

# The $\beta$ 2-Adrenergic Receptor Is a Global Regulator of Wound Healing *In Vivo*

C. E. Pullar<sup>1</sup> and R. R. Isseroff<sup>1</sup>

## 1 Introduction

Wound healing is a complex process, requiring the coordinated, temporal orchestration of numerous processes, including galvanotaxis, to repair damaged tissue. Over the past five years, we have established a role for the  $\beta$ 2 adrenergic receptor ( $\beta$ 2-AR) in regulating skin wound healing by investigating the effect of  $\beta$ 2-AR activation and blockade on the cellular processes of both human keratinocytes (K) and dermal fibroblasts (DF) in vitro and on wound healing in ex-vivo human skin models (1-6). Our prior research suggests that an endogenous catecholamine/ $\beta$ 2-AR network might regulate wound healing in vivo. In this study, we established colonies of FVB  $\beta$ 2-AR  $+/+$  (wild-type, WT) and FVB  $\beta$ 2-AR  $-/-$  (knock-out, KO) mice to investigate the role of the  $\beta$ 2-AR in murine wound healing in vivo.

## 2 Materials and Methods

Murine K and DF were isolated from both WT and KO neonates and the effect of  $\beta$ 2-AR ligands on murine cell migration, ERK phosphorylation and galvanotaxis was studied as previously described (1-6). Two 6mm full-thickness wounds were created on the shaved backs of WT and KO mice, treated daily, topically, with gel alone or gel containing 0.1%  $\beta$ 2-AR agonist or antagonist ( $n = 5$ ). Wounds were photographed daily then excised after either 3 or 5 days, fixed, embedded and sectioned for both hematoxylin/eosin and smooth muscle  $\alpha$ -actin (SM $\alpha$ A) immunostaining.

## 3 Results

$\beta$ 2-AR activation decreased both ERK

phosphorylation and the migratory capacity of WT murine K, while blinding them to an applied electric field (EF).  $\beta$ 2-AR blockade increased ERK phosphorylation, migration rate and galvanotaxis in WT murine K. KO murine K did not respond to  $\beta$ 2-AR ligands but migrated faster and more directionally in an EF than their WT counterparts. In contrast, both  $\beta$ 2-AR activation and blockade increased ERK phosphorylation and speed of migration in WT murine DF, while KO murine DF did not respond to  $\beta$ 2-AR ligands but migrated faster than WT. Finally, we detected catecholamine synthesis enzymes and measured epinephrine in both WT and KO K extracts.

$\beta$ 2-AR agonist treatment delayed while  $\beta$ 2-AR antagonist treatment or loss of the  $\beta$ 2-AR accelerated wound closure.  $\beta$ 2-AR activation decreased wound re-epithelialization by 15% while blockade or loss of receptor increased re-epithelialization by 30% in wounds excised 5 days post wounding. Wound contraction correlates with the appearance of Sm $\alpha$ A expressing myofibroblasts in the granulation tissue. WT mice exhibited dense staining for Sm $\alpha$ A within the dermis below the hyperproliferative epithelial wound margin. SM $\alpha$ A staining was markedly reduced in  $\beta$ -AR agonist-treated WT wounds (by 62%) and markedly increased in  $\beta$ 2-AR antagonist-treated WT wounds (2.2 fold) and KO wounds (27%).

## 4 Conclusion

In summary, we have demonstrated that  $\beta$ 2-AR agonist treatment significantly delays, while  $\beta$ 2-AR antagonist treatment significantly augments wound healing in vivo by modulating both re-epithelialization (K) and wound contraction (DF). Additionally, the fact that wound healing is

<sup>1</sup>Department of Dermatology, University of California, Davis: Davis California 95616, USA

accelerated in  $\beta$ 2-AR KO mice provides convincing evidence that the  $\beta$ 2-AR/catecholamine network regulates the rate of wound healing. Our work demonstrates that  $\beta$ 2-AR blockade could be a potential therapy for promoting healing in chronic wounds.

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### References

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