

## Influence of Biochar on Nitrogen Use Efficiency and Root Morphology of Rice-Seedling in Two Contrasting Paddy Soils

Lei Chu<sup>1,2</sup>, Yu Zhang<sup>1,2</sup>, Long Qian<sup>1,2</sup>, Dandan Zhu<sup>1,2</sup> and Haijun Sun<sup>1,2,\*</sup>

<sup>1</sup>Co-Innovation Center for Sustainable Forestry in Southern China, College of Forestry, Nanjing Forestry University, Nanjing, 210037, China

<sup>2</sup>Key Laboratory of Soil and Water Conservation and Ecological Restoration of Jiangsu Province, Nanjing Forestry University, Nanjing, 210037, China

\*Corresponding Author: Haijun Sun. Email: hjsun@njfu.edu.cn

Received: 03 October 2020; Accepted: 14 October 2020

**Abstract:** Biochar may affect the root morphology and nitrogen (N) use efficiency (NUE) of rice at seedling stage, which has not been clearly verified until now. To clarify it, we conducted a pot experiment regarding to two soil types (Hydragric Anthrosol and Haplic Acrisol), two biochar application rates (0.5 wt% and 1.5 wt %) and two rice varieties (common rice var. Xiushui134 and hybrid super rice var. Zhongkejiayou12-6) meanwhile. Seedling NUE of common rice Xiuhui134 was significantly increased ( $p < 0.05$ ) by 78.2% in Hydragric Anthrosol and by 91.4% in Haplic Acrisol following biochar addition with 1.5 wt%. However, biochar addition exerted no influence on seedling NUE of super rice Zhongkejiayou12-6 in both soils. Overall, 0.09–0.10 units higher soil pH and 105–116% higher soil  $\text{NH}_4^+\text{-N}$  were observed in Xiushui134 growing two soils with 1.5 wt% biochar. In addition, improved root morphology (including longer root length, larger root surface area, bigger root volume, and more root tips) contributed to the higher seedling NUE of Xiushui134 in two soils. The soil pH and  $\text{NH}_4^+\text{-N}$  content, also the root morphology were influenced by biochar, which though could not thoroughly explained the NUE of Zhongkejiayou12-6. In conclusion, biochar application to paddy soil changed soil pH and  $\text{NH}_4^+\text{-N}$  content, root growth, and the consequent seedling NUE of rice, which effects are relative with rice cultivar, biochar addition rate, and soil type.

**Keywords:** Ammonium; biochar; nitrogen management; rice paddy soil; root morphology; super rice

### 1 Introduction

Rice (*Oryza sativa* L.) is one of the most important cereal crops, feeding more than half of the world's population. However, in order to improve yields, rice paddy fields are often over fertilized with nitrogen (N) [1], which is a serious problem in Asia, especially China. Excessive N application not only reduces N use efficiency (NUE), but also causes N leaching and runoff losses, increases ammonia volatilization and nitrous oxide emission [2,3]. Therefore, effective improvement the NUE of rice under optional N input rate is of great significance in ensuring food security and protecting the environment meanwhile [4]. To



This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

reach this aim, we should firstly try to enhance the NUE of rice seedling and ideal root morphological traits [5]. As for rice, root morphological characteristics are closely associated with NUE and yield [6].

Biochar is a kind of pyrolysis material with large specific surface area and rich pore structure mainly prepared from plant materials under no or limited oxygen condition [7]. Recently, biochar is widely studied and used to rice paddy soil, attributing to its functions in improving soil properties [8], reducing greenhouse gas emission and ammonia volatilization [9,10], and enhancing NUE and thereby the grain production [11]. Thus we can reduce the application of N and keep grain yields in biochar-added rice paddy soil. What is more, there are reports showing that increases in grain yield and NUE was mainly attributed to root biomass, root length, root tips, and root-oxidizing activity [5,12]. According to previous studies, biochar could improve soil physical structure, which consequently impact the rice root development [13,14]. Field with application of biochar increased root biomass (+32%), root volume (+29%) and root surface area (+39%) of crops [13]. It has been proved that biochar can increase lateral roots and fine roots of crops [14]. For rice crops, ammonium ( $\text{NH}_4^+\text{-N}$ ) and nitrate ( $\text{NO}_3^-\text{-N}$ ) are the main forms of N available to the young rice plants [5,15]. However, root morphology of rice affects NUE and biochar also has an effect on the conversion of N forms [16]. Therefore, it is necessary to explore the response mechanism between rice root morphology and biochar application. However, whether biochar affects NUE by changing rice seedling root morphology has been little documented.

Asia, as the main rice production area all over the world, has many soil types distributed with varied properties, and also many rice varieties, including traditional cultivated and super hybrid rice cultivars, which was also in China [5,17]. Therefore, the hypothesis of current work is biochar can influence the NUE of rice seedling via changing the root morphology, which effect might depended on soil type, rice variety and biochar application rate. Here, we conduct a pot experiment to clarify the responses of rice seedling NUE and root morphology to biochar application with low/higher rates and the potential mechanisms.

## 2 Material and Methods

### 2.1 Background Information and Soil Pots Installation

Two types of typical paddy soils were collected from Yixing (31°28' N, 119°59' E), Jiangsu province, and from Yingtan (28°12' N, 117°10' E), Jiangxi province, respectively. The Yixing soil is classified as Hydragric Anthrosol paddy soil and the Yingtan soil is Haplic Acrisol paddy soil according to WRB [18]. These two soils represented two dominant soil types in Chinese rice production systems. After being dried and crushed passing 2-mm sieve, soil samples were correspondingly repacked to transparent pot (inner diameter 12.5 cm, height 20 cm) with 2.4 kg soil each. Biochar was produced by continuous slow pyrolysis of wheat straw without oxygen at 500°C. The biochar has a BET (Brunauer-Emmett-Teller) surface area of 51.5  $\text{m}^2/\text{g}$  and contains 174 g/kg ash. The quantitative biochar (biochar: soil = 0.5 wt% and 1.5 wt%) were homogeneously mixed with the soil sample during repacking practice. The selected properties of tested soils and the biochar are shown in Tab. 1.

**Table 1:** Selected properties of two tested soils and wheat straw biochar

Soils/biochar	pH	Total N	Total P	Total K	SOC	CEC
			g/kg			cmol/kg
Hydragric anthrosol	6.38	1.56	0.96	4.12	22.8	15.0
Haplic acrisol	5.05	1.90	1.29	4.44	18.1	8.89
Wheat straw biochar	9.51	13.3	4.40	20.9	–	27.5

Note: SOC: soil organic carbon; CEC: cation exchange capacity.

A common rice variety Xiushui134 (XS) and a hybrid super rice variety Zhongkejiayou12-6 (ZK), according to previous work [5], were selected for the current experiment. The 28-day rice seedlings were transplanted to the pots (2 seedlings per pot).

## 2.2 Experimental Design and Management

There were three treatments for each soil type planted with each rice variety: Urea (receiving urea N fertilizer), Urea + 0.5%BC (receiving urea N fertilizer plus 0.5 wt% biochar), and Urea + 1.5%BC (receiving urea N fertilizer plus 1.5 wt% biochar). Every type of soil planted with each rice variety included one control treatment (no application of N and biochar) to calculate the NUE of rice seedling. Three replicates were maintained for each treatment.

Pre-flooding irrigation for each pot was formed one week prior to rice transplanting. Floodwater was continuously maintained at a depth of 3–5 cm in all soil pots. We homogeneously broadcasted 0.25 g urea to each pot (approximately equal to 240 kg N/ha under field condition).

## 2.3 Sample and Measurement

Fourteen days after transplanting, all treated rice seedling were harvested for the measurements of shoot biomass, N content. At the same time, the rice seedling roots were washed with deionized water and immediately for observation of root morphology. Soils were sampled for pH (soil: water = 1: 5),  $\text{NH}_4^+$ -N and  $\text{NO}_3^-$ -N determinations.

The harvested rice seedling samples were placed in a drying oven at 105°C and then were dried at 75°C to a consistent weight, when we recorded the shoot biomass (dry weight). Dry plant samples were ground into powder and digested with  $\text{H}_2\text{SO}_4\text{-H}_2\text{O}_2$ , then the total N content of rice plant was determined using the Kjeldahl method [5]. The NUE was calculated as the percentage of applied fertilizer N recovered in above-ground biomass minus that of the control treatment. The soil  $\text{NH}_4^+$ -N and  $\text{NO}_3^-$ -N were extracted by 2.0 mol/L KCl, then the  $\text{NH}_4^+$ -N and  $\text{NO}_3^-$ -N concentrations of the KCl extracted soil solution were measured by a San<sup>++</sup> Continuous Flow Analyzer (Skalar, Netherlands).

Rice roots and shoots were separated and washed with deionized water. Root morphology including total root length, root volume, root surface area, average root diameter, and root tip number in the two rice cultivars were measured using a root analysis instrument WinRhizo-LA1600 (Regent Instruments Inc., Quebec, Canada) [5].

## 2.4 Statistical Analysis

Analysis of variance (one-way ANOVA) was used to determine the significance of the difference between treatments. Duncan multiple-comparison test was conducted to determine the differences among treatments ( $p < 0.05$ ) (SPSS Ver. 16.0 for Windows, SPSS Inc., Chicago, IL, USA).

# 3 Results

## 3.1 Shoot Biomass and NUE of Rice at Seedling Stage

Compared to the control, application of N fertilizer significantly increased ( $p < 0.05$ ) shoot biomass of two rice seedlings planted in two soils (Tab. 2). Generally, biochar amendment exerted no remarkable effect on rice shoot biomass. Exceptionally, averaged 19.0% higher shoot biomass was observed under Urea + 1.5%BC than that under Urea only treatment for Xiushui134 planted in Haplic Acrisol. The NUEs of Xiushui134 seedlings were only 6.62–11.8% and 4.89–9.36%, when grown in Hydragric Anthrosol and Haplic Acrisol, respectively. Zhongkejiayou12-6 had relatively higher NUEs, reached 10.7–12.1% in Hydragric Anthrosol and 16.2–18.2% in Haplic Acrisol. Data in Tab. 1 suggested that biochar amendment did not improve the NUE of Zhongkejiayou tested under both soils. However, NUEs of Xiushui134 under biochar-added treatments were higher than that under Urea treatment. Particularly, Urea

+ 1.5%BC had 78.2% and 91.4% significantly higher ( $p < 0.05$ ) NUEs than that of Urea, in Hydragric Anthrosol and Haplic Acrisol, respectively.

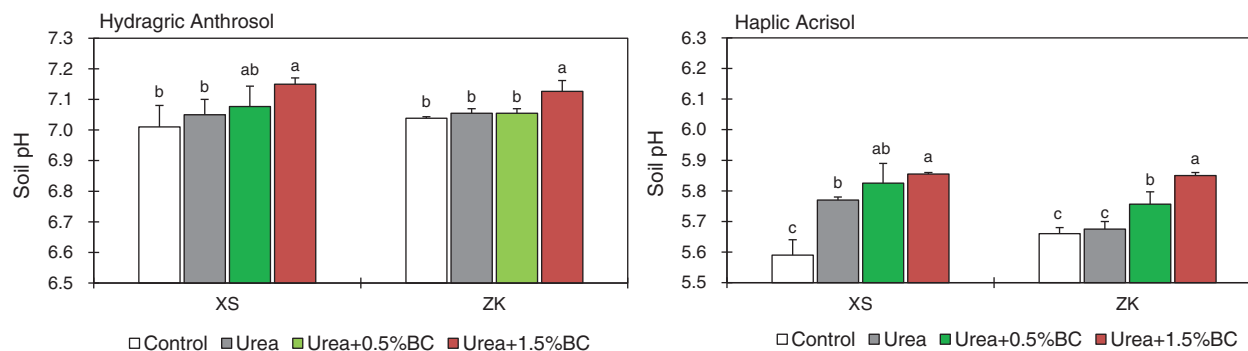
**Table 2:** Impacts of biochar addition on shoot biomass and nitrogen use efficiency (NUE)

Soil type	Rice variety	Treatment	Shoot biomass (g)	NUE (%)
Hydragric Anthrosol	Xiushui134	Control	0.22 ± 0.02 b	–
		Urea	0.27 ± 0.03 a	6.62 ± 3.21 b
		Urea + 0.5%BC	0.28 ± 0.02 a	9.09 ± 2.72 ab
		Urea + 1.5%BC	0.29 ± 0.03 a	11.8 ± 1.21 a
	Zhongkejiayou12-6	Control	0.21 ± 0.03 b	–
		Urea	0.29 ± 0.04 a	12.1 ± 7.10 a
		Urea + 0.5%BC	0.27 ± 0.02 a	10.7 ± 6.18 a
		Urea + 1.5%BC	0.29 ± 0.02 a	11.7 ± 1.99 a
Haplic Acrisol	Xiushui134	Control	0.15 ± 0.01 c	–
		Urea	0.21 ± 0.01 b	4.89 ± 0.84 b
		Urea + 0.5%BC	0.23 ± 0.02 ab	5.39 ± 2.25 ab
		Urea + 1.5%BC	0.25 ± 0.02 a	9.36 ± 2.43 a
	Zhongkejiayou12-6	Control	0.19 ± 0.03 b	–
		Urea	0.34 ± 0.04 a	18.2 ± 5.34 a
		Urea + 0.5%BC	0.32 ± 0.04 a	16.5 ± 3.37 a
		Urea + 1.5%BC	0.29 ± 0.02 a	16.2 ± 3.30 a

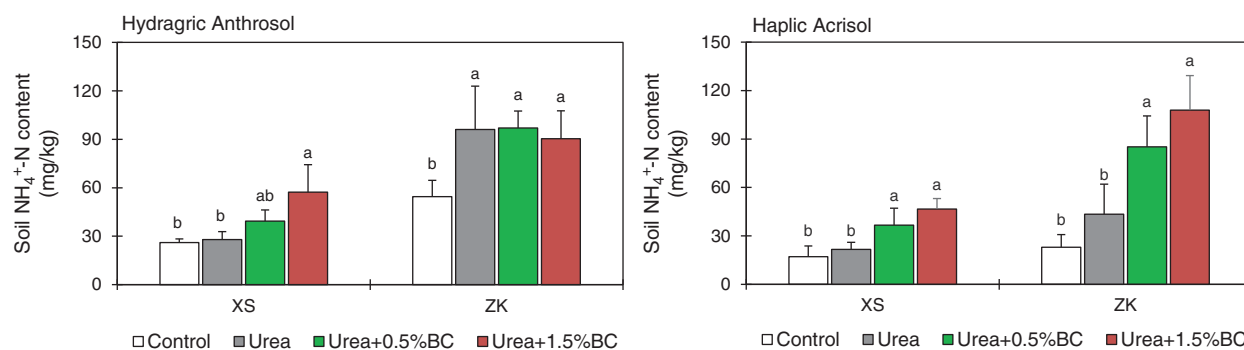
Note: Values shown are the means ± SD of three replicates. Different letters in same column for single rice variety indicate significant differences between each treatment within same soil type and rice variety according to Duncan's multiple range test at  $p < 0.05$ .

### 3.2 Soil pH and $\text{NH}_4^+$ -N Content

The soil pH and  $\text{NH}_4^+$ -N content after two-week seedlings harvested were presented in Figs. 1 and 2, respectively. Urea fertilizer input slightly increased soil pH, excepted for Haplic Acrisol planted with Xiushui134. Overall, soil pH increased following the application of biochar, particularly with higher rate (1.5 wt%). When applied with 1.5%, biochar increased soil pH by 0.07–0.18 units, compared to the urea only treatments.



**Figure 1:** Impacts of biochar addition on soil pH at rice seedling stage. XS and ZK referred to Xiushui134 and Zhongkejiayou12-6, respectively. Error bars represent the standard deviation (SD) for three replicates. Different letters indicate significant differences between each treatment within same rice variety according to Duncan's multiple range test at  $p < 0.05$



**Figure 2:** Impacts of biochar addition on soil NH<sub>4</sub><sup>+</sup>-N content at rice seedling stage. XS and ZK referred to Xiushui134 and Zhongkejiayou12-6, respectively. Error bars represent the standard deviation (SD) for three replicates. Different letters indicate significant differences between each treatment within same rice variety according to Duncan's multiple range test at  $p < 0.05$

Fig. 2 showed that NH<sub>4</sub><sup>+</sup>-N contents of two contrasting soil cultivated with Zhongkejiayou12-6 were relatively higher than that cultivated with Xiushui134. For Xiushui134 planted soils, urea N addition did not influence soil NH<sub>4</sub><sup>+</sup>-N content. However, soil NH<sub>4</sub><sup>+</sup>-N content increased significantly ( $p < 0.05$ ) by 76.3–88.7% following the N fertilization (Fig. 2). Except for Zhongkejiayou12-6 cultivated Hydragric Anthrosol, biochar application raised soil NH<sub>4</sub><sup>+</sup>-N contents by 40.9–148.6%, compared with Urea treatment. What is more, this improve effect was stronger when biochar was applied at higher rate (1.5 wt%).

### 3.3 Root Morphology Indexes

Impacts of biochar addition on rice seedling planted in two contrasting soils were presented in Tab. 3. For Xiushui134 in Hydragric Anthrosol, we found that biochar addition promoted the seedling root morphology, with longer root length (+87.6–88.2%), larger surface area (+58.9–64.3%), higher root volume (+38.5–46.2%), and more root tips (+67.9–69.4%), compared with the Urea treatment. This effect was also confirmed by Zhongkejiayou12-6 in Hydragric Anthrosol and Xiushui134 in Haplic Acrisol (Tab. 3). The improvement effect of biochar on seedling root was independent with biochar applied rate for two rice varieties in Hydragric Anthrosol, but not for Xiushui134 in Haplic Acrisol, which had best root morphology when biochar applied with a relative higher rate (1.5 wt%).

**Table 3:** Root total length, surface area, root volume, average diameter and root tips of Xiushui134 and Zhongkejiayou12-6 seedlings planted in Hydragric Anthrosol and Haplic Acrisol with biochar-added

Soil type	Rice variety	Treatment	Root length (cm)	Root surface area (cm <sup>2</sup> )	Root volume (cm <sup>3</sup> )	Average diameter (cm)	Root tips
Hydragric Anthrosol	Xiushui134	Control	176.3 ± 33.3 b	16.7 ± 2.2 b	0.13 ± 0.01 b	0.30 ± 0.02 a	2068 ± 379 b
		Urea	168.8 ± 56.5 b	16.8 ± 3.7 b	0.13 ± 0.01 b	0.33 ± 0.05 a	1940 ± 726 b
		Urea + 0.5%BC	317.7 ± 44.7 a	27.6 ± 3.2 a	0.19 ± 0.02 a	0.28 ± 0.01 a	3258 ± 185 a
		Urea + 1.5%BC	316.6 ± 74.4 a	26.7 ± 4.2 a	0.18 ± 0.02 a	0.27 ± 0.02 a	3287 ± 842 a
	Zhongkejiayou12-6	Control	252.6 ± 35.0 c	24.4 ± 2.3 b	0.19 ± 0.01 c	0.30 ± 0.02 a	2554 ± 581 b
		Urea	346.7 ± 25.4 b	31.9 ± 9.2 b	0.23 ± 0.06 bc	0.30 ± 0.04 a	3054 ± 509 b
		Urea + 0.5%BC	672.3 ± 53.7 a	51.3 ± 8.1 a	0.31 ± 0.07 ab	0.24 ± 0.02 b	6161 ± 401 a
		Urea + 1.5%BC	739.9 ± 46.7 a	57.4 ± 8.3 a	0.35 ± 0.04 a	0.25 ± 0.02 b	6772 ± 727 a
Haplic Acrisol	Xiushui134	Control	194.1 ± 56.4 b	15.8 ± 3.6 b	0.10 ± 0.02 b	0.26 ± 0.02 ab	1820 ± 867 b
		Urea	219.0 ± 52.9 b	19.0 ± 3.5 b	0.13 ± 0.02 b	0.28 ± 0.02 a	2011 ± 274 b
		Urea + 0.5%BC	293.5 ± 84.9 b	21.6 ± 4.9 b	0.13 ± 0.03 b	0.24 ± 0.02 b	2491 ± 846 b
		Urea + 1.5%BC	422.0 ± 17.9 a	31.0 ± 0.6 a	0.18 ± 0.02 a	0.23 ± 0.01 b	4020 ± 732 a

(Continued)

**Table 3 (continued).**

Soil type	Rice variety	Treatment	Root length (cm)	Root surface area (cm <sup>2</sup> )	Root volume (cm <sup>3</sup> )	Average diameter (cm)	Root tips
	Zhongkejiayou12-6	Control	261.6 ± 50.4 c	23.3 ± 4.6 c	0.16 ± 0.04 c	0.28 ± 0.02 a	2500 ± 736 c
		Urea	876.9 ± 61.7 a	68.7 ± 4.4 a	0.43 ± 0.03 a	0.25 ± 0.00 b	6837 ± 678 a
		Urea + 0.5%BC	484.8 ± 72.9 b	38.0 ± 6.8 b	0.24 ± 0.05 b	0.25 ± 0.01 b	4758 ± 206 b
		Urea + 1.5%BC	430.1 ± 81.2 b	33.6 ± 2.6 b	0.21 ± 0.03 bc	0.25 ± 0.01 b	4414 ± 257 b

Note: Values shown are the means ± SD of three replicates. Different letters in same column for single rice variety indicate significant differences between each treatment under same soil type and rice variety according to Duncan's multiple range test at  $p < 0.05$ .

Nevertheless, the development of seedling root was likely retarded following the biochar amendment. Data in [Tab. 3](#) showed that 44.7–51.0% shorter root length, 44.7–51.1% smaller surface area, 44.2–51.2% lower root volume, and 30.4–35.4% less root tips were recorded under Zhongkejiayou12-6 in Haplic Acrisol added with biochar (both at 0.5 and 1.5 wt%) than that under their counterpart Urea treatment. Results in [Tab. 3](#) also suggested that the influences of biochar on average root diameter varied with rice variety and soil type.

#### 4 Discussion

China initiated a “super rice” breeding program in 1996, focusing on hybrid rice lines, with the goals of producing high-yield rice cultivars through morphological improvement and through the utilization of N fertilizer [5]. In the present study, as can be seen from [Tab. 2](#), biochar has different effects on NUE of the two rice varieties, hybrid super rice variety Zhongkejiayou12-6 indeed showed higher NUE than common rice variety Xiushui134. What is more, Zhongkejiayou12-6 developed better root traits than Xiushui134, regardless of soil type and biochar addition rate ([Tab. 3](#)).

Nitrogen is one of the key nutrient for rice, and its availability in soil profile or root rhizosphere can influence the plant root morphology, total N uptake, and the NUE [12]. It had been demonstrated that  $\text{NH}_4^+\text{-N}$  a preferred N source, and is the main form of N available to rice seedling [15]. Data in [Tab. 2](#) suggested that biochar addition, particularly when applied at higher rate (1.5 wt%), indeed enhanced rice seedling NUE. Improvement on soil physical properties likely explained the higher NUE of Xiushui134 in biochar-added soils [7,13]. Well-know, biochar always has a basic features (biochar tested in current work was with a pH of 9.51), which could increase soil pH, particularly under acidic Haplic Acrisol ([Fig. 1](#)). After been transplanted, seedlings are more sensitive to pH changes, so changes in soil pH may be benefit to seedling growth. What is more, biochar can inhibit the soil N nitrification process that more  $\text{NH}_4^+\text{-N}$  could be retained in soil [19,20]. In the present study, except for Zhongkejiayou12-6 in Hydragric Anthrosol, biochar-added soils recorded higher  $\text{NH}_4^+\text{-N}$  contents than that of only urea added soils ([Fig. 2](#)). Higher soil  $\text{NH}_4^+\text{-N}$  content provided more N available to youth rice seedling that exerted higher NUE. Moreover, the rich functional groups and large specific surface area of biochar can improve soil CEC [21], absorb more nutrient ions besides N [22]. However, this could not explain why biochar addition had no significant influence on NUE of Zhongkejiayou12-6, a hybrid super rice cultivar, which should be explored in future study.

The change of soil pH has a direct influence on the growth and development of crops, especially the root system [23,24]. That is to say, biochar significantly promoted the growth and development of rice root system during Xiushui134 seedling stage ([Tab. 3](#)), which is consistent with a previous report [16]. Similarly, studies have shown that biochar has a catalytic effect on other plants. For example, biochar increased the root growth of citrus rootstock by 50% and the roots prefer to grow around biochar particles [23]. Miller et al. [24] also reported that roots preferentially grow around biochar to extend the rhizosphere. Therefore, the root system of the crop can extend to capture more N in the biochar-added soil. Nevertheless, biochar retarded the



Zhongkejiayou12-6 root growth in acidic Haplic Acrisol, though had not exert negative effect on NUE of seedling (Tabs. 2 and 3). Thus, the long-term effect and the underlying mechanisms of biochar on root morphology of varied rice cultivars in contrasting soils should be studied in future.

## 5 Conclusion

A soil pot experiment was conducted to observe the effects of biochar application with different rates on NUE, root morphology of rice seedling and the underlying relationship considering varied soil type and rice cultivars. Common rice Xiuhui134's NUE was increased after biochar addition, which effect was significant in higher rate of biochar-added two soils. The enhanced NUE of Xiuhui134 was likely attributing to the improved soil pH, increased soil  $\text{NH}_4^+$ -N content, and developed seedling root morphology. However, biochar addition had no impact on seedling NUE of super rice Zhongkejiayou12-6, which could not be thoroughly explained by the presently observed index. This phenomenon should be further explored in future study. In conclusion, biochar amendment indeed influence the soil pH and  $\text{NH}_4^+$ -N content, and the seedling root morphology and possibly impact the seedling NUE, which effects are dependent on rice cultivar, soil type and biochar application rate.

**Funding Statement:** This research is funded by the National Natural Science Foundation of China (31972518), and the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD).

**Conflicts of Interest:** The authors declare that they have no conflicts of interest to report regarding the present study.

## References

1. Kopittke, P. M., Menzies, N. W., Wang, P., Mckenna, B. A., Lombi, E. (2019). Soil and the intensification of agriculture for global food security. *Environment International*, 132, 105078. DOI 10.1016/j.envint.2019.105078.
2. Pratiwi, E. P. A., Shinogi, Y. (2016). Endita Prima Ari and Yoshiyuki Shinogi, Rice husk biochar application to paddy soil and its effects on soil physical properties, plant growth, and methane emission. *Paddy and Water Environment*, 14(4), 521–532. DOI 10.1007/s10333-015-0521-z.
3. Sun, H. J., Dan, A., Feng, Y. F., Vithanage, M., Mandal, S. et al. (2019). Floating duckweed mitigated ammonia volatilization and increased grain yield and nitrogen use efficiency of rice in biochar amended paddy soils. *Chemosphere*, 237, 124532. DOI 10.1016/j.chemosphere.2019.124532.
4. Kamau, S., Karanja, N. K., Ayue, F. O., Lehmann, J. (2019). Short-term influence of biochar and fertilizer-biochar blends on soil nutrients, fauna and maize growth. *Biology and Fertility of Soils*, 55(7), 661–673. DOI 10.1007/s00374-019-01381-8.
5. Chen, M., Chen, G., Di, D. W., Kronzucker, H. J., Shi, W. M. (2020). Higher nitrogen use efficiency in hybrid super rice links to improved morphological and physiological traits in seedling roots. *Journal of Plant Physiology*, 251, 153191. DOI 10.1016/j.jplph.2020.153191.
6. Ju, C. X., Buresh, R. J., Wang, Z. Q., Zhang, H., Liu, L. J. et al. (2015). Root and shoot traits for rice varieties with higher grain yield and higher nitrogen use efficiency at lower nitrogen rates application. *Field Crops Research*, 175, 47–55. DOI 10.1016/j.fcr.2015.02.007.
7. Chu, Q. N., Xu, S., Xue, L. H., Liu, Y., Feng, Y. F. et al. (2020). Bentonite hydrochar composites mitigate ammonia volatilization from paddy soil and improve nitrogen use efficiency. *Science of the Total Environment*, 718, 137301. DOI 10.1016/j.scitotenv.2020.137301.
8. Chu, L., Hennayake, D. H. M. K., Sun, H. J. (2019). Biochar effectively reduces ammonia volatilization from nitrogen-applied soils in tea and bamboo plantations. *Phyton-International Journal of Experimental Botany*, 88 (3), 261–267.
9. Yu, S., Xue, L. H., Feng, Y. F., Liu, Y., Song, Z. Z. et al. (2020). Hydrochar reduced  $\text{NH}_3$  volatilization from rice paddy soil: Microbial-aging rather than water-washing is recommended before application. *Journal of Cleaner Production*, 268, 122233. DOI 10.1016/j.jclepro.2020.122233.

10. Chu, Q. N., Xue, L. H., Cheng, Y. Q., Liu, Y., Feng, Y. F. et al. (2020). Microalgae-derived hydrochar application on rice paddy soil: Higher rice yield but increased gaseous nitrogen loss. *Science of the Total Environment*, 717, 137127. DOI 10.1016/j.scitotenv.2020.137127.
11. Yu, H. W., Zou, W. X., Chen, J. J., Chen, H., Yu, Z. B. et al. (2019). Biochar amendment improves crop production in problem soils: A review. *Journal of Environmental Management*, 232, 8–21. DOI 10.1016/j.jenvman.2018.10.117.
12. Drescher, G. L., da Silva, L. S., Sarfaraz, Q., Roberts, T. L., Nicoloso, F. T. et al. (2020). Available nitrogen in paddy soils depth: Influence on rice root morphology and plant nutrition. *Journal of Soils and Sediments*, 20, 1029–1041.
13. Sun, C. X., Chen, X., Cao, M. M., Li, M. Q., Zhang, Y. L. (2017). Growth and metabolic responses of maize roots to straw biochar application at different rates. *Plant and Soil*, 416(1–2), 487–502. DOI 10.1007/s11104-017-3229-6.
14. Xiang, Y. Z., Deng, Q., Duan, H. L., Guo, Y. (2017). Effects of biochar application on root traits: A meta-analysis. *Global Change Biology Bioenergy*, 9(10), 1563–1572. DOI 10.1111/gcbb.12449.
15. Britto, D. T., Kronzucker, H. J. (2002).  $\text{NH}_4^+$  toxicity in higher plants: A critical review. *Journal of Plant Physiology*, 159(6), 567–584. DOI 10.1078/0176-1617-0774.
16. Razaq, M., Salahuddin, Shen, H. L., Sher, H., Zhang, P. (2017). Influence of biochar and nitrogen on fine root morphology, physiology, and chemistry of Acer mono. *Scientific Reports*, 7(1), 5367. DOI 10.1038/s41598-017-05721-2.
17. Ubaidillah, M., Faperta, M., Kim, K. M. (2019). Identification of phytohormone changes and its related genes under abiotic stresses in transgenic rice. *Biocell*, 43(3), 215–224. DOI 10.32604/biocell.2019.07549.
18. Gong, Z. T., Chen, Z. C., Zhang, G. L. (2003). World reference base for soil resources (WRB): Establishment and development. *Soils*, 35(4), 271–278.
19. Sun, H. J., Lu, H. Y., Chu, L., Shao, H. B., Shi, W. M. (2017). Biochar applied with appropriate rates can reduce N leaching, keep N retention and not increase  $\text{NH}_3$  volatilization in a coastal saline soil. *Science of the Total Environment*, 575, 820–825. DOI 10.1016/j.scitotenv.2016.09.137.
20. Glaser, B., Lehmann, J., Zech, W. (2002). Ameliorating physical and chemical properties of highly weathered soils in the tropics with charcoal-A review. *Biology and Fertility of Soils*, 35(4), 219–230. DOI 10.1007/s00374-002-0466-4.
21. Laird, D. A., Fleming, P., Davis, D. D., Horton, R., Wang, B. et al. (2010). Impact of biochar amendments on the quality of a typical Midwestern agricultural soil. *Geoderma*, 158(3–4), 443–449. DOI 10.1016/j.geoderma.2010.05.013.
22. Lehmann, J. (2007). A handful of carbon. *Nature*, 447(7141), 143–144. DOI 10.1038/447143a.
23. Głuszek, S., Paszt, L. S., Sumorok, B., Kozera, R. (2017). Biochar-rhizosphere interactions—A review. *Polish Journal of Microbiology*, 66(2), 151–161. DOI 10.5604/01.3001.0010.6288.
24. Duvall, M., Sohi, S. P., Prendergast-Miller, M. T. (2014). Biochar-root interactions are mediated by biochar nutrient content and impacts on soil nutrient availability. *European Journal of Soil Science*, 65, 73–185.