Pseudogene HMGN2P46 as a microRNA sponge to regulate HMGN2 expression via competing for miR-590-3p in severe acute pancreatitis

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Abstract: HMGN2 have functions in inflammatory response. However, the role of HMGN2 in severe acute pancreatitis (SAP) remains unclear. Here, our study was to discuss the role and regulatory mechanism of HMGN2 in SAP. In this study, the SAP cell model of AR42J was used to study the function and mechanism of HMGN2 in SAP. The protein expression in cells and serums were examined by western blot and ELISA assay. qPCR was used to test the transcriptional RNA level. Cell viability were examined by MTT assay. Luciferase assay was used to evaluate the interaction between gene and gene. Our results showed that HMGN2 was significantly upregulated in SAP patients. The database predicted and luciferase assay data indicated the HMGN2 was directly binding with miR-590-3p. ELISA, MTT and western blot experiments showed that the HMGN2 were promoted the cell proliferation, reduced the inflammation, and repressed the cell autophagy. Mechanism studies showed that the pseudogene HMGN2P46 level was positively correlated with HMGN2 and upregulated HMGN2 expression by competing for miR-590-3p in SAP. Taken together, all over these results showed upregulation of HMGN2 alleviates SAP, this process was regulated by HMGN2P46 competitively binding with miR-590-3p, which may provide a new insight for the treatment and intervention in SAP. Pseudogene HMGN2P46 was a miRNA sponge to regulate HMGN2 level by competing for miR-590-3p to alleviate the process of SAP. It provided a novel strategy for the diagnosis and treatment of severe acute pancreatitis.

Introduction

Severe acute pancreatitis (SAP) is a subtype of acute pancreatitis with multiple organ failure and systemic inflammatory response syndrome (Pezzilli *et al.*, 2015). At present, the incidence of acute pancreatitis continues to increase worldwide, and it is one of the most common gastrointestinal causes of hospitalization in China. Treatment for SAP includes intravenous fluid resuscitation, feeding, prophylactic antibiotics, probiotics, and endoscopic retrograde cholangiopancreatography (ERCP). In addition, interventions such as abstinence from alcohol and smoking cessation are important to reduce readmissions or prevent pancreatitis (Greenberg *et al.*, 2016; Zerem, 2014). Despite improvements in the treatment of SAP, disease-related mortality rates are still as high as 30%–40% (Garg and Singh, 2019). Therefore, the pathogenesis of SAP has been the focus

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of much research. Current studies suggest that oxidative stress, excessive release of pro-inflammatory cytokines, calcium overload, and dysregulation of autophagy are the main mechanisms that cause disease pathology (Gukovskaya *et al.*, 2017; Mandalia *et al.*, 2018).

HMGN2 is a gene which belongs to the high mobility group chromosome of the histone (HMG) members of the family and is the most abundant non-histone family in the nucleus of vertebrates and invertebrates. Previous studies have suggested that HMGN2 is related to DNA replication and transcriptional activity. Current studies have shown that HMGN2 plays a certain role in mouse embryo transfer (Li *et al.*, 2017a), M2 cell polarization (Wang *et al.*, 2019), and stem cell fate (Garza-Manero *et al.*, 2019). In addition, HMGN2 has a good antibacterial (Li *et al.*, 2017b), antiinflammatory (Deng *et al.*, 2012) and anticancer activities (Chen *et al.*, 2018; Su *et al.*, 2014). However, the role of HMGN2 in SAP is still unclear.

In recent years, non-coding RNA has received great attention in describing the molecular pathogenesis of SAP. Among them, microRNAs (miRNAs) are evolutionarily





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conserved pleiotropic small RNAs that inhibit gene expression after transcription. MiRNAs lead to translation inhibition of a large number of genes by sequence-specific binding to the 3'untranslated regions (UTRs) of homologous mRNAs (Pu *et al.*, 2019). It is reported that miR-590-3p is an interesting gene that play an exactly opposite effect in different cancers (Ge and Gong, 2017; Salem *et al.*, 2018) and also have function in inflammation injury (Liu *et al.*, 2020b). But there have been no related reports in SAP.

Similar to miRNAs, pseudogenes belong to another class of non-coding RNAs, defined as genomic elements that resemble real genes. Pseudogenes are common and abundant in the genome. They originate from the decay of repeated genes in the process of evolution, and are similar to functional genes, but have defects such as termination codon, frameshift and deletion in the coding sequence. Pseudogenes were previously considered biologically insignificant because they retain mutations and stop codons. However, nucleotide sequences within pseudogenes may be conserved due to evolutionary selection pressures, suggesting that they may indeed have an important role (Zheng et al., 2018). Pseudogenes were expressed tissuespecific and abnormally activated in some diseases and malignancies (Grandér and Johnsson, 2015; Tutar et al., 2018), suggesting that may be involved in some diseases and malignancies, although the mechanism remains unclear. It has been shown to pseudogenes act as affinity endogenous RNAs that regulate other RNA transcripts through affinity shared miRNAs (An et al., 2017). Interestingly, the frequency of pseudogenes such as HMG, OCT4 and Nanog was higher than expected (Lum et al., 2000; Pain et al., 2005), suggesting that it may be involved in the coordination and maintenance of related functions of their parent genes. Liu and his teams reported that pseudogene HMGA1P6 acted as a competitive endogenous RNA to promote ovarian cancer cell malignancy by upregulating HMGA1 and HMGA2 expression (Tian et al., 2020). de Martino et al. (2020) showed HMGA1P7 promoted B-cell lymphoma in mice by regulating HMGA1 and other cancer-related genes through miRNA. HMGN2P46 belongs to pseudogenes of HMGN2, but the functions also remain unknown.

In this study, we reported that HMGN2 and its pseudogene HMGN2P46 were abnormally expressed in SAP patient serum. At the same times, the expression levels of the two genes are directly related. In addition, we found that as a competitive endogenous RNA, HMGN2P46 can protect HMGN2 transcription from inhibition by miR-590-3p to play a protective role in the SAP process. This study provides the first evidence of a remarkable interaction among HMGN2, miR-590-3p, and HMGN2P46 and offered a new sight on the treatment of SAP.

Materials and Methods

Patients' serum samples

10 pairs SAP patient serum and normal serum samples were collected in our hospital from 2017 to 2020. All of SAP samples were provided by an average age of 41.3 years. Normal serum samples were taken from 12 normal volunteers which have a 33.1-year average age. All the serum samples were centrifuged at 4°C, stored in a -80° C ultra-low

temperature refrigerator. Each patient signed informed consent and ethics committee documents of Ethics Committee of our hospital to approve this study.

Cell culture, transfection, and SAP model

AR42Jcell was maintained with 15% FBS (Hyclone, USA), 100 µg/mL penicillin/streptomycin (Thermo, USA) at 37°C in a CO_2 incubator (5% CO_2 and 95% air, 95% humidity). The cell was cultured by passage after fusion at 70%-80%. WT/mut HMNG2, WT/mut HMNG2P46, sh-HMGN2, sh-HMGN2P46 and miR-590-3p mimic were designed, synthesized, and encapsulated by company (RiboBio, Guangzhou China). AR42Jcell was transfected with lentivirus plasmid and collected at 72 h after transfection. The AR42J SAP cell model in vitro was stimulated by lipopolysaccharide (LPS) in different concentrations (0, 1, 10, 100 mg/L) and different times (0, 6, 12, 18, and 24 h). The TNF-a level, cell survival and apoptosis were detected by Western blot and MTT assay. No LPS condition cell was used as control. sh-HMGN2 and sh-HMGN2P46 sense Oligonucleotide sequence showed in Table 1.

Enzyme linked immunosorbent assay (ELISA)

The ELISA assay was used to detect the HMGN2 level in SAP patient serums. The quantification of HMGN2 was assayed by Human HMGN2 ELISA Kit (No. JL38663, Jianglaibio, Shanghai China) according to the manufacturer's instructions. OD values were measured by microplate at the wavelength of 450 nm.

MTT assay

The proliferation of AR42J cells by LPS treatment or plasmid transfection were detected by MTT assay. Firstly, the treated experimental cells were seeded and incubated in 96-well plates at density of $5-8 \times 10^3$ cells/well and detected at 12, 24, 48, and 72 h, respectively. Then MTT solution (Thermo, USA) was added and incubated with the cells (dark), 150 µL DMSO (Sigma) was added after 4 h. Finally, the plates were shaken, and the absorbance was detected at 490 nm by a microplate reader (Gene, Hong Kong) after blue formazan crystals dissolved. The cell survival rate (%) = OD (experimental group)/OD (blank control group) × 100%. The experiment was repeated three times.

Real-time qPCR

The total RNA was extracted from patient serums and treatment cells by using TRIzol reagent (Thermo, USA) according to the manufacturer's information. Then the cDNA was synthesized by SuperScript VILOTM cDNA kit (Thermo, USA) following the manufacturer's manual. The reaction system and conditions of qPCR was referred to SYBRTM Green PCR Supermix kit instructions (Thermo, USA). The primers of HMGN2, HMGN2P46, miR-590-3p, and β -actin were shown in Table 2, β -actin as an internal control.

Western blot analysis

The total protein was extracted from SAP patient serums and LPS-treated cells. The concentration of protein was quantified by PierceTM BCA assay kit (Thermo, USA). In western blot analysis process, the total protein (30 μ g) was separated by 10% SDS-PAGE Gel. Then the separated proteins were

TABLE 1

Sense oligonucleotide sequence

| Gene name | Sense oligonucleotide sequence (5'-3') |
|-------------|--|
| sh-HMGN2 | CCGGCCCTGCAAAGAAGGGAGAGAACTCGAGTTCTCTCCCCTTCTTTGCAGGGTTTTT |
| sh-HMGN2P46 | CCGGGAAAGCTAAAGGTGCTGGAGACTCGAGTCTCCAGCACCTTTAGCTTTCTTT |

TABLE 2

Primer sequence

| Gene name | Sequence (5'-3') |
|------------|---|
| HMGN2 | Forward: CGATTGTCTGCCCATGTCCT |
| | Reverse: GCAGAACGTACCCTGTTCCA |
| HMGN2P46 | Forward: TCCGGATGTGACTCCTGTTA |
| | Reverse: TCTGGGACCTTGATGGCTCTA |
| miR-590-3p | Reverse transcriptional primer: CTCAACTGGTGTCGTGGAGTCGGCAATTCAGTTGAGCACTAGCTT |
| miR-590-3p | Forward: TCGGCAGGTAATTTTATGTAT |
| | Reverse: CTCAACTGGTGTCGTGGA |
| β-actin | Forward: GGAGATTACTGCCCTGGCTCCTA |
| | Reverse: GACTCATCGTACTCCTGCTTGCTG |

transferred to the PVDF membrane (Millipore, USA). The blots were incubated with HMGN2 (#9437, 1:500), TNF- α (#3707, 1:800), IL-10 (#12163, 1:1000), IL-6 (#12153, 1:1000), PARP1 (#9532, 1:800), cleaved caspase3 (#9664, 1:1000), Bcl2 (#15071, 1:1000), ULK1 (#6439, 1:1000), Beclin1 (#3738, 1:600), SQSTM1 (#88588, 1:900), and β -actin (#4970, 1:1000) (CST, USA) overnight at 4°C. The enhanced chemiluminescence detection kit (CST, USA) was used to visualize the target band after the second antibody incubation (goat anti-rabbit IgG 1:2000, CST, USA).

Luciferase reporter assay

The potential target miRNAs of HMGN2 were searched in Targetscan, StarBase and miRDB database website. The sequence of HMGN2 and HMGN2P46 with miR-590-3p binding sites were amplified by PCR and cloned to construct HMGN2, HMGN2P46 wild-type (WT) reporter vector, respectively. The mutant miR-590-3p binding sites was inserted into HMGN2, HMGN2P46 mutant-type (mut) reporter vector by site-directed mutagenesis (Thermo, USA), respectively. All the plasmid was constructed by company (RiboBio, Guangzhou, China). In the luciferase reporter assay process, the HEK293 or AR42J cells were cultured in 24-well plates ($3-5 \times 10^4$ cells/well) and transfected alone or co-transfected with the constructed reporter vector and miR-590-3p mimics. After transfected 48 h, the relative luciferase activity was detected by the Dual-Luciferase Reporter Assay System (Promega, USA).

Statistical analysis

All the data were analyzed by GraphPad Prism 6 (GraphPad Software, San Diego, CA USA). Two-way ANOVA, One-way ANOVA and Student's *t*-test were performed, and the results showed as mean \pm SD. *P* < 0.05 was considered to have statistical significance. All experiments repeated at least three times.

Results

HMGN2 is upregulated during SAP and targeted by miR-590-3p At the beginning, we detected and found the HMGN2 level was significantly increased in SAP patients' serum compared with normal serum (Figs. 1a and 1b) (P < 0.05). Besides, the bioinformatics analysis predicted that miR-590-3p may be the target gene of HMGN2 and the luciferase data showed HMGN2 was directly binding with miR-590-3p (Figs. 1c and 1d) (P < 0.05) and negatively regulated by miR-590-3p (Figs. 1e and 1f) (P < 0.05). These data suggested HMGN2 may play an important role in SAP and regulated by miR-590-3p.

HMGN2P46 level is positively correlated with HMGN2 in SAP Then we speculated that the expression level of miR-590-3p in SAP might be negatively correlated with HMGN2 because of HMGN2 directly targeting miR-590-3p. However, there was no significant correlation between miR-590-3p and HMGN2 protein level (r = 0.1367, P = 0.2368) (Fig. 2a), suggesting that there was other regulatory mechanism may be involved in HMGN2 expression. Studies have shown that pseudogenes play a role in many physiological and pathological processes and participate in the function of genes by regulating the mRNA expression of coding genes with similar sequences (Lum et al., 2000; Pain et al., 2005). In our study, whether there is a pseudogene of HMGN2 to regulate its expression. The corresponding pseudogenes of HMGN2 were searched by NCBI database and selected transcriptional sequences genes. It was found that HMGN2P46 was significant positively related to the expression of HMGN2 (r = 0.7217, P = 0.0001) (Fig. 2b).

HMGN2P46 is a processed pseudogene located at 15q21. It is highly homologous to the coding sequence of HMGN2. The



FIGURE 1. HMGN2 increased in SAP patients and regulated by miR-590-3p directly. (a–b) The HMGN2 protein levels in serum of SAP patients and normal serum were detected by ELISA and western blot assay. (c) The potential target miRNAs of HMGN2 in Venn diagram were predicted in Targetscan, StarBase and miRDB database. (d) Relative luciferase activity mediated by reporter constructs harboring the WT or mut 3'-UTR of HMGN2 upon transfection with 100 nMmiR-NC or miR-590-3p in HEK293 cell. (e–f) The HMGN2 level in HEK293 cell were detected by qRT-PCR and western blot assay. β -actin as an internal gene control. **P* < 0.05, ***P* < 0.01. Data are present the mean ± SD of three independent experiments.

HMGN2P46 lacked the HMGN2 3'-UTR sequence which binding with miR-590-3p, but actually the sequence variations of HMGN2P46 contribute to a new match with miR-590-3p (Fig. 2c) and the luciferase assay data was confirmed this combination (Fig. 2d). In addition, we used the LPS to induce the SAP model in AR42J cell line at different times and concentrations and the results showed the SAP cell model was success (Suppl. Fig. S1). Then we detected the HMGN2, HMGN2P46, and miR-590-3p levels in LPS-induced SAP cell model and the qPCR results showed that HMGN2, HMGN2P46 were upregulated, while miR-590-3p decreased after 10 mg/L LPS stimulated 24 h in AR42J cell (Fig. 2e) (P < 0.05). Besides, further qRT-PCR analysis confirmed miR-590-3p has only effect on the HMGN2P46 mRNA level, the HMGN2P46 level was remarkably repressed by miR-590-3p (Fig. 2f) (P < 0.05). All the data indicated that HMGN2P46 level is positively correlated with HMGN2 and regulated by miR0-590-3p in SAP.

HMGN2P46 upregulates HMGN2 expression by competing for miR-590-3p in SAP

In order to evaluate the role of HMGN2P46 in regulating HMGN2 expression level, we used the luciferase reporter



FIGURE 2. The HMGN2 levels have a significant correlation with HMGN2P46 in SAP. (a) Correlation between HMGN2 protein level and miR-590-3p level. (b) Correlation between HMGN2 protein level and HMGN2P46 level. (c) Bioinformatic analysis of HMGN2 and its pseudogene HMGN2P46. (d) Relative luciferase activity was detected after transfection with 100 nM miR-590-3p upon reporter constructs harboring the WT/mut-HMGN2P46. (e) The expression of HMGN2, HMGN2P46, and miR-590-3p levels in LPS-induced SAP cell model. (f) The HMGN2P46 levels in LPS-induced SAP cell model after transfection with 100 nM miR-590-3p. *P < 0.05, **P < 0.01. Data are present the mean \pm SD of three independent experiments.

assay which including wild-type HMGN2 3'-UTR and the results showed the miR-590-3p overexpression significantly downregulated the luciferase signal activity by bind with WT-HMGN2 3'-UTR in both HEK293 and AR42J cells, while WT-HMGN2P46 overexpression significantly increased the luciferase activity because of competing interaction with miR-590-3p to keep HMGN2 level but not mut-HMGN2P46 which unbinding with miR-590-3p (Figs. 3a and 3b) (P <0.05). Besides, the HMGN2P46 downregulation could decreased the luciferase activity, while anti-miR-590-3p reversed the effect of HMGN2P46 downregulation by binding with miR-590-3p (Figs. 3c and 3d) (P < 0.05). In addition, the western blot results showed that HMGN2P46 inhibition also downregulated the HMGN2 protein level, and anti-miR-590-3p overexpression rescued the effect of HMGN2P46 inhibition via interacting with miR-590-3p to keep more HMGN2 translation (Figs. 3e and 3f) (P < 0.05). All the results suggested that HMGN2P46 upregulated HMGN2 expression by competing interaction with miR-590-3p.

HMGN2 promotes the cell survival in SAP

To explore the effect of HMGN2 and HMGN2P46 on AR42J cell proliferation upon a SAP cell model, the cells were transfected with sh-control, sh-HMGN2, sh-HMGN2P46, sh-HMGN2 + sh-HMGN2P46, and sh-HMGN2P46 + anti-miR-590-3p knockdown plasmid and detected by MTT assay, respectively. The results showed both of HMGN2 and HMGN2P46 repression aggravated LPS-induced AR42J cell injury, while miR-590-3p inhibition could rescued this effect (Fig. 4a) (P <0.05). Besides, we assessed the effect on cell apoptosis protein levels by western blot and both of HMGN2 and HMGN2P46 downregulation increased SAP AR42J cell apoptosis, while miR-590-3p downregulation revered the further aggravation which caused by HMGN2P46 inhibition (Fig. 4b) (P < 0.05). Furthermore, only WT-HMGN2P46 could affected the HMGN2 and cell apoptosis key protein levels in SAP cell model but not mut-HMGN2P46 (Figs. 4c and 4d) (P < 0.05), indicating the HMGN2P46 as a decoy to interact with miR-590-3p for regulate HMGN2 level and cell proliferation. All



FIGURE 3. HMGN2P46 competitive binding with miR-590-3p to upregulate HMGN2 expression in SAP. (a–b) The luciferase activity results in both HEK293 and AR42J cells after transfected with control, miR-590-3p, WT-HMGN2P46 or mut-HMGN2P46, and WT-/mut-HMGN2P46 + miR-590-3p, respectively. (c) The mRNA levels of HMGN2 and HMGN2P46 in AR42J cell after HMGN2 and/or HMGN2P46 downregulation were detected by qPCR. (d) The luciferase activity in AR42J cells after transfected with sh-control, sh-HMGN2P46, and sh-HMGN2P46 + anti-miR-590-3p. (e–f) The western blot and statistical results of HMGN2 protein levels in SAP cell model after transfected with sh-control, sh-HMGN2P46, and sh-HMGN2P46, and sh-HMGN2P46 + anti-miR-590-3p. β -actin as an internal gene control. **P* < 0.05, ***P* < 0.01. Data are present the mean ± SD of three independent experiments.

data showed that HMGN2 was regulated by the combination HMGN2P46 and miR-590-3p to benefit AR42J cells survival from SAP injury.

HMGN2 reduces cell inflammation response in SAP

Excessive release of pro-inflammatory cytokines is one of the pathological mechanisms in SAP, so we detected the effect of HMGN2 on the inflammatory response after HMGN2 inhibition. The results showed that downregulation of HMGN2 or HMGN2P46 significantly repressed anti-inflammatory factor IL-10 level and increased pro-inflammatory cytokines TNF- α and IL-6 levels compared with the control, while the miR-590-3p inhibition significantly decreased the TNF- α and IL-6 levels and upregulated the IL-10 (Figs. 5a and 5b) (P < 0.05). In addition, only WT-HMGN2P46 could decreased the key

inflammation response factors TNF- α and IL-6 levels and upregulated the IL-10 level in SAP cell model (Figs. 5c and 5d) (P < 0.05). All the results indicated HMGN2P46 may reduce the release of inflammation factors as a decoy to interact with miR-590-3p for regulate HMGN2 level in SAP.

HMGN2 regulates the autophagy in SAP

Autophagy is one of the pathological mechanisms of SAP, so we also examined the changes of autophagy after HMGN2 inhibition. The results indicated that both HMGN2 and HMGN2P46 inhibition significantly decreased autophagy key factors such as ULK1, SQSTM1, and Beclin1 protein levels compared with the control, especially ULK1, while the downregulation of miR-590-3p significantly reversed this effect (Figs. 6a and 6b) (P < 0.05). Besides, only



FIGURE 4. The HMGN2 regulated the cell proliferation in SAP. (a) The cell proliferation upon a SAP cell model were detected by MTT assay after transfected with sh-control, sh-HMGN2, sh-HMGN2P46, sh-HMGN2 + sh-HMGN2P46, and sh-HMGN2P46 + anti-miR-590-3p knockdown plasmid. (b) The protein levels of PARP1, cleaved caspase3, and Bcl-2 were assessed by western blot after transfected with a series of knockdown plasmid. (c–d) The HMGN2, PARP1, cleaved caspase3, and Bcl-2 protein levels in SAP cell were detected by western blot after transfected with WT- or mut-HMGN2P46 plasmid. β -actin as an internal control. "*": compared with sh-control, " Δ ": compared with sh-HMGN2, " ϕ ": compared with sh-HMGN2P46, all mean *P* < 0.05. Data are present the mean ± SD of three independent experiments.



FIGURE 5. The HMGN2 affected inflammatory response in SAP cell model. (a-b) The inflammation response of AR42J cell with LPS-induced for 24 h after HMGN2, HMGN2P46, and miR-590-3p downregulation were detected by western blot. (c-d) The inflammatory related factors IL-10, TNF- α , and IL-6 protein levels in SAP cell model after transfected with WT- or mut-HMGN2P46 plasmid were evaluated by western blot. β -actin as an internal control. "*": compared with sh-control, " Δ ": compared with sh-HMGN2, "\phi": compared with sh-HMGN2P46, all mean P < 0.05. Data are present the mean ± SD of three independent experiments.

WT-HMGN2P46 reduced the autophagy factors ULK1, SQSTM1, and Beclin1 protein levels in SAP cell model (Figs. 6c and 6d) (P < 0.05). All the data suggested that HMGN2P46 involved the autophagy processas a decoy to compete with miR-590-3p to increase HMGN2 level in SAP.

Discussion

Studies have shown that pseudogenes were conserved in biological evolution, expressed tissue-specific and abnormally activated in partial diseases and cancers (Grandér and Johnsson, 2015; Tutar et al., 2018; Zheng et al., 2018), which means the pseudogenes may play an important role in the progress of some diseases and cancers. Tan et al. (2019) stated PIWI-36712 interacted with pseudogene SEPW1P to inhibit breast cancer progression and chemoresistance. Liu et al. (2020a) reported that AGPG pseudogene regulated cancer glycolytic process by targeting PFKFB3. Yu et al. (2019) showed that pseudogene POU5F1B was increased in cervical cancer and promoted cell proliferation and migration. In other diseases, Wang et al. reported that pseudogene PTENP1 regulated PTEN level to promote differentiation process of osteoclast and reduce the osteoporosis by targeting miR-214 (Wang et al., 2020). Our study focused on the function and mechanism of HMGN2 and its pseudogene HMGN2P46 in SAP.

In our research, HMGN2 upregulated significantly inpatient serum and AR42J cell model of SAP. Besides, HMGN2 directly binding with miR-590-3p, and the expression does not have much correlation with miR-590-3p level but positively correlated with its pseudogene HMGN2P46 in SAP patient serum. Furthermore, the downregulation of HMGN2 and HMGN2P46 significantly inhibited cell growth, promoted inflammatory response and autophagy process in SAP cell model. In molecular mechanisms, the functions of HMGN2P46 were achieved by regulating HMGN2 protein level, which competing interaction with miR-590-3p to increase HMGN2 protein level. Our molecular mechanisms, HMGN2P46 as a decoy competing with miR-590-3p to upregulate HMGN2 expression in SAP, are consistent with previous studies.

HMG family members, which including HMGA, HMGB, and HMGN, are play an important role in inflammation and infection of body. HMGA1 and HMGA2 as two driver of inflammation, aggravated LPS-induced cell inflammation and apoptosis by different pathway (Cai *et al.*, 2020; Li *et al.*, 2020). Besides, HMGB1, as a controlling target for inflammation, is a therapeutic target for severe pulmonary inflammation (Andersson *et al.*, 2020), sepsis and trauma-induced inflammation (Deng *et al.*, 2019). There are a few studies on the anti-inflammatory effects of HMGN. In our results, HMGN2 relieved inflammation in SAP by regulating pseudogene HMGN2P46/miR-590-3p axis, which provide a new sight of HMGN family on pathological mechanism and therapeutic targets in SAP.

Besides, autophagy process is also regulated by HMG family. HMGA1 inhibited autophagy to promote myocardial dysfunction in diabetic cardiomyopathy (Wu *et al.*, 2020). HMGA2 have function in Cr(VI)-induced autophagy (Yang *et al.*, 2017). HGMB1 is a critical regulator of autophagy (Tang *et al.*, 2010), but there are also few reports on autophagy function ofHMGN at present. In our study showed that HMGN2 was regulated by pseudogene HMGN2P46/miR-590-3p axis to promote cell autophagy in SAP. It adds autophagy regulation to HMGN2 and provide a new idea of HMG family on pathological mechanism in SAP.



FIGURE 6. The HMGN2 affected the autophagy process in SAP cell model. (a-b) The cell autophagyin SAP cell model after HMGN2, HMGN2P46, and miR-590-3p downregulation were evaluated by western blot. (c-d) The protein levels of autophagy-related factors ULK1, SQSTM1 and Beclin1 in SAP cell model after transfected with WT- or mut-HMGN2P46 plasmid were tested by western blot. $\beta\text{-actin}$ as an internal control. "*": compared with sh-control, " Δ ": compared with sh-HMGN2, "φ": compared with sh-HMGN2P46, all mean P < 0.05. Data are present the mean ± SD of three independent experiments.

MiRNAs are evolutionarily conserved small RNAs that inhibit gene expression after transcription. MiRNAs typically lead to translation inhibition or mRNA degradation by interacting with the 3'-UTR sequences of homologous mRNAs (Pu *et al.*, 2019). MiR-590-5p was always showed in many cancers (Ge and Gong, 2017; Salem *et al.*, 2018). Moreover, Ma *et al.* (2019) reported that miR-590-3p inhibited TNFRAF-6 to attenuate acute kidney injury in septic mice. MiR-590-3p also inhibited the LPS-induced sepsis apoptosis and inflammation (Liu *et al.*, 2020b). In our study, miR-590-3p as a sponge to regulate inflammatory response of SAP and we did not discuss the function of miR-590-3p in SAP need more experiment to confirm.

Taken together, HMGN2P46 as a decoy competing with miR-590-3p to upregulate HMGN2 expression in SAP, thus regulated cell viability, inflammation, and autophagy to attenuate the SAP injury. Our study provides a new insight for understanding HMGN2 roles in SAP and a new therapeutic target for SAP.

Availability of Data and Materials: All data generated or analysed during this study are included in this published articleand its supplementary information files.

Author Contribution: The authors confirm contribution to the paper as follows: study conception and design: Hongqiang Zhao, Jiangyang Lu; data collection: Hongqiang Zhao, Qi Liu; analysis and interpretation of results: Hongqiang Zhao, Qi Liu, Haorun Liu; draft manuscript preparation: Hongqiang Zhao, Weimin Li. All authors reviewed the results and approved the final version of the manuscript.

Ethics Approval: This study and all experiments involved are under approval of the Chinese PLA General Hospital.

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Conflicts of Interest: The authors declare that they have no conflicts of interest to report regarding the present study.

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SUPPLEMENTARY FIGURE S1. The cell model construction of severe acute pancreatitis *in vitro*. (a) The TNF- α protein levels in AR42J cell after stimulated by LPS in different concentrations (0, 1, 10, 100 mg/L) for 24 h were detected by Western blot. (b) The TNF- α protein levels in AR42J cell after stimulated by 10 mg/L LPS at different time (0, 6, 12, 18, and 24 h) were assessed by Western blot. (c) The AR42J cell proliferation after stimulated by 10 mg/L LPS treatment at different time was tested by MTT assay. (d) The apoptosis protein levels of PARP1, cleaved capase3, and Bcl-2 in AR42J cell after 10 mg/L LPS treatment were detected by Western blot. β -actin as a negative control. **P* < 0.05, ***P* < 0.01. Data are present the mean ± SD of three independent experiments.