ORIGINAL ARTICLE

WILEY Congenital Heart Disease

Analysis of *DICER1* in familial and sporadic cases of transposition of the great arteries

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Abstract

Objective: We previously identified a pathogenic germline *DICER1* variant in a child with transposition of the great arteries who was a member of a family with DICER1 syndrome. In view of a report linking *Dicer1* knockout in murine cardiomyocytes to cardiac outflow defects, we investigated the involvement of DICER1 in transposition of the great arteries.

Design: We used Fluidigm access array followed by next generation sequencing to screen for variants in the coding exons, their exon/intron boundaries and the 3' untranslated region of *DICER1* in patient DNA.

Cases: Germline DNA was collected from 129 patients with either sporadic or familial forms of transposition of the great arteries from two sites in Australia and Italy.

Results: Most cases (85%) did not have any germline *DICER1* variants. In the remaining 15% of cases, we identified 16 previously reported variants (5 synonymous, 6 intronic, and 5 missense) and 2 novel variants (1 intronic and 1 missense). None of the identified variants were predicted to be pathogenic.

Conclusions: Here, we report that neither likely pathogenic nor pathogenic variants in *DICER1* appear to play a major role in transposition of the great arteries.

KEYWORDS

DICER1, fluidigm, next-generation sequencing, transposition of the great arteries

1 | INTRODUCTION

DICER1 is an endoribonuclease that plays an essential role in modulating the expression of genes by producing mature microRNAs (miRNA), which are small, single stranded RNA molecules that bind to and thereby inhibit target mRNAs. *DICER1*-related diseases are referred to collectively as DICER1 syndrome and result from germline pathogenic and likely pathogenic variants in individuals with rare childhood cancers such as: pleuropulmonary blastoma, cystic nephroma, Sertoli-Leydig cell tumor, embryonal rhabdomyosarcoma,

Abbreviations: CHD, congenital heart defect; miRNA, microRNAs; TGA, Transposition of the great arteries.

and other rare tumors.¹ Several years ago, we identified a pathogenic germline *DICER1* variant (c.2117-1G > A, in intron 13 at the junction with exon 14, predicted to result in p.Gly706Aspfs*8) in a child with transposition of the great arteries (TGA), associated with a bicuspid pulmonary valve, an atrial septal defect and a patent ductus arteriosus.¹ Later, at the age of 18, he developed a solitary nodule in the left lobe of the thyroid gland. Two years later, he was found to have further nodules and cysts in the same lobe. Other heterozygotes for this pathogenic variant in the family also had phenotypes consistent with DICER1 syndrome.¹

TGA is a cyanotic congenital heart defect (CHD) characterized by ventriculo-arterial discordance and represents 5% to 7% of CHD.² It is often accompanied by other structural changes that allow mixing of

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oxygenated and deoxygenated blood; although there have been studies looking for the genetic causes of TGA, data so far have been inconclusive.³ Saxena and Tabin previously reported cardiac outflow defects in mice with a conditional knock-out of Dicer in the developing murine heart.⁴ In light of this, observation of this phenotype in a patient with a pathogenic variant in *DICER1* prompted us to screen for *DICER1* variants in additional familial and sporadic cases with TGA.

2 | METHODS

We screened 129 germline DNA samples from children with sporadic (n = 91) or familial (n = 38) forms of TGA for *DICER1* variants using targeted capture with a Fluidigm access array followed by next-generation sequencing and confirmatory Sanger sequencing.⁵

Variants were visualized against read alignments using Integrative Genomic Viewer (IGV)₆ version 2.3 (Broad Institute, Cambridge, Massachusetts). The mean and median coverage calculations were done using GATK DepthOfCoverage (Broad Institute).⁷

Eighty-two cases were referred to our study from The Heart Centre for Children, The Children's Hospital at Westmead, Westmead, Australia, and forty-seven from the Department of Medical Genetics, Bambino Gesù Pediatric Hospital, Rome, Italy. Details of the cases studied are shown in Tables 1–3. All patients signed an Institutional Review Board-approved consent form to participate in the study.

3 | RESULTS

No DICER1 variants were detected in 110 cases (85%), of which 77 cases were sporadic and 33 cases were familial. Seventy-one of these sporadic TGA cases had malformation of outflow tracts and 6 had a functional single ventricle (Table 1). Of the 33 familial cases, 22 had malformation of outflow tracts including 3 pairs of cousins where one of the cousins had malformation of outflow tracts and the other had either tetralogy of Fallot, or atrial septal defect or ventricular septal defect, respectively; 6 had one functional ventricle, and 3 had heterotaxy (Table 2). Nineteen individuals carried one or more variants in DICER1 including 5 synonymous, 7 intronic and 6 missense variants. A list of the individual variants and the associated physical manifestation for each carrier, where known, is presented in Table 3. As was the case for individuals with no variants detected, most cases presented with malformation of outflow tracts. All samples were sequenced to at least 1000x median coverage (except one sample at 644x), and 90% of the samples at 2000x median coverage. The scaled CADD⁸ and REVEL⁹ scores are listed in Table 3.

| TGA clinical info | No. of cases | Extra-cardiac features | Genetic tests (other than DICER1 sequencing) |
|-------------------------|-----------------|--|--|
| Functional single | 1 | Asthma | None |
| ventricle $n = 6$ | 1 | Asplenia | Normal karvotype and FISH ^b |
| | 4 | None ^c | None |
| Malformatin of | 19 | None ^c | None |
| outflow tracts $n = 71$ | 1 | None ^c | Normal karyotype |
| | 1 | None ^c | Normal karyotype |
| | 3 | None ^c | Normal karyotype and FISH ^d |
| | 1 | Heterozygous for cystic fibrosis | None |
| | 1 | Congenital talipes equinovarus, divergent strabis- mus | None |
| | 1 | Capillary hemangioma on left neck and face | None |
| | 1 | Hip dysplasia | None |
| | 1 | Delay in gross motor skills | None |
| | 1 | Tongue tie, asthma, autosomal dominant polycystic kidney disease, mild speech delay | None |
| | 1 | Congenital asplenia, gut malrotation, bowel ob- struction secondary to adhesions | Agilent (Santa Clara, California) SurePrint G3 ISCA Targeted Microarray. No abnormality |
| | 1 | Delayed receptive and expressive language skills | Balanced reciprocal translocation involving chr 3 and 6. No FISH abnormality ^d |
| | 1 | Speech delay | Normal karyotype and FISH ^d |
| | 1 | Possible liver mass (intrahepatic) | Normal karyotype, normal FISH at the TUPLE1 (VCFS) locus |
| | 1 | Subarachnoid hemorrhage on left hemisphere with small right parietal white matter infarction. | Normal karyotype and FISH ^d |
| | 1 | Intermittent right eye exotropia | Normal karyotype and FISH ^b |
| | 35 | Not known | Not known |

TABLE 1DICER1 negative sporadica cases of TGA, n = 77

Abbreviations: FISH, fluorescent in situ hybridization; HIRA, histone regulator A; ISCA, International Standards for Cytogenomics Arrays; TBX1, T-Box1; TUPLE1, TUP1-like enhancer of split protein; VCFS, velocardiofacial syndrome; 22q11.2, long arm of chromosome 22.

^aSporadic TGA was defined as any individual without any family history of CHD.

^bAt the HIRA (VYSIS,^e TUPLE1) locus.

^cTGA cases with no extra-cardiac features could be referred to as isolated TGA.

^dAt the HIRA (VYSIS, TUPLE1) or TBX1 (22q11.2) loci.

^eAbbott Molecular, Des Plaines, Illinois.

TABLE 2 DICER1 negative familial^a cases of TGA, n = 33

| TGA clinical info | No. of cases | Extra-cardiac features | Genetic tests (other than DICER1 sequencing) |
|---|---|---|---|
| Functional single ventricle n = 6 | 2 1 1 1 1 | None ^b None ^b Annular pancreas, duodenal atresia Functional asplenia Cerebral palsy, jejunal, and sigmoid atresia, microcephaly | None Normal karyotype, normal FISH ^c Normal karyotype None None |
| Heterotaxy n = 3 | 1 1 1 | Central liver with left sided stomach and spleen. Malposition of the superior mesenteric vein. Hip dysplasia, congenital asplenia, duodenal atresia, tracheal stenosis Congenital asplenia, situs ambigous, midline liver, right sided stomach | None None Normal karyotype, normal FISH ^c Agilent SurePrint G3 Targeted Microarray—normal |
| Malformation of outflow tracts n = 22 | 7 1 1 1 1 1 1 1 1 3 1 3 ^{def} | None ^b None ^b Delayed speech development Delay in receptive and expressive language skills Delayed expressive language at 2 years Multiple intestinal atresia, situs inversus Hypoxic ischemic encephalopathy secondary to shoulder dystocia, global developmental de- lay Mild encephalopathy resulting from early acidosis and hypoxemia None ^b None ^b Iron deficiency anemia NA | None Normal karyotype, normal FISH at the HIRA (VCFS) locus None None Normal karyotype Normal karyotype, normal FISH at the TUPLE1 (VCFS) locus Normal karyotype, Normal FISH at the HIRA (VYSIS, TUPLE1) locus Agilent aCGH 60K targeted array—normal Normal karyotype, normal FISH ^c none NA |
| Tetralogy of Fallot | 1 ^e | NA | NA |
| Atrial septal defect | 1 ^f | NA | NA |

Abbreviations: aCGH, array comparative genomic hybridization; FISH, fluorescent in situ hybridization; HIRA, histone regulator A; NA, not available; TBX1, T-Box1; TUPLE1, TUP1-like enhancer of split protein1; VCFS, velocardiofacial syndrome; 22q11.2, long arm of chromosome 22. ^aFamilial TGA was defined as any individual with immediate or extended family history of CHD.

^bTGA cases with no extra-cardiac features could be referred to as isolated TGA.

^cAt the HIRA (VYSIS, TUPLE1) or TBX1 (22g11.2) loci.

^d, ^e, and ^f are 3 pairs of cousins (^done cousin with malformation of outflow tracts and the other cousin with ventricular septal defects [included in Table 3]).

We listed the extra-cardiac features in our TGA cases and the genetic testing done, where available, in Tables 1–3.

4 DISCUSSION

The full spectrum of phenotypes associated with pathogenic germline variants in *DICER1* is still being defined, and newly associated phenotypes such as pituitary blastoma⁵ and macrocephaly¹⁰ are still emerging. As such, it is important to fully explore all possible associations. Here, no likely pathogenic or pathogenic variants in *DICER1* were observed among 129 patients affected by TGA. Extra-cardiac malformations are common in patients with congenital heart disease.¹¹ Furthermore, 16.5% of total cardiovascular MRI studies in children (145 studies) have noncardiovascular findings, thus proving one estimate of the prevalence of comorbid conditions in pediatric cardiovascular disease.¹² We did not detect any evidence of the presence of DICER1 syndrome in the TGA patients (Tables 1–3).

Two of the variants observed (Table 3), c.307 + 13T > C and c.4886C > T, were novel intronic and missense variants, respectively. The c.4886C > T variant is predicted to result in a protein with an amino acid change from serine to leucine, (p.S1629L) at position 1629. The algorithms SIFT (Sorting Intolerant from Tolerant)¹³ and Polyphen 2 (Polymorphism Phenotyping-2)¹⁴ predicted this variant to be "tolerated and benign," respectively. The G93E variant along with the missense at codon Y1835S and T60I in the transcript are all classified as variants of unknown significance (VUS₁₅). The variants R1599Q and C1153Y are both predicted to be tolerated by SIFT and benign by Polyphen 2, their REVEL and CADD scaled scores are lower than 0.75 and 30, respectively. S1629L is a novel variant that is tolerated by SIFT and benign by Polyphen 2, respectively. Most predictions for the missense variants varied from possibly damaging to benign by Polyphen 2, but all variants identified were predicted to be tolerated by SIFT (Table 3). For the missense variants we considered REVEL scores lower than 0.75 and CADD scaled scores lower than 30 to be likely benign.¹⁶ In our

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| TABLE 3 DICER1 variants in TGA stur | dy | | | | | | | | |
|--|----------------|---|---|--|----------------|---------------------|----------------|---------------------------------------|---------------------------------------|
| Variant | | Clinical information/genetic tests | | | Prediction | ns | | | MAF |
| Missense $n = \delta$ | z | TGA (sporadic/familial) | Extra-cardiac features | Genetic tests | REVEL | CADD (scaled) | SIFT | Polyphen 2 | ExAC (MAF/ count) |
| c.1278A > G p.E426E rs878855242*** | 1^{a} | Malformation of outflow tracts (sporadic) | None ^b | None | NA | 0.352 | NA | NA | Not found |
| c.1935G > A p.P645P rs61751177 | 4 | Ventricular septal defect ^c (familial, $n = 1$) | NA | NA | NA | 16.49 | NA | NA | 0.0095/1148 |
| | | Malformation of outflow tracts (sporadic $n = 3$) | None ^b | None | AN | | | | |
| c.2337A > G p.T779T rs747210633 c.2718C > T p.R906R rs370692165 c.5145C > T p.L1715L rs139500905 | ~ ~ ~ | Malformation of outflow tracts (familial) Malformation of outflow tracts (sporadic) Malformation of outflow tracts (sporadic) | None ^b NA None ^b | None NA None | N N N N N N | 2.59 NA 11.22 | NA NA NA | N N N N N N N N N N N N N N N N N N N | 0.00003/4 0.00008/10 0.0015/179 |
| Intronic $n = 7$ | | | | | | | | | |
| c.307 + 13T > C | 1 ª | Malformation of outflow tracts (sporadic) | None ^b | None | NA | NA | NA | NA | Novel |
| c.574-5G > A rs368253792 c.1377-4T > G rs192490028 | | Malformation of outflow tracts (sporadic) Heterotaxy (sporadic) | hypokalemia mild congenital tracheomalacia, congenital intestinal | None None | A A Z | 1.77 4.98 | A N A | A A A Z | 0.00006/7 0.0033/401 |
| c.1377-4T>G rs192490028 | - | Malformation of outflow tracts (snoradic) | Mairotation | ٨٨ | AN | 4.98 | NA | NA | 0.0033/401 |
| c.2040 + 29T > C rs370866625 | 1 ^d | Malformation of outflow tracts (familial) | Seborrhoeic dermatitis, asthma | Translocation, normal FISH at HIRA (VVSIS TUPI F1) ^e | NA | 2.57 | NA | NA | 0.0002/19 |
| c.3093 + 149_3093 + 153delGTTTT rs575610432 | - | Malformation of outflow tracts (sporadic) | None ^b | None | NA | NA | NA | NA | 0.0008/4*** |
| c.5364 + 18C > T rs777415635 c.5527 + 19A > G rs765497219 | н н | Malformation of outflow tracts (sporadic) Malformation of outflow tracts (familial) | NA Seborrhoeic dermatitis, asthma | NA Translocation, normal FISH at HIRA (VYSIS, TUPLE1) ^e | NA | 1.16 1.16 | NA | AN AN | 0.00007/8 0.000008/1 |
| Missense n = 6 c.179C > T p.T60l rs587778228 | , , | Malformation of outflow tracts (familial) | None ^b | None | 0.32 | 15.46 | Tolerated | Benign | 0.00005/6 |
| c.z/8G > A p.G93E rs//6219930 | H | Malformation of outflow tracts (sporadic) | AN | NA | 0.39 | 23./ | lolerated | Possibly damaging | 0.00003/4 |
| c.3458G > A p.C1153Y rs762999390 | 1^{d} | Malformation of outflow tracts (familial) | Seborrhoeic dermatitis, asthma | Translocation, normal FISH at HIRA (VYSIS, TUPLE1) ^e | 0.085 | 7.8 | Tolerated | Benign | 0.000008/1 |
| c.4796G > A p.R1599Q rs569615549 | \leftarrow | Malformation of outflow tracts (sporadic) | Early onset occipital epilepsy (Panayiotopoulos syndrome), speech delay | Normal Karyotype, normal FISH at the HIRA (VYSIS, TU- PLE1) or TBX1 (22q11.2) loci | 0.046 | 20.8 | Tolerated | Benign | 0.0002/23 |
| | | | | | | | | | (Continues) |

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| Aissense n = 6 | z | TGA (sporadic/familial) | Extra-cardiac features | Genetic tests | REVEL | CADD (scaled) | SIFT | Polyphen 2 | ExAC (MAF/ count) |
| c.4886C > T p.S1629L | 1 | Malformation of outflow tracts (familial) | None ^b | None | NA | NA | Tolerated | Benign | novel |
| c.5504A > C p.Y1835S rs747510783 | 1 | Malformation of outflow tracts (sporadic) | NA | NA | 0.68 | 25.7 | Tolerated | Possibly damaging | 0.00006/7 |

of split Abbreviations: chr 4, chromosome 4; FISH, fluorescent in situ hybridization; HIRA, histone regulator A; MAF, minor allele frequency; NA, not available; TBX1, T-Box1; TUPLE1, TUP1-like enhancer protein1; VCFS, velocardiofacial syndrome; 22q11.2, long arm of chromosome 22.

Genomes. From 1000

^aIndividual with 2 variants.

³TGA cases with no extra-cardiac features could be referred to as isolated TGA 6

^cCousin of an individual with malformation of outflow tracts (Table

Satellited chromosome regarded as heteromorphism with no expected clinical effect. and Nucleolar Organising Region (NOR) show translocation of NOR to the long arm of chr4. FISH to 4q sub-telomere 3 varian with ¹Individual

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study, all missense variants found have a REVEL value lower than 0.75 and all CADD scaled scores are lower than 30 (Table 3). In addition, the ones with CADD scores above 15 had no other significant pathogenicity flags. Notably, we did not identify any variants predicted to result in a truncated protein. Thus far, most diseaseassociated germline pathogenic variants in DICER1 are predicted to truncate the protein.¹⁷ Our results suggest that TGA is not likely to be caused by DICER1 pathogenic variants in humans and that occurrence of TGA in our patient carrying a pathogenic germline DICER1 variant may have been coincidental.

We recognize that we may have missed deletions/duplications as we did not investigate our sample set for large duplications/deletions via multiplex ligation-dependent probe amplification (MLPA).

This study did not address the potential of finding deleterious variants in DICER1 in other tissues (somatic mosaicism).¹⁸

5 CONCLUSION

Here, we report that TGA does not appear to be part of the DICER1 syndrome. The genetics underlying predisposition to TGA remain enigmatic² and it is likely that whole genome approaches in a large series of cases will be required to identify causal variants and genetic modifiers.

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CONFLICT OF INTEREST STATEMENT

The authors declare that they have no conflict of interest.

AUTHOR CONTRIBUTIONS

The manuscript was reviewed and edited by all authors, who commented on and approved the final version. All authors read and approved the final manuscript.

Nelly Sabbaghian Analyzed and validated the results and wrote the manuscript.

William D. Foulkes co-wrote the manuscript with Nelly Sabbaghian and oversaw the study.

Maria C. Digilio provided the samples, Gillian M. Blue, and David S. Winlaw provided the samples and the results of the additional genetic tests and extra-cardiac features.

Timothée Revil provided help with the bioinformatics analyses.

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REFERENCES

- Foulkes WD, Bahubeshi A, Hamel N, et al. Extending the phenotypes associated with DICER1 mutations. *Hum Mutat.* 2011;32(12): 1381–1384.
- [2] De Luca A, Sarkozy A, Consoli F, et al. Familial transposition of the great arteries caused by multiple mutations in laterality genes. *Heart* (*British Cardiac Society*). 2010;96(9):673–677.
- [3] Muncke N, Jung C, Rudiger H, et al. Missense mutations and gene interruption in PROSIT240, a novel TRAP240-like gene, in patients with congenital heart defect (transposition of the great arteries). *Circulation*. 2003;108(23):2843–2850.
- [4] Saxena A, Tabin CJ. miRNA-processing enzyme Dicer is necessary for cardiac outflow tract alignment and chamber septation. Proc Natl Acad Sci U S A. 2010;107(1):87–91.
- [5] de Kock L, Sabbaghian N, Plourde F, et al. Pituitary blastoma: a pathognomonic feature of germ-line DICER1 mutations. *Acta Neuropathol.* 2014;128(1):111–122.
- [6] Robinson JT, Thorvaldsdottir H, Winckler W, et al. Integrative genomics viewer. Nat Biotechnol. 2011;29(1):24–26.
- [7] McKenna A, Hanna M, Banks E, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 2010;20(9):1297–1303.
- [8] Kircher M, Witten DM, Jain P, O'roak BJ, Cooper GM, Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet*. 2014;46(3):310–315.
- [9] Ioannidis NM, Rothstein JH, Pejaver V, et al. REVEL: an ensemble method for predicting the pathogenicity of rare missense variants. *Am J Hum Genet*. 2016;99(4):877–885.

- [10] Khan NE, Bauer AJ, Doros L, Schultz KA, Decastro RM, Harney LA, et al. Macrocephaly associated with the DICER1 syndrome. *Genet Med.* 2016;19(2):244–248.
- [11] Rosa RC, Rosa RF, Zen PR, Paskulin GA. Congenital heart defects and extracardiac malformations. *Rev Paul Pediatr.* 2013;31(2):243–251.
- [12] Mahani MG, Morani AC, Lu JC, et al. Non-cardiovascular findings in clinical cardiovascular magnetic resonance imaging in children. *Pediatr Radiol.* 2016;46(4):473–482.
- [13] Ng PC, Henikoff S. SIFT: predicting amino acid changes that affect protein function. Nucleic Acids Res. 2003;31(13):3812–3814.
- [14] Sunyaev SR, Eisenhaber F, Rodchenkov IV, Eisenhaber B, Tumanyan VG, Kuznetsov EN. PSIC: profile extraction from sequence alignments with position-specific counts of independent observations. *Prot Eng.* 1999;12(5):387–394.
- [15] Database resources of the National Center for Biotechnology Information. Nucleic Acids Res. 2016;44(D1):D7–19.
- [16] Kim J, Field A, Schultz KAP, Hill DA, Stewart DR. The prevalence of DICER1 pathogenic variation in population databases. Int J Cancer. 2017;141(10):2030–2036.
- [17] Foulkes WD, Priest JR, Duchaine TF. DICER1: mutations, micro-RNAs and mechanisms. *Nat Rev Cancer*. 2014;14(10):662–672.
- [18] de Kock L, Wang YC, Revil T, et al. High-sensitivity sequencing reveals multi-organ somatic mosaicism causing DICER1 syndrome. *J Med Genet.* 2016;53(1):43–52.

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