *Journal of Nephropathology* **6**: 264–269. DOI 10.15171/jnp.2017.42.
- Garber A, Regueiro M (2019). Extraintestinal manifestations of inflammatory bowel disease: Epidemiology, etiopathogenesis, and management. *Current Gastroenterology Reports* **21**: 31. DOI 10.1007/s11894-019-0698-1.
- Gilis E, Mortier C, Venken K, Debusschere K, Vereecke L, Elewaut D (2018). The role of the microbiome in gut and joint

- inflammation in psoriatic arthritis and spondyloarthritis. *Journal of Rheumatology Supplement* **94**: 36–39.
- Gionchetti P, Calabrese C, Rizzello F (2015). Inflammatory Bowel Diseases and Spondyloarthropathies. *The Journal of Rheumatology Supplement* **93**: 21–23. DOI 10.3899/jrheum.150628.
- Gkentzis A, Kimuli M, Cartledge J, Traxer O, Biyani CS (2016). Urolithiasis in inflammatory bowel disease and bariatric surgery. *World Journal of Nephrology* **5**: 538–546. DOI 10.5527/wjn.v5.i6.538.
- Gondim FA, Brannagan TH, III, Sander HW, Chin RL, Latov N (2005). Peripheral neuropathy in patients with inflammatory bowel disease. *Brain* **128**: 867–879. DOI 10.1093/brain/awh429.
- Grant AJ, Lalor PF, Salmi M, Jalkanen S, Adams DH (2002). Homing of mucosal lymphocytes to the liver in the pathogenesis of hepatic complications of inflammatory bowel disease. *Lancet* **359**: 150–157. DOI 10.1016/S0140-6736(02)07374-9.
- Gray MT, Woulfe JM (2015). Striatal blood–brain barrier permeability in Parkinson’s disease. *Journal of Cerebral Blood Flow & Metabolism* **35**: 747–750. DOI 10.1038/jcbfm.2015.32.
- Green S, Wahli W (1994). Peroxisome proliferator-activated receptors: Finding the orphan a home. *Molecular and Cellular Endocrinology* **100**: 149–153. DOI 10.1016/0303-7207(94)90294-1.
- Greuter T, Vavricka SR (2019). Extraintestinal manifestations in inflammatory bowel disease—epidemiology, genetics, and pathogenesis. *Expert Review of Gastroenterology & Hepatology* **13**: 307–317. DOI 10.1080/17474124.2019.1574569.
- Gupta G, Gelfand JM, Lewis JD (2005). Increased risk for demyelinating diseases in patients with inflammatory bowel disease. *Gastroenterology* **129**: 819–826. DOI 10.1053/j.gastro.2005.06.022.
- Hansen TH, Gobel RJ, Hansen T, Pedersen O (2015). The gut microbiome in cardio-metabolic health. *Genome Medicine* **7**: 33. DOI 10.1186/s13073-015-0157-z.
- Hansson GK (2005). Inflammation, atherosclerosis, and coronary artery disease. *New England Journal of Medicine* **353**: 429–430, author reply 429–430.
- Harbord M, Annese V, Vavricka SR, Allez M, Barreiro-de Acosta M, Boberg KM, Colitis O (2016). The first European evidence-based consensus on extra-intestinal manifestations in inflammatory bowel disease. *Journal of Crohn’s and Colitis* **10**: 239–254. DOI 10.1093/ecco-jcc/jjv213.
- Hart A L, Al-Hassi HO, Rigby RJ, Bell SJ, Emmanuel AV, Knight SC, Kamm MA, Stagg AJ (2005). Characteristics of intestinal dendritic cells in inflammatory bowel diseases. *Gastroenterology* **129**: 50–65. DOI 10.1053/j.gastro.2005.05.013.
- Hatoum OA, Binion DG, Otterson MF, Gutterman DD (2003). Acquired microvascular dysfunction in inflammatory bowel disease: Loss of nitric oxide-mediated vasodilation. *Gastroenterology* **125**: 58–69. DOI 10.1016/S0016-5085(03)00699-1.
- Hermann E, Yu DT, Meyer zum Buschenfelde KH, Fleischer B (1993). HLA-B27-restricted CD8 T cells derived from synovial fluids of patients with reactive arthritis and ankylosing spondylitis. *Lancet* **342**: 646–650. DOI 10.1016/0140-6736(93)91760-J.
- Higa A, Chevet E (2012). Redox signaling loops in the unfolded protein response. *Cellular Signalling* **24**: 1548–1555. DOI 10.1016/j.cellsig.2012.03.011.
- Hirschfield GM, Karlsen TH, Lindor KD, Adams DH (2013). Primary sclerosing cholangitis. *Lancet* **382**: 1587–1599. DOI 10.1016/S0140-6736(13)60096-3.
- Hoban DB, Connaughton E, Connaughton C, Hogan G, Thornton C, Mulcahy P, Moloney T C, Dowd E (2013). Further characterisation of the LPS model of Parkinson’s disease: A comparison of intra-nigral and intra-striatal lipopolysaccharide administration on motor function, microgliosis and nigrostriatal neurodegeneration in the rat. *Brain, Behavior, and Immunity* **27**: 91–100. DOI 10.1016/j.bbi.2012.10.001.
- Horai R, Caspi RR (2019). Microbiome and autoimmune uveitis. *Frontiers in Immunology* **10**: 232. DOI 10.3389/fimmu.2019.00232.
- Horai R, Zarate-Blades CR, Dillenburg-Pilla P, Chen J, Kielczewski JL, Silver PB, Jittayasothorn Y, Chan CC, Yamane H, Honda K, Caspi RR (2015). Microbiota-dependent activation of an autoreactive T cell receptor provokes autoimmunity in an immunologically privileged site. *Immunity* **43**: 343–353. DOI 10.1016/j.immuni.2015.07.014.
- Horowitz S, Binion DG, Nelson VM, Kanaa Y, Javadi P, Lazarova Z, Andrekopoulos C, Kalyanaraman B, Otterson MF, Rafiee P (2007). Increased arginase activity and endothelial dysfunction in human inflammatory bowel disease. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **292**: G1323–G1336. DOI 10.1152/ajpgi.00499.2006.
- Howell KW, Meng X, Fullerton DA, Jin C, Reece TB, Cleveland JC Jr (2011). Toll-like receptor 4 mediates oxidized LDL-induced macrophage differentiation to foam cells. *Journal of Surgical Research* **171**: e27–e31. DOI 10.1016/j.jss.2011.06.033.
- Huang X, Ye Z, Cao Q, Su G, Wang Q, Deng J, Zhou C, Kijlstra A, Yang P (2018). Gut microbiota composition and fecal metabolic phenotype in patients with acute anterior uveitis. *Investigative Ophthalmology & Visual Science* **59**: 1523–1531. DOI 10.1167/iovs.17-22677.
- Hudson M, Hutton RA, Wakefield AJ, Sawyerr AM, Pounder RE (1992). Evidence for activation of coagulation in Crohn’s disease. *Blood Coagulation & Fibrinolysis* **3**: 773–778. DOI 10.1097/00001721-199212000-00011.
- Hueppelshaeuser R, von Unruh GE, Habbig S, Beck BB, Buderus S, Hesse A, Hoppe B (2012). Enteric hyperoxaluria, recurrent urolithiasis, and systemic oxalosis in patients with Crohn’s disease. *Pediatric Nephrology* **27**: 1103–1109. DOI 10.1007/s00467-012-2126-8.
- Hussein IAH, Tohme R, Barada K, Mostafa MH, Freund JN, Jurjus RA, Karam W, Jurjus A (2008). Inflammatory bowel disease in rats: Bacterial and chemical interaction. *World Journal of Gastroenterology* **14**: 4028–4039. DOI 10.3748/wjg.14.4028.
- Jacques P, Elewaut D (2008). Joint expedition: Linking gut inflammation to arthritis. *Mucosal Immunology* **1**: 364–371. DOI 10.1038/mi.2008.24.
- Janowitz C, Nakamura YK, Metea C, Gligor A, Yu W, Karstens L, Rosenbaum JT, Asquith M, Lin P (2019). Disruption of intestinal homeostasis and intestinal microbiota during experimental autoimmune uveitis. *Investigative Ophthalmology & Visual Science* **60**: 420–429. DOI 10.1167/iovs.18-24813.
- Janse M, Lamberts LE, Franke L, Raychaudhuri S, Ellinghaus E, Muri Boberg K, Melum E, Folseraas T, Schrupf E, Bergquist A, Björnsson E, Fu J, Jan Westra H, Groen HJM, Fehrmann RSN, Smolonska J, van den Berg LH, Ophoff RA, Porte RJ, Weismüller TJ, Wedemeyer J, Schramm C, Sterneck M, Günther R, Braun F, Vermeire S, Henckaerts L, Wijmenga C,

- Ponsioen CY, Schreiber S, Karlsen TH, Franke A, Weersma RK (2011). Three ulcerative colitis susceptibility loci are associated with primary sclerosing cholangitis and indicate a role for IL2, REL, and CARD9. *Hepatology* **53**: 1977–1985. DOI 10.1002/hep.24307.
- Jonassen JA, Kohjimoto Y, Scheid CR, Schmidt M (2005). Oxalate toxicity in renal cells. *Urological Research* **33**: 329–339. DOI 10.1007/s00240-005-0485-3.
- Jorens PG, Hermans CR, Haber I, Kockx MM, Vermylen J, Parizel GA (1990). Acquired protein C and S deficiency, inflammatory bowel disease and cerebral arterial thrombosis. *Blut* **61**: 307–310. DOI 10.1007/BF01732883.
- Joseph L, Fink LM, Hauer-Jensen M (2002). Cytokines in coagulation and thrombosis: A preclinical and clinical review. *Blood Coagulation & Fibrinolysis* **13**: 105–116. DOI 10.1097/0001721-200203000-00005.
- Kamada N, Núñez G (2013). Role of the gut microbiota in the development and function of lymphoid cells. *Journal of Immunology* **190**: 1389–1395. DOI 10.4049/jimmunol.1203100.
- Kane S (2006). Urogenital complications of Crohn's disease. *American Journal of Gastroenterology* **101**: S640–S643. DOI 10.1111/j.1572-0241.2006.00662.x.
- Karlsen TH, Franke A, Melum E, Kaser A, Hov JR, Balschun T, Lie BA, Bergquist A, Schramm C, Weismüller TJ, Gotthardt D, Rust C, Philipp EER, Fritz T, Henckaerts L, Weersma RK, Stokkers P, Ponsioen CY, Wijmenga C, Sterneck M, Nothnagel M, Hampe J, Teufel A, Runz H, Rosenstiel P, Stiehl A, Vermeire S, Beuers U, Manns MP, Schrupf E, Boberg KM, Schreiber S (2010). Genome-wide association analysis in primary sclerosing cholangitis. *Gastroenterology* **138**: 1102–1111. DOI 10.1053/j.gastro.2009.11.046.
- Karremans MC, Luime JJ, Hazes JMW, Weel A (2017). The prevalence and incidence of axial and peripheral spondyloarthritis in inflammatory bowel disease: A systematic review and meta-analysis. *Journal of Crohn's and Colitis* **11**: 631–642.
- Kaur CP, Vadivelu J, Chandramathi S (2018). Impact of *Klebsiella pneumoniae* in lower gastrointestinal tract diseases. *Journal of Digestive Diseases* **19**: 262–271. DOI 10.1111/1751-2980.12595.
- Knutson CG, Mangerich A, Zeng Y, Raczynski AR, Liberman RG, Kang P, Ye W, Prestwich EG, Lu K, Wishnok JS, Korzenik JR, Wogan GN, Fox JG, Dedon PC, Tannenbaum SR (2013). Chemical and cytokine features of innate immunity characterize serum and tissue profiles in inflammatory bowel disease. *Proceedings of the National Academy of Sciences of the United States of America* **10**: 2332–2341. DOI 10.1073/pnas.1222669110.
- Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, Britt EB, Fu X, Wu Y, Li L, Smith JD, DiDonato JA, Chen J, Li H, Wu GD, Lewis JD, Warrier M, Brown JM, Krauss RM, Tang WHW, Bushman FD, Lusis AJ, Hazen SL (2013). Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nature Medicine* **19**: 576–585. DOI 10.1038/nm.3145.
- Kohjima M, Enjoji M, Higuchi N, Kato M, Kotoh K, Yoshimoto T, Fujino T, Yada M, Yada R, Harada N, Takayanagi R, Nakamura M (2007). Re-evaluation of fatty acid metabolism-related gene expression in nonalcoholic fatty liver disease. *International Journal of Molecular Medicine* **20**: 351–358.
- Kolios G, Valatas V, Ward SG (2004). Nitric oxide in inflammatory bowel disease: A universal messenger in an unsolved puzzle. *Immunology* **113**: 427–437. DOI 10.1111/j.1365-2567.2004.01984.x.
- Kumar R, Ghoshal UC, Singh G, Mittal RD (2004). Infrequency of colonization with *Oxalobacter formigenes* in inflammatory bowel disease: Possible role in renal stone formation. *Journal of Gastroenterology and Hepatology* **19**: 1403–1409. DOI 10.1111/j.1440-1746.2004.03510.x.
- Kurilshikov A, van den Munckhof ICL, Chen L, Bonder MJ, Schraa K, Rutten JHW, Riksen NP, de Graaf J, Oosting M, Sanna S, Joosten LAB, van der Graaf M, Brand T, Koonen DPY, van Faassen M, Xavier RJ, Kuipers F, Hofker MH, Wijmenga C, Netea MG, Zhernakova A, Fu J (2019). Gut microbial associations to plasma metabolites linked to cardiovascular phenotypes and risk. *Circulation Research* **124**: 1808–1820. DOI 10.1161/CIRCRESAHA.118.314642.
- Larsen S, Bendtzen K, Nielsen OH (2010). Extraintestinal manifestations of inflammatory bowel disease: Epidemiology, diagnosis, and management. *Annals of Medicine* **42**: 97–114. DOI 10.3109/07853890903559724.
- Lee IA, Kim DH (2011). *Klebsiella pneumoniae* increases the risk of inflammation and colitis in a murine model of intestinal bowel disease. *Scandinavian Journal of Gastroenterology* **46**: 684–693. DOI 10.3109/00365521.2011.560678.
- Leung TM, Nieto N (2013). CYP2E1 and oxidant stress in alcoholic and non-alcoholic fatty liver disease. *Journal of Hepatology* **58**: 395–398. DOI 10.1016/j.jhep.2012.08.018.
- Levi M, van der Poll T, Schultz M (2012). New insights into pathways that determine the link between infection and thrombosis. *Netherlands Journal of Medicine* **70**: 114–120.
- Liao L, Schneider KM, Galvez EJC, Frissen M, Marschall HU, Su H, Hatting M, Wahlström A, Haybaeck J, Puchas P, Mohs A, Peng J, Bergheim I, Nier A, Hennings J, Reißing J, Zimmermann HW, Longrich T, Strowig T, Liedtke C, Cubero FJ, Trautwein C (2019). Intestinal dysbiosis augments liver disease progression via NLRP3 in a murine model of primary sclerosing cholangitis. *Gut* **68**: 1477–1492. DOI 10.1136/gutjnl-2018-316670.
- Lin CH, Kadakia S, Frieri M (2014a). New insights into an autoimmune mechanism, pharmacological treatment and relationship between multiple sclerosis and inflammatory bowel disease. *Autoimmunity Reviews* **13**: 114–116. DOI 10.1016/j.autrev.2013.09.011.
- Lin P, Bach M, Asquith M, Lee AY, Akileswaran L, Stauffer P, Davin S, Pan Y, Cambonne ED, Dorris M, Debelius JW, Lauber CL, Ackermann G, Baeza YV, Gill T, Knight R, Colbert RA, Taurog JD, Van Gelder RN, Rosenbaum JT, Bereswill S (2014b). HLA-B27 and human β 2-microglobulin affect the gut microbiota of transgenic rats. *PLoS One* **9**: e105684. DOI 10.1371/journal.pone.0105684.
- Liu HH, Shih TS, Huang HR, Huang SC, Lee LH, Huang YC (2013). Plasma homocysteine is associated with increased oxidative stress and antioxidant enzyme activity in welders. *Scientific World Journal* **2013**: 370487. DOI 10.1155/2013/370487.
- Liu M, Koh H, Kurtz ZD, Battaglia T, PeBenito A, Li H, Nazzari L, Blaser MJ (2017). *Oxalobacter formigenes*-associated host features and microbial community structures examined using the American Gut Project. *Microbiome* **5**: 108. DOI 10.1186/s40168-017-0316-0.
- Lossos A, River Y, Eliakim A, Steiner I (1995). Neurologic aspects of inflammatory bowel disease. *Neurology* **45**: 416–421. DOI 10.1212/WNL.45.3.416.

- Lyte M, Vulchanova L, Brown DR (2011). Stress at the intestinal surface: Catecholamines and mucosa-bacteria interactions. *Cell and Tissue Research* **343**: 23–32. DOI 10.1007/s00441-010-1050-0.
- Maes M, Berk M, Goehler L, Song C, Anderson G, Galecki P, Leonard B (2012). Depression and sickness behavior are Janus-faced responses to shared inflammatory pathways. *BMC Medicine* **10**: 66. DOI 10.1186/1741-7015-10-66.
- Maggs JR, Chapman RW (2008). An update on primary sclerosing cholangitis. *Current Opinion in Gastroenterology* **24**: 377–383. DOI 10.1097/MOG.0b013e3282f9e239.
- Magri S, Paduano D, Chicco F, Cingolani A, Farris C, Delogu G, Tumbarello F, Lai M, Melis A, Casula L, Fantini MC, Usai P (2019). Nonalcoholic fatty liver disease in patients with inflammatory bowel disease: Beyond the natural history. *World Journal of Gastroenterology* **25**: 5676–5686. DOI 10.3748/wjg.v25.i37.5676.
- Mahmud N, O'Toole D, O'Hare N, Freyne PJ, Weir DG, Kelleher D (2002). Evaluation of renal function following treatment with 5-aminosalicylic acid derivatives in patients with ulcerative colitis. *Alimentary Pharmacology & Therapeutics* **16**: 207–215. DOI 10.1046/j.1365-2036.2002.01155.x.
- Makrides SC (1998). Therapeutic inhibition of the complement system. *Pharmacological Reviews* **50**: 59–87.
- Manichanh C, Borrueal N, Casellas F, Guarner F (2012). The gut microbiota in IBD. *Nature Reviews Gastroenterology & Hepatology* **9**: 599–608. DOI 10.1038/nrgastro.2012.152.
- Mari M, Caballero F, Colell A, Morales A, Caballeria J, Fernandez A, Enrich C, Fernandez-Checa JC, Garcia-Ruiz C (2006). Mitochondrial free cholesterol loading sensitizes to TNF- and Fas-mediated steatohepatitis. *Cell Metabolism* **4**: 185–198. DOI 10.1016/j.cmet.2006.07.006.
- Maziere C, Conte MA, Dantin F, Maziere JC (1999). Lipopolysaccharide enhances oxidative modification of low density lipoprotein by copper ions, endothelial and smooth muscle cells. *Atherosclerosis* **143**: 75–80. DOI 10.1016/S0021-9150(98)00277-9.
- McAleer IM, Kaplan GW, Bradley JS, Carroll SF, Griffith DP (2003). Endotoxin content in renal calculi. *Journal of Urology* **169**: 1813–1814. DOI 10.1097/01.ju.0000061965.51478.79.
- McConnell N, Campbell S, Gillanders I, Rolton H, Danesh B (2002). Risk factors for developing renal stones in inflammatory bowel disease. *BJU International* **89**: 835–841. DOI 10.1046/j.1464-410X.2002.02739.x.
- Mehta M, Goldfarb DS, Nazzal L (2016). The role of the microbiome in kidney stone formation. *International Journal of Surgery* **36**: 607–612. DOI 10.1016/j.ijso.2016.11.024.
- Mendes FD, Levy C, Enders FB, Loftus EV Jr, Angulo P, Lindor KD (2007). Abnormal hepatic biochemistries in patients with inflammatory bowel disease. *American Journal of Gastroenterology* **102**: 344–350. DOI 10.1111/j.1572-0241.2006.00947.x.
- Mikocka-Walus A, Knowles SR, Keefer L, Graff L (2016). Controversies revisited: A systematic review of the comorbidity of depression and anxiety with inflammatory bowel diseases. *Inflammatory Bowel Disease* **22**: 752–762. DOI 10.1097/MIB.0000000000000620.
- Miura K, Kodama Y, Inokuchi S, Schnabl B, Aoyama T, Ohnishi H, Olefsky JM, Brenner DA, Seki E (2010). Toll-like receptor 9 promotes steatohepatitis by induction of interleukin-1 β in mice. *Gastroenterology* **139**: 323–334.e7. DOI 10.1053/j.gastro.2010.03.052.
- Moreira Mde L, Tsuji M, Corbett AJ, Araújo MSS, Teixeira-Carvalho A, Martins-Filho OA, Peruhype-Magalhães V, Coelho-dos-Reis JG (2017). MAIT-cells: A tailor-made mate in the ancient battle against infectious diseases? *Immunology Letters* **187**: 53–60. DOI 10.1016/j.imlet.2017.05.007.
- Moris G (2014). Inflammatory bowel disease: An increased risk factor for neurologic complications. *World Journal of Gastroenterology* **20**: 1228–1237. DOI 10.3748/wjg.v20.i5.1228.
- Moura FA, De Andrade KQ, Santos JCF, Araújo ORP, Goulart MOF (2015). Antioxidant therapy for treatment of inflammatory bowel disease: Does it work? *Redox Biology* **6**: 617–639. DOI 10.1016/j.redox.2015.10.006.
- Moura FA, de Andrade KD, Araújo ORP, Nunes-Souza V, Santos JCF, Rabelo LA, Goulart MD (2016). Colonic and hepatic modulation by lipoic acid and/or N-acetylcysteine supplementation in mild ulcerative colitis induced by dextran sodium sulfate in rats. *Oxidative Medicine and Cellular Longevity* **2016**: 4047362. DOI 10.1155/2016/4047362.
- Muriel P (2009). Role of free radicals in liver diseases. *Hepatology International* **3**: 526–536. DOI 10.1007/s12072-009-9158-6.
- Muruve DA, Pétrilli V, Zais AK, White LR, Clark SA, Ross PJ, Parks RJ, Tschopp J (2008). The inflammasome recognizes cytosolic microbial and host DNA and triggers an innate immune response. *Nature* **452**: 103–107. DOI 10.1038/nature06664.
- Nakamoto N, Sasaki N, Aoki R, Miyamoto K, Suda W, Teratani T, Suzuki T, Koda Y, Chu PS, Taniki N, Yamaguchi A, Kanamori M, Kamada N, Hattori M, Ashida H, Sakamoto M, Atarashi K, Narushima S, Yoshimura A, Honda K, Sato T, Kanai T (2019). Gut pathobionts underlie intestinal barrier dysfunction and liver T helper 17 cell immune response in primary sclerosing cholangitis. *Nature Microbiology* **4**: 492–503. DOI 10.1038/s41564-018-0333-1.
- Nakamura YK, Metea C, Karstens L, Asquith M, Gruner H, Mosciobroci C, Lee I, Brislawn CJ, Jansson JK, Rosenbaum JT, Lin P (2016). Gut microbial alterations associated with protection from autoimmune uveitis. *Investigative Ophthalmology & Visual Science* **57**: 3747–3758. DOI 10.1167/iovs.16-19733.
- Navaneethan U (2014). Hepatobiliary manifestations of ulcerative colitis: An example of gut-liver crosstalk. *Gastroenterology Reports* **2**: 193–200. DOI 10.1093/gastro/gou036.
- Nemati R, Mehdizadeh S, Salimipour H, Yaghoobi E, Alipour Z, Tabib SM, Assadi M (2019). Neurological manifestations related to Crohn's disease: A boon for the workforce. *Gastroenterology Reports* **7**: 291–297. DOI 10.1093/gastro/gox034.
- Nguyen GC, Bernstein CN, Bitton A, Chan AK, Griffiths AM, Leontiadis GI, Geerts W, Bressler B, Butzner JD, Carrier M, Chande N, Marshall JK, Williams C, Keaton C (2014). Consensus statements on the risk, prevention, and treatment of venous thromboembolism in inflammatory bowel disease: Canadian Association of Gastroenterology. *Gastroenterology* **146**: 835–848.e6. DOI 10.1053/j.gastro.2014.01.042.
- Nielsen NM, Frisch M, Rostgaard K, Wohlfahrt J, Hjalgrim H, Koch-Henriksen N, Melbye M, Westergaard T (2008). Autoimmune diseases in patients with multiple sclerosis and their first-degree relatives: A nationwide cohort study in Denmark. *Multiple Sclerosis Journal* **14**: 823–829. DOI 10.1177/1352458508088936.
- Nishida A, Inoue R, Inatomi O, Bamba S, Naito Y, Andoh A (2018). Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clinical Journal of Gastroenterology* **11**: 1–10. DOI 10.1007/s12328-017-0813-5.
- Nishino K, Nishida A, Inoue R, Kawada Y, Ohno M, Sakai S, Inatomi O, Bamba S, Sugimoto M, Kawahara M, Naito Y, Andoh A (2018). Analysis of endoscopic brush samples identified mucosa-

- associated dysbiosis in inflammatory bowel disease. *Journal of Gastroenterology* **53**: 95–106. DOI 10.1007/s00535-017-1384-4.
- Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG (2000). Multiple sclerosis. *New England Journal of Medicine* **343**: 938–952. DOI 10.1056/NEJM200009283431307.
- O'Mahony S, Anderson N, Nuki G, Ferguson A (1992). Systemic and mucosal antibodies to Klebsiella in patients with ankylosing spondylitis and Crohn's disease. *Annals of the Rheumatic Diseases* **51**: 1296–1300. DOI 10.1136/ard.51.12.1296.
- Oikonomou K, Kapsoritakis A, Eleftheriadis T, Stefanidis I, Potamianos S (2011). Renal manifestations and complications of inflammatory bowel disease. *Inflammatory Bowel Diseases* **17**: 1034–1045. DOI 10.1002/ibd.21468.
- Olpin JD, Sjöberg BP, Stilwill SE, Jensen LE, Rezvani M, Shaaban AM (2017). Beyond the bowel: Extraintestinal manifestations of inflammatory bowel disease. *Radiographics* **37**: 1135–1160. DOI 10.1148/rg.2017160121.
- Orchard TR, Chua CN, Ahmad T, Cheng H, Welsh KI, Jewell DP (2002). Uveitis and erythema nodosum in inflammatory bowel disease: Clinical features and the role of HLA genes. *Gastroenterology* **123**: 714–718. DOI 10.1053/gast.2002.35396.
- Osna NA, Carter WG, Ganesan M, Kirpich IA, McClain CJ, Petersen DR, Shearn CT, Tomasi ML, Kharbanda KK (2016). Aberrant post-translational protein modifications in the pathogenesis of alcohol-induced liver injury. *World Journal of Gastroenterology* **22**: 6192–6200. DOI 10.3748/wjg.v22.i27.6192.
- Oussalah A, Gueant JL, Peyrin-Biroulet L (2011). Meta-analysis: Hyperhomocysteinaemia in inflammatory bowel diseases. *Alimentary Pharmacology & Therapeutics* **34**: 1173–1184. DOI 10.1111/j.1365-2036.2011.04864.x.
- Pagliassotti MJ (2012). Endoplasmic reticulum stress in nonalcoholic fatty liver disease. *Annual Review of Nutrition* **32**: 17–33. DOI 10.1146/annurev-nutr-071811-150644.
- Palumbo C, Restellini S, Chao CY, Aruljothy A, Lemieux C, Wild G, Afif W, Lakatos PL, Bitton A, Cocciolillo S, Ghali P, Bessissow T, Sebastiani G (2019). Screening for nonalcoholic fatty liver disease in inflammatory bowel diseases: A cohort study using transient elastography. *Inflammatory Bowel Diseases* **25**: 124–133. DOI 10.1093/ibd/izy200.
- Panes J, Granger DN (1998). Leukocyte-endothelial cell interactions: Molecular mechanisms and implications in gastrointestinal disease. *Gastroenterology* **114**: 1066–1090. DOI 10.1016/S0016-5085(98)70328-2.
- Papa A, Scaldaferrri F, Danese S, Guglielmo S, Roberto I, Bonizzi M, Mocchi G, Felice C, Ricci C, Andrisani G, Fedeli G, Gasbarrini G, Gasbarrini A (2008). Vascular involvement in inflammatory bowel disease: Pathogenesis and clinical aspects. *Digestive Diseases* **26**: 149–155. DOI 10.1159/000116773.
- Park S, Chun J, Han KD, Soh H, Choi K, Kim JH, Lee J, Lee C, Im JP, Kim JS (2018). Increased end-stage renal disease risk in patients with inflammatory bowel disease: A nationwide population-based study. *World Journal of Gastroenterology* **24**: 4798–4808. DOI 10.3748/wjg.v24.i42.4798.
- Parks JH, Worcester EM, O'Connor RC, Coe FL (2003). Urine stone risk factors in nephrolithiasis patients with and without bowel disease. *Kidney International* **63**: 255–265. DOI 10.1046/j.1523-1755.2003.00725.x.
- Pimentel-Santos FM, Matos M, Ligeiro D, Mourão AF, Ribeiro C, Costa J, Santos H, Barcelos A, Pinto P, Cruz M, Sousa E, Santos RA, Fonseca JE, Trindade H, Guedes-Pinto H, Branco JC (2013). HLA alleles and HLA-B27 haplotypes associated with susceptibility and severity of ankylosing spondylitis in a Portuguese population. *Tissue Antigens* **82**: 374–379. DOI 10.1111/tan.12238.
- Poulou AC, Goumas KE, Dandakis DC, Tyrmpas I, Panagiotaki M, Georgouli A, Soutos DC, Archimandritis A (2006). Microproteinuria in patients with inflammatory bowel disease: Is it associated with the disease activity or the treatment with 5-aminosalicylic acid? *World Journal of Gastroenterology* **12**: 739–746. DOI 10.3748/wjg.v12.i5.739.
- Primas C, Novacek G, Schweiger K, Mayer A, Eser A, Papay P, Gratzner C, Angelberger S, Dejaco C, Reinisch W, Vogelsang H (2013). Renal insufficiency in IBD—prevalence and possible pathogenetic aspects. *Journal of Crohn's and Colitis* **7**: e630–e634. DOI 10.1016/j.crohns.2013.05.001.
- Purchiaroni F, Tortora A, Gabrielli M, Bertucci F, Gigante G, Ianiro G, Ojetti V, Scarpellini E, Gasbarrini A (2013). The role of intestinal microbiota and the immune system. *European Review for Medical and Pharmacological Sciences* **17**: 323–333.
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Li S, Jian M, Zhou Y, Li Y, Zhang X, Li S, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J, Bork P, Ehrlich S D, Wang J (2010). A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* **464**: 59–65. DOI 10.1038/nature08821.
- Qin L, Liu Y, Hong JS, Crews FT (2013). NADPH oxidase and aging drive microglial activation, oxidative stress, and dopaminergic neurodegeneration following systemic LPS administration. *Glia* **61**: 855–868. DOI 10.1002/glia.22479.
- Quevrain E, Maubert MA, Michon C, Chain F, Marquant R, Tailhades J, Miquel S, Carlier L, Bermúdez-Humarán LG, Pigneur B, Lequin O, Kharrat P, Thomas G, Rainteau D, Aubry C, Breyner N, Afonso C, Lavielle S, Grill JP, Chassaing G, Chatel JM, Trugnan G, Xavier R, Langella P, Sokol H, Seksik P (2016). Identification of an anti-inflammatory protein from *Faecalibacterium prausnitzii*, a commensal bacterium deficient in Crohn's disease. *Gut* **65**: 415–425. DOI 10.1136/gutjnl-2014-307649.
- Rahman I, Biswas SK (2004). Non-invasive biomarkers of oxidative stress: Reproducibility and methodological issues. *Redox Report* **9**: 125–143. DOI 10.1179/135100004225005219.
- Ratajczak AE, Szymczak-Tomczak A, Skrzypczak-Zielińska M, Rychter AM, Zawada A, Dobrowolska A, Krela-Kaźmierczak I (2020). Vitamin C deficiency and the risk of osteoporosis in patients with an inflammatory bowel disease. *Nutrients* **12**: 2263. DOI 10.3390/nu12082263.
- Restellini S, Chazouilleres O, Frossard JL (2017). Hepatic manifestations of inflammatory bowel diseases. *Liver International* **37**: 475–489. DOI 10.1111/liv.13265.
- Reyes-Gordillo K, Shah R, Muriel P (2017). Oxidative stress and inflammation in hepatic diseases: Current and future therapy. *Oxidative Medicine and Cellular Longevity* **2017**: 3140673. DOI 10.1155/2017/3140673.
- Robertson G, Leclercq I, Farrell GC (2001). Nonalcoholic steatosis and steatohepatitis. II. Cytochrome P-450 enzymes and oxidative stress. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **281**: G1135–G1139. DOI 10.1152/ajpgi.2001.281.5.G1135.

- Roifman I, Sun YC, Fedwick JP, Panaccione R, Buret AG, Liu H, Rostom A, Anderson TJ, Beck PL (2009). Evidence of endothelial dysfunction in patients with inflammatory bowel disease. *Clinical Gastroenterology and Hepatology* **7**: 175–182. DOI 10.1016/j.cgh.2008.10.021.
- Sabino J, Vieira-Silva S, Machiels K, Joossens M, Falony G, Ballet V, Ferrante M, Van Assche G, Van der Merwe S, Vermeire S, Raes J (2016). Primary sclerosing cholangitis is characterised by intestinal dysbiosis independent from IBD. *Gut* **65**: 1681–1689. DOI 10.1136/gutjnl-2015-311004.
- Sadaf H, Raza SI, Hassan SW (2017). Role of gut microbiota against calcium oxalate. *Microbial Pathogenesis* **109**: 287–291. DOI 10.1016/j.micpath.2017.06.009.
- Salmi M, Jalkanen S (2001). Human leukocyte subpopulations from inflamed gut bind to joint vasculature using distinct sets of adhesion molecules. *Journal of Immunology* **166**: 4650–4657. DOI 10.4049/jimmunol.166.7.4650.
- Santos J, Saunders PR, Hanssen NP, Yang PC, Yates D, Groot JA, Perdue MH (1999). Corticotropin-releasing hormone mimics stress-induced colonic epithelial pathophysiology in the rat. *American Journal of Physiology* **277**: G391–G399.
- Scaldaferri F, Lancellotti S, Pizzoferrato M, De Cristofaro R (2011). Haemostatic system in inflammatory bowel diseases: New players in gut inflammation. *World Journal of Gastroenterology* **17**: 594–608. DOI 10.3748/wjg.v17.i5.594.
- Scaldaferri F, Sans M, Vetrano S, Correale C, Arena V, Pagano N, Rando G, Romeo F, Potenza AE, Repici A, Malesci A, Danese S (2009). The role of MAPK in governing lymphocyte adhesion to and migration across the microvasculature in inflammatory bowel disease. *European Journal of Immunology* **39**: 290–300. DOI 10.1002/eji.200838316.
- Scher JU, Littman DR, Abramson SB (2016). Microbiome in inflammatory arthritis and human rheumatic diseases. *Arthritis & Rheumatology* **68**: 35–45. DOI 10.1002/art.39259.
- Schinzari F, Armuzzi A, De Pascalis B, Mores N, Tesaro M, Melina D, Cardillo C (2008). Tumor necrosis factor- α antagonism improves endothelial dysfunction in patients with Crohn's disease. *Clinical Pharmacology & Therapeutics* **83**: 70–76. DOI 10.1038/sj.clpt.6100229.
- Schuermann GM, Aber-Bishop AE, Facer P, Lee JC, Rampton DS, Dore CJ, Polak JM (1993). Altered expression of cell adhesion molecules in uninvolved gut in inflammatory bowel disease. *Clinical & Experimental Immunology* **94**: 341–347. DOI 10.1111/j.1365-2249.1993.tb03455.x.
- Seldin MM, Meng Y, Qi H, Zhu WF, Wang Z, Hazen SL, Lusic AJ, Shih DM (2016). Trimethylamine N-oxide promotes vascular inflammation through signaling of mitogen-activated protein kinase and nuclear factor- κ B. *Journal of the American Heart Association* **5**: e002767. DOI 10.1161/JAHA.115.002767.
- Sen U, Mishra PK, Tyagi N, Tyagi SC (2010). Homocysteine to hydrogen sulfide or hypertension. *Cell Biochemistry and Biophysics* **57**: 49–58. DOI 10.1007/s12013-010-9079-y.
- Serviddio G, Bellanti F, Vendemiale G, Altomare E (2011). Mitochondrial dysfunction in nonalcoholic steatohepatitis. *Expert Review of Gastroenterology & Hepatology* **5**: 233–244. DOI 10.1586/egh.11.11.
- Shearn CT, Fennimore B, Orlicky DJ, Gao YR, Saba LM, Battista KD, Aivazidis S, Assiri M, Harris PS, Michel C, Merrill GF, Schmidt EE, Colgan SP, Petersen DR (2019). Cholestatic liver disease results increased production of reactive aldehydes and an atypical periportal hepatic antioxidant response. *Free Radical Biology and Medicine* **143**: 101–114. DOI 10.1016/j.freeradbiomed.2019.07.036.
- Shearn CT, Orlicky DJ, Petersen DR (2018). Dysregulation of antioxidant responses in patients diagnosed with concomitant Primary Sclerosing Cholangitis/Inflammatory Bowel Disease. *Experimental and Molecular Pathology* **104**: 1–8. DOI 10.1016/j.yexmp.2017.11.012.
- Shemer A, Erny D, Jung S, Prinz M (2015). Microglia plasticity during health and disease: An immunological perspective. *Trends in Immunology* **36**: 614–624. DOI 10.1016/j.it.2015.08.003.
- Sheth T, Pitchumoni CS, Das KM (2015). Management of musculoskeletal manifestations in inflammatory bowel disease. *Gastroenterology Research and Practice* **2015**: 387891. DOI 10.1155/2015/387891.
- Siener R, Bangen U, Sidhu H, Honow R, von Unruh G, Hesse A (2013). The role of Oxalobacter formigenes colonization in calcium oxalate stone disease. *Kidney International* **83**: 1144–1149. DOI 10.1038/ki.2013.104.
- Silva J, Brito BS, Silva INN, Nóbrega VG, Silva MCSM, Gomes HDN, Fortes FM, Pimentel AM, Mota J, Almeida N, Surlo VC, Lyra A, Rocha R, Santana GO (2019). Frequency of hepatobiliary manifestations and concomitant liver disease in inflammatory bowel disease patients. *BioMed Research International* **2019**: 7604939. DOI 10.1155/2019/7604939.
- Silverman MD, Tumuluri RJ, Davis M, Lopez G, Rosenbaum JT, Lelkes PI (2002). Homocysteine upregulates vascular cell adhesion molecule-1 expression in cultured human aortic endothelial cells and enhances monocyte adhesion. *Arteriosclerosis, Thrombosis, and Vascular Biology* **22**: 587–592. DOI 10.1161/01.ATV.0000014221.30108.08.
- Singer II, Kawka DW, Scott S, Weidner JR, Mumford RA, Riehl TE, Stenson WF (1996). Expression of inducible nitric oxide synthase and nitrotyrosine in colonic epithelium in inflammatory bowel disease. *Gastroenterology* **111**: 871–885. DOI 10.1016/S0016-5085(96)70055-0.
- Singh S, Kumar N, Loftus EV Jr, Kane SV (2013). Neurologic complications in patients with inflammatory bowel disease: Increasing relevance in the era of biologics. *Inflammatory Bowel Disease* **19**: 864–872. DOI 10.1002/ibd.23011.
- Siva S, Barrack ER, Reddy GPV, Thamilselvan V, Thamilselvan S, Menon M, Bhandari M (2009). A critical analysis of the role of gut Oxalobacter formigenes in oxalate stone disease. *BJU International* **103**: 18–21. DOI 10.1111/j.1464-410X.2008.08122.x.
- Specia S, Dubuquoy L (2017). Chronic bowel inflammation and inflammatory joint disease: Pathophysiology. *Joint Bone Spine* **84**: 417–420. DOI 10.1016/j.jbspin.2016.12.016.
- Stadnicki A (2012). Involvement of coagulation and hemostasis in inflammatory bowel diseases. *Current Vascular Pharmacology* **10**: 659–669. DOI 10.2174/157016112801784495.
- Stewart CS, Duncan SH, Cave DR (2004). Oxalobacter formigenes and its role in oxalate metabolism in the human gut. *FEMS Microbiology Letters* **230**: 1–7. DOI 10.1016/S0378-1097(03)00864-4.
- Suh HY, Lee WJ, Na SY (2019). Dermatologic manifestations in inflammatory bowel disease. *Korean J Gastroenterol* **73**: 285–293.
- Sunny NE, Parks EJ, Browning JD, Burgess SC (2011). Excessive hepatic mitochondrial TCA cycle and gluconeogenesis in humans with nonalcoholic fatty liver disease. *Cell Metabolism* **14**: 804–810. DOI 10.1016/j.cmet.2011.11.004.
- Szeto CC, McIntyre CW, Li PK (2018a). Circulating bacterial fragments as cardiovascular risk factors in CKD. *Journal of the American Society of Nephrology* **29**: 1601–1608. DOI 10.1681/ASN.2018010068.

- Szeto W, van der Bent A, Petty CR, Reich J, Farraye F, Fishman LN (2018b). Use of social media for health-related tasks by adolescents with inflammatory bowel disease: A step in the pathway of transition. *Inflammatory Bowel Diseases* **24**: 1114–1122. DOI 10.1093/ibd/izy021.
- Takahashi K, Nishida A, Fujimoto T, Fujii M, Shiyoa M, Imaeda H, Inatomi O, Bamba S, Sugimoto M, Andoh A (2016). Reduced abundance of butyrate-producing bacteria species in the fecal microbial community in Crohn's Disease. *Digestion* **93**: 59–65.
- Takemura T, Okada M, Yagi K, Kuwajima H, Yanagida H (2002). An adolescent with IgA nephropathy and Crohn disease: Pathogenetic implications. *Pediatric Nephrology* **17**: 863–866. DOI 10.1007/s00467-002-0943-x.
- Taleban S, Li D, Targan SR, Ippoliti A, Brant SR, Cho JH, Duerr RH, Rioux JD, Silverberg MS, Vasilias EA, Rotter JL, Haritunians T, Shih DQ, Dubinsky M, Melmed GY, McGovern DPB (2016). Ocular manifestations in inflammatory bowel disease are associated with other extra-intestinal manifestations, gender, and genes implicated in other immune-related traits. *Journal of Crohn's and Colitis* **10**: 43–49. DOI 10.1093/ecco-jcc/jjv178.
- Tang WH, Hazen SL (2014). The contributory role of gut microbiota in cardiovascular disease. *Journal of Clinical Investigation* **124**: 4204–4211. DOI 10.1172/JCI72331.
- Taugrog JD, Richardson JA, Croft JT, Simmons WA, Zhou M, Fernández-Sueiro JL, Balish E, Hammer RE (1994). The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. *Journal of Experimental Medicine* **180**: 2359–2364. DOI 10.1084/jem.180.6.2359.
- Timani S, Mutasim DF (2008). Skin manifestations of inflammatory bowel disease. *Clinics in Dermatology* **26**: 265–273. DOI 10.1016/j.clindermatol.2007.10.018.
- Tito RY, Cypers H, Joossens M, Varkas GB, Van Praet L, Glorieus E, Van den Bosch F, De Vos M, Raes J, Elewaut D (2017). Brief report: Dialister as a microbial marker of disease activity in spondyloarthritis. *Arthritis & Rheumatology* **69**: 114–121. DOI 10.1002/art.39802.
- Tsaitas C, Semertzidou A, Sinakos E (2014). Update on inflammatory bowel disease in patients with primary sclerosing cholangitis. *World Journal of Hepatology* **6**: 178–187. DOI 10.4254/wjh.v6.i4.178.
- Tyagi N, Sedoris KC, Steed M, Ovechkin AV, Moshal KS, Tyagi SC (2005). Mechanisms of homocysteine-induced oxidative stress. *American Journal of Physiology-Heart and Circulatory Physiology* **289**: H2649–H2656. DOI 10.1152/ajpheart.00548.2005.
- van Assche G, Dignass A, Bokemeyer B, Danese S, Gionchetti P, Moser G, Beaugerie L, Gomollón F, Häuser W, Herrlinger K, Oldenburg B, Panes J, Portela F, Rogler G, Stein JC, Tilg H, Travis S, Lindsay JO (2013). Second European evidence-based consensus on the diagnosis and management of ulcerative colitis part 3: Special situations. *Journal of Crohn's and Colitis* **7**: 1–33. DOI 10.1016/j.crohns.2012.09.005.
- van Hogezaand RA, Hamdy NA (2006). Skeletal morbidity in inflammatory bowel disease. *Scandinavian Journal of Gastroenterology Supplement* **243**: 59–64. DOI 10.1080/00365520600664276.
- van Staa TP (2006). The pathogenesis, epidemiology and management of glucocorticoid-induced osteoporosis. *Calcified Tissue International* **79**: 129–137. DOI 10.1007/s00223-006-0019-1.
- van Staa TP, Leufkens HG, Abenham L, Zhang B, Cooper C (2005). Use of oral corticosteroids and risk of fractures. *Journal of Bone and Mineral Research* **20**: 1487–1494, discussion 1486.
- Vavricka SR, Brun L, Ballabeni P, Pittet V, Vavricka BMP, Zeitz J, Rogler G, Schoepfer AM (2011). Frequency and risk factors for extraintestinal manifestations in the Swiss inflammatory bowel disease cohort. *American Journal of Gastroenterology* **106**: 110–119. DOI 10.1038/ajg.2010.343.
- Vavricka SR, Schoepfer A, Scharl M, Lakatos PL, Navarini A, Rogler G (2015). Extraintestinal manifestations of inflammatory bowel disease. *Inflammatory Bowel Disease* **21**: 1982–1992. DOI 10.1097/MIB.0000000000000392.
- Venkatesh PG, Navaneethan U, Shen B (2011). Hepatobiliary disorders and complications of inflammatory bowel disease. *Journal of Digestive Diseases* **12**: 245–256. DOI 10.1111/j.1751-2980.2011.00511.x.
- Vernon G, Baranova A, Younossi ZM (2011). Systematic review: The epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. *Alimentary Pharmacology & Therapeutics* **34**: 274–285. DOI 10.1111/j.1365-2036.2011.04724.x.
- Villaran RF, Espinosa-Oliva AM, Sarmiento M, De Pablos RDM, Argüelles S, Delgado-Cortés MDJ, Sobrino V, Van Rooijen N, Venero JL, Herrera AJ, Cano J, Machado A (2010). Ulcerative colitis exacerbates lipopolysaccharide-induced damage to the nigral dopaminergic system: Potential risk factor in Parkinson's disease. *Journal of Neurochemistry* **114**: 1687–1700. DOI 10.1111/j.1471-4159.2010.06879.x.
- Villumsen M, Aznar S, Pakkenberg B, Jess T, Brudek T (2019). Inflammatory bowel disease increases the risk of Parkinson's disease: A Danish nationwide cohort study 1977–2014. *Gut* **68**: 18–24. DOI 10.1136/gutjnl-2017-315666.
- Walker JR, Ediger JP, Graff LA, Greenfeld JM, Clara I, Lix L, Rawsthorne P, Miller N, Rogala L, McPhail CM, Bernstein CN (2008). The Manitoba IBD cohort study: A population-based study of the prevalence of lifetime and 12-month anxiety and mood disorders. *American Journal of Gastroenterology* **103**: 1989–1997. DOI 10.1111/j.1572-0241.2008.01980.x.
- Wang G, Woo CW, Sung FL, Siow YL, Karmin O (2002). Increased monocyte adhesion to aortic endothelium in rats with hyperhomocysteinemia: Role of chemokine and adhesion molecules. *Arteriosclerosis, Thrombosis, and Vascular Biology* **22**: 1777–1783. DOI 10.1161/01.ATV.0000035404.18281.37.
- Wang Z, Roberts AB, Buffa JA, Levison BS, Zhu W, Org E, Gu X, Huang Y, Zamanian-Daryoush M, Culley MK, DiDonato AJ, Fu X, Hazen JE, Krajcik D, DiDonato JA, Lusis AJ, Hazen SL (2015). Non-lethal inhibition of gut microbial trimethylamine production for the treatment of atherosclerosis. *Cell* **163**: 1585–1595. DOI 10.1016/j.cell.2015.11.055.
- Wang Z, Zhao Y (2018). Gut microbiota derived metabolites in cardiovascular health and disease. *Protein & Cell* **9**: 416–431. DOI 10.1007/s13238-018-0549-0.
- Wei W, Ding M, Zhou K, Xie H, Zhang M, Zhang C (2017). Protective effects of wedelolactone on dextran sodium sulfate induced murine colitis partly through inhibiting the NLRP3 inflammasome activation via AMPK signaling. *Biomedicine & Pharmacotherapy* **94**: 27–36. DOI 10.1016/j.biopha.2017.06.071.
- Wiesner P, Choi SH, Almazan F, Benner C, Huang W, Diehl CJ, Gonen A, Butler S, Witztum JL, Glass CK, Miller YI (2010). Low doses of lipopolysaccharide and minimally oxidized low-density lipoprotein cooperatively activate macrophages via NF-kappaB and AP-1: Possible mechanism for acceleration of atherosclerosis by subclinical endotoxemia. *Circulation Research* **107**: 56–65. DOI 10.1161/CIRCRESAHA.110.218420.

- Worcester EM (2002). Stones from bowel disease. *Endocrinology and Metabolism Clinics of North America* **31**: 979–999. DOI 10.1016/S0889-8529(02)00035-X.
- Wu P, Jia F, Zhang B, Zhang P (2017). Risk of cardiovascular disease in inflammatory bowel disease. *Experimental and Therapeutic Medicine* **13**: 395–400. DOI 10.3892/etm.2016.3966.
- Yang L, Wang L, Wang X, Xian CJ, Lu H (2016). A possible role of intestinal microbiota in the pathogenesis of ankylosing spondylitis. *International Journal of Molecular Sciences* **17**: 12.
- Yuan J, Chen C, Cui J, Lu J, Yan C, Wei X, Zhao X, Li NN, Li S, Xue G, Cheng W, Li B, Li H, Lin W, Tian C, Zhao J, Han J, An D, Zhang Q, Wei H, Zheng M, Ma X, Li W, Chen X, Zhang Z, Zeng H, Ying S, Wu JX, Yang R, Liu D (2019). Fatty liver disease caused by high-alcohol-producing *Klebsiella pneumoniae*. *Cell Metabolism* **30**: 1172. DOI 10.1016/j.cmet.2019.11.006.
- Zanoli L, Rastelli S, Inserra G, Castellino P (2015). Arterial structure and function in inflammatory bowel disease. *World Journal of Gastroenterology* **21**: 11304–11311. DOI 10.3748/wjg.v21.i40.11304.
- Zezos P, Kouklakis G, Saibil F (2014). Inflammatory bowel disease and thromboembolism. *World Journal of Gastroenterology* **20**: 13863–13878. DOI 10.3748/wjg.v20.i38.13863.
- Zhang Y, Ye P, Luo L, Bai Y, Xu R, Xiao W, Liu D, Wu H (2014). Association between arterial stiffness and risk of coronary artery disease in a community-based population. *Chinese Medical Journal* **127**: 3944–3947.
- Zhao C, Ling Z, Newman MB, Bhatia A, Carvey PM (2007). TNF- α knockout and minocycline treatment attenuates blood-brain barrier leakage in MPTP-treated mice. *Neurobiology of Disease* **26**: 36–46. DOI 10.1016/j.nbd.2006.11.012.
- Zhou W, Cao Q, Peng Y, Zhang QJ, Castrillon DH, DePinho RA, Liu ZP (2009). FoxO4 inhibits NF- κ B and protects mice against colonic injury and inflammation. *Gastroenterology* **137**: 1403–1414. DOI 10.1053/j.gastro.2009.06.049.
- Zhu W, Gregory JC, Org E, Buffa JA, Gupta N, Wang Z, Li L, Fu X, Wu Y, Mehrabian M, Sartor RB, McIntyre TM, Silverstein RL, Tang WHW, DiDonato JA, Brown JM, Luscis AJ, Hazen SL (2016). Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. *Cell* **165**: 111–124. DOI 10.1016/j.cell.2016.02.011.
- Zieman SJ, Melenovsky V, Kass DA (2005). Mechanisms, pathophysiology, and therapy of arterial stiffness. *Arteriosclerosis, Thrombosis, and Vascular Biology* **25**: 932–943. DOI 10.1161/01.ATV.0000160548.78317.29.
- Zmora N, Levy M, Pevsner-Fischer M, Elinav E (2017). Inflammasomes and intestinal inflammation. *Mucosal Immunology* **10**: 865–883. DOI 10.1038/mi.2017.19.