

## Chilling Injury, Physicochemical Properties, and Antioxidant Enzyme Activities of Red Pitahaya (*Hylocereus polyrhizus*) Fruits under Cold Storage Stress

Kai Sheng<sup>1</sup>, Saichao Wei<sup>1</sup>, Jun Mei<sup>1,2,3,4,\*</sup> and Jing Xie<sup>1,2,3,4,\*</sup>

<sup>1</sup>College of Food Science and Technology, Shanghai Ocean University, Shanghai, 201306, China.

<sup>2</sup>National Experimental Teaching Demonstration Center for Food Science and Engineering Shanghai Ocean University, Shanghai, 201306, China

<sup>3</sup>Shanghai Engineering Research Center of Aquatic Product Processing and Preservation, Shanghai, 201306, China

<sup>4</sup>Shanghai Professional Technology Service Platform on Cold Chain Equipment Performance and Energy Saving Evaluation, Shanghai, 201306, China

\*Corresponding Authors: Jun Mei. Email: jmei@shou.edu.cn; Jing Xie. Email: jxie@shou.edu.cn

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**Abstract:** Low-temperature storage is extensively used to optimize the postharvest life of various fresh fruits. However, red pitahaya (*Hylocereus polyrhizus*) fruits are sensitive to chilling injury (CI), which leads to the limitation of low-temperature storage. In this study, red pitahaya fruits were stored at 2, 4, 6, 8, and 10°C, respectively, for 27 days to determine the appropriate storage temperature. During the storage of red pitahaya fruits, storage at 8°C was more effective in suppressing decay and maintaining quality than other low temperatures. Low-temperature (2, 4, and 6°C) storage decreased weight loss (WL) and maintained higher content of titratable acidity (TA), soluble sugars (SS), and total phenolics (TP) but different degrees of CI were detected. No CI was observed at 8°C and 10°C. Red pitahaya as stored at 8 and 10°C were associated with better color evaluation, lower electrolyte leakage (EL), respiration rate, and lipoxygenase (LOX) activity, and higher fruit firmness, superoxide dismutase (SOD) activity, and catalase (CAT) activity. However, higher storage temperature (10°C) resulted in higher metabolic activity leading to lower quality and antioxidant capacities compared with 8°C. Therefore, our results demonstrated that red pitahaya stored at 8°C exhibited a protective effect on fruit quality and resisted CI development during storage.

**Keywords:** Red pitahaya; quality; low temperature; chilling injury

### 1 Introduction

Red pitahayas (*Hylocereus polyrhizus*), originating from Latin America and the West Indies [1], have attracted much attention worldwide due to attractive appearance, high nutritional value, and various biological activities [2–4]. However, the fruits are highly perishable, mainly caused by fungal diseases, degreening, and physiological disorders [5,6], which shortens its postharvest shelf life during distribution and marketing. Since the major quantum of fruits undergoes for fresh consumption [7], adequate and



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sound preservation technology for fruits' quality maintenance and long-term storage are required for both the consumers and food processing markets.

Some methods preserving post-harvest red pitahaya fruits have been developed, such as Gamma irradiation [8], 1-methylcyclopropene treatment [5], and active package [9]. Nevertheless, these preservation technologies combined with low-temperature storage of fruits could have a more beneficial influence on maintaining fruit quality. Low-temperature storage can effectively delay fruit decay and prolong the storage period of red pitahayas. Obenland et al. [10] and Punitha et al. [11] reported that the fruits stored at 5°C or 6°C could maintain the quality and visual appearance better than those at higher temperatures. However, red pitahayas are sensitive to low temperatures such as 2°C resulting in CI, which limits the application of low-temperature storage [12,13].

CI is a suite of physiological defects that affects the marketability and storability of most tropical fruits [14,15]. In recent years, reports on horticultural products found that fruits responded to chilling temperatures mainly in two ways. On the one hand, the molecular level of CI affects bio-membrane conformation and structure [16], with an increase in electrolyte leakage and lipid peroxidation [17,18], causing a drastic fruit softening [19]. On the other hand, low-temperature storage may induce an uncoupling in the respiratory chain, giving rise to reactive oxygen species (ROS) [20] and the excessive accumulation of ROS can further poison cells and contribute to CI [21]. Yet, fruits can naturally protect themselves against oxidative damage by producing antioxidant enzymes, such as SOD and CAT [22].

Little information is available on the comprehensive evaluation of the physio-chemical changes concerning CI in red flesh pitahayas during cold storage. The object of this study was to investigate the physio-chemical changes in red pitahayas during cold storage at 2, 4, 6, 8, and 10°C and to determine optimum low-temperature without CI in red pitahayas.

## 2 Materials and Methods

### 2.1 Fruit and Treatment

Red pitahayas were purchased from a local supermarket. Fresh fruits that were free from disease and of uniform size ( $560 \pm 50$  g per fruit), shape, and color were randomly divided into five groups and stored at 2, 4, 6, 8, and 10°C ( $\pm 0.2^\circ\text{C}$ ), respectively. The relative humidity was set within the range of 70%–80%. The quality evaluation of red pitahaya samples was performed on 0, 6th, 12th, 15th, 18th, 21st, 24th, and 27th day. The samples were transferred to  $23 \pm 1^\circ\text{C}$  with 70% R.H. for 24 h before the quality determinations.

### 2.2 CI Evaluation

CI evaluation was done in the sensory assessment laboratory and controlled white lighting. In each session, 6 trained panelists tested 30 codified samples (6 fruits from each group). Each panelist was asked to rate the extent of the CI based on the percentage of pitting, browning, and shriveling [23] in the peel surface as follows: 0 = no injury, 0% of the area affected; 1 = slight injury, up to 25%; 2 = moderate injury, from 26% to 50%; 3 = severe injury, from 51% to 75%; and 4 = extremely severe injury, higher than 75%. Therefore, the Chilling injury index (CII) ranged from 0 to 4. The CII was expressed as follows:

$$\text{CII} = \frac{\text{CI level} \times \text{number of fruits at the CI level}}{4 \times \text{total number of fruits}} \quad (1)$$

### 2.3 Color Evaluation

Lightness (L), chroma (C), and hue angle (H) were evaluated on four locations around the equatorial plane of the fruit peel using a colorimeter (CR-400, Konica Minolta Investment Ltd., Shanghai, China).

#### **2.4 Weight Loss (WL)**

The WL of red pitahaya was recorded before storage and after storage. The WL was expressed as the percentage of fruit weight.

#### **2.5 Fruit Firmness**

Fruit firmness was determined in the equatorial region of the fruits using a penetrometer (GY-4, Yueqing Handpi Instruments Co., Ltd., Zhejiang, China) with a 3-mm diameter tip and the results were expressed in Newton (N).

#### **2.6 Determination of EL**

EL was measured according to Ruiling et al. [5] with some modifications. Cylindrical fruits (6 mm in diameter and 2 mm thick) were rinsed with deionized water and incubated in distilled water at 20°C for 1 h to determine the initial EL ( $E_0$ ). The samples were boiled in water for 30 min, cooled at room temperature, and the total conductivity ( $E_t$ ) was measured. The rate of EL was calculated as follows:

$$EL = \frac{E_0}{E_t} \times 100\% \quad (2)$$

#### **2.7 Respiration Rate**

Respiration rate was estimated by the method of Deng et al. [24] with some modifications. One fruit was sealed in one glass container that contained 10 mL 0.4 mol/L NaOH at room temperature for 30 min. Then, 5 mL saturated BaCl<sub>2</sub> and two drops of phenolphthalein were added, and the solution was titrated with 0.1 mol/L oxalic acid until the endpoint. The respiration rate of the samples was expressed as mg·kg<sup>-1</sup>·h<sup>-1</sup> CO<sub>2</sub>.

#### **2.8 Titratable Acidity (TA)**

TA was calculated based on malic acid as the predominant acid and was expressed as percent (w/v) of malic acid at 25°C. Measurements were performed in triplicate.

#### **2.9 Soluble Sugars (SS)**

SS was determined according to the method by Zhang et al. [25] with some modifications. 1 g sample was homogenized with 10 mL distilled water. The slurry was centrifuged at 4 000 rpm for 10 min at 4°C. The supernatant was quantitative to 25 mL and used for SS analysis. Results were expressed as a percentage of glucose.

#### **2.10 Total Phenolics (TP)**

TP was measured with Folin-Ciocalteu reagent according to Ruiling et al. [5] with some modifications. Samples (2 g) were homogenized with 5 mL of pre-cooled methanol (80%, v/v) in darkness at 4°C. After centrifugation, the supernatant (1 mL) was added to the reaction solution consisted of 4 mL of 7.5% (w/v) Na<sub>2</sub>CO<sub>3</sub> and 5 mL of Folin-Ciocalteu reagent. Then the reaction mixture was incubated in darkness at 25°C for 1 h. The absorbance at 765 nm was measured using a spectrophotometer (UV-2100, Unico (Shanghai) instrument Co., Ltd, China). The results were calculated from a standard curve of gallic acid and expressed as gallic acid equivalents (GAE) based on fresh weight basis (g·kg<sup>-1</sup>).

#### **2.11 Enzyme Assays**

Frozen pulp tissue (5 g) was homogenized in various precooled extraction buffer with a tissue grinder (Crl-6010, Kinematica, Kriens-LU, Switzerland) to prepare crude extracts for assay of the following enzymes: 10 mL of 100 mmol/L sodium phosphate (pH 6.0) containing polyvinyl polypyrrolidone

(PVPP) for SOD. 20 mL of 50 mmol/L sodium phosphate buffer (pH 7.0) containing PVPP for CAT. 25 mL of 1 mol/L Tris-HCl (pH 7.6) for LOX.

One unit of SOD was calculated as the amount of enzyme which leads to 50% inhibition of the reduction rate of nitro tetrazolium blue chloride (NBT) per second and the results were expressed as  $U \cdot g^{-1}$  based on fresh weight. CAT activity was calculated from the change at 240 nm, and the results were expressed as  $U \cdot g^{-1}$  based on fresh weight, where  $U = 0.01 \Delta A_{240}$  nm per min. The activity of LOX was assayed according to Xu et al. [26]. LOX activity was calculated from the change at 234 nm, and the results were expressed as  $U \cdot g^{-1}$  based on fresh weight, where  $U = 0.01 \Delta A_{234}$  nm per min.

### 2.12 Statistical Analysis

Measurements of all the indices were conducted in sextuplicate. The data of the figures were shown as the mean  $\pm$  standard deviation. The experimental data was analyzed using Duncan multiple comparison test (One-way ANOVA) using SPSS version 21.0 (IBM Corp., New York, USA). The significance level among all treatments was set equal to 5% ( $p < 0.05$ ). All graph preparation was performed in OriginPro version 2020 (OriginLab Corp., Northampton, MA, USA).

## 3 Results and Discussion

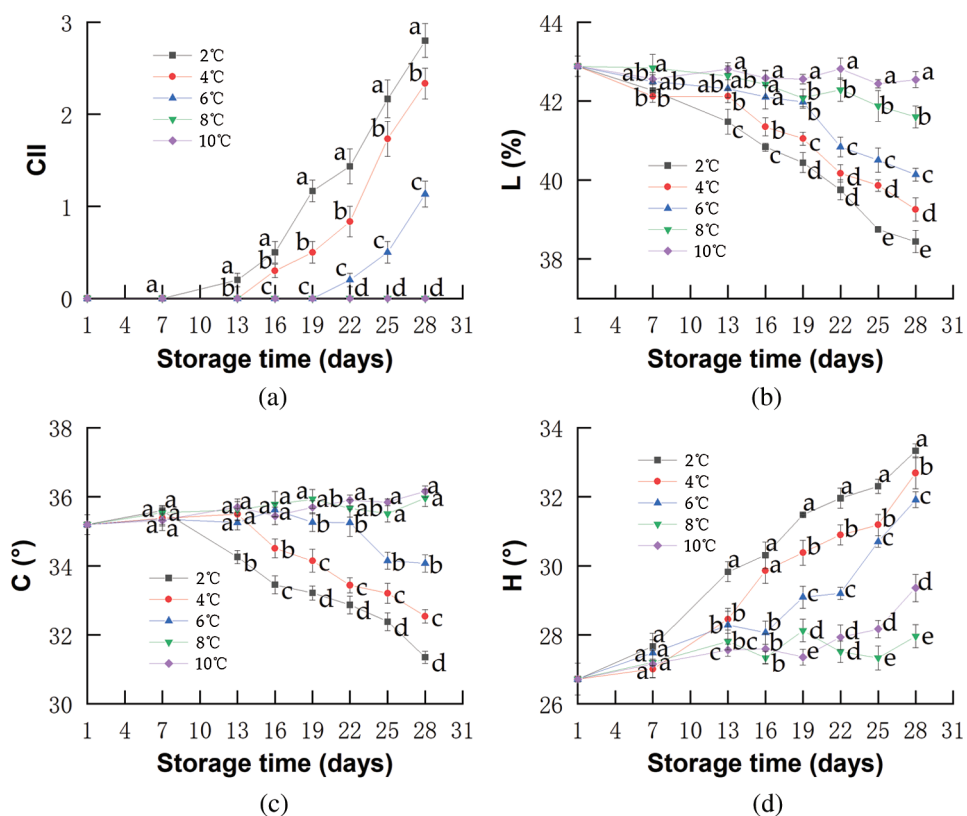
### 3.1 CI Results

CI in red pitahaya has some typical symptoms including uneven brown specks of the skin, browning and shriveling, and abnormal ripening [27,28]. The red pitahayas stored at 2°C showed some pitting at the end of storage and CI did not appear in others. As shown in Fig. 1a, different degrees of CI appearance occurred on the fruits stored at room temperature for 24 h after cold storage at 2, 4, and 6°C, which was similar to those of Liu et al. [29] and Wang et al. [30]. They found that the CI did not appear until the pitahaya fruits were transferred from the low temperature to the room temperature (about 20°C). The CI of red pitahayas stored at 2, 4, and 6°C began to appear on the 12th, 15th, and 21st day, respectively, and the CI index sharply increased from that time. Probably because cold storage conditions triggered metabolic adaptation, for example, degradation of reserves, fermentation, and amino acid mobilization [31]. The temperatures no more than 6°C were too low for defense mechanisms of red pitahayas to cure themselves, which made fruits different. Based on these differences, after fruits were exposed to the room temperature, various degrees of CI appeared. No visual CI symptoms were observed on red pitahaya fruits stored at 8°C and 10°C during storage. These results were in agreement with the findings of Balois-Morales et al. [32], who found the CI of pitahayas stored at 3°C was significantly higher than those of 7°C. Similarly, Nair et al. [33] pointed out that CI appeared more intensely in mango fruits stored at 0°C than those of 5°C and CI was more severe for longer storage time.

### 3.2 Color Evaluation

The color attributes (L, C, and H) values of exocarp adjoining to the equator of red pitahayas were shown in Figs. 1b–1d. The  $L^*$  values of red pitahayas stored at 10°C were maintained stable for 27 days. The reduction of  $L^*$  value was observed in other treatments throughout storage due to the development of brown surface. This result revealed that fruits were significantly darker when stored at lower temperatures, which was in agreement with previous reports [32,34]. Furthermore, the remarkable reduction of  $L^*$  values of red pitahayas stored at 2, 4, and 6°C might result from pigmentation caused by CI. The association between a reduced  $L^*$  value and an increased CI during cold storage has been reported in pineapple [35] and pomegranate [36]. Low-temperature storage prevented the development of chroma. Chroma values of red pitahayas stored at 2, 4, and 6°C decreased notably (Fig. 1c) and chroma was lower in red pitahayas stored at the lower temperatures. This suggests that the red color lost purity at low temperatures (below 6°C) [37]. However, there was no significant difference ( $p > 0.05$ ) of chroma

between red pitahayas stored at 8 and 10°C, and chroma increased slightly. The results were in accordance with Nyanjage et al. [38] who observed a decrease in chroma value of mango fruits stored at 4°C with CI and an increasing tendency for chroma during storage at 13 and 20°C without CI. This was because the fruits with CI did not have the capacity to increase color saturation in post-cold storage, an incapacity interpreted as a manifestation of CI [39,40]. Red pitahayas stored at lower temperatures showed higher H (Fig. 1d), suggesting a tendency to orange colorations, which was due to the irregular ripening with low temperature [39]. However, the hue value of fruits stored at 10°C for 27 days was higher than those stored at 8°C for 27 days due to rot and chlorophyll degradation as the ripening process progressed [41]. Corrales Garcia et al. [42] also found the pitayas rewarming from 4°C decreased the pinkish-red color and the pitayas rewarming from 8°C were not greatly affected. In short, cold storage significantly influenced the color of red pitahaya and 8°C was an effective way to maintain good appearances such as higher L value, higher C value, and lower H value.

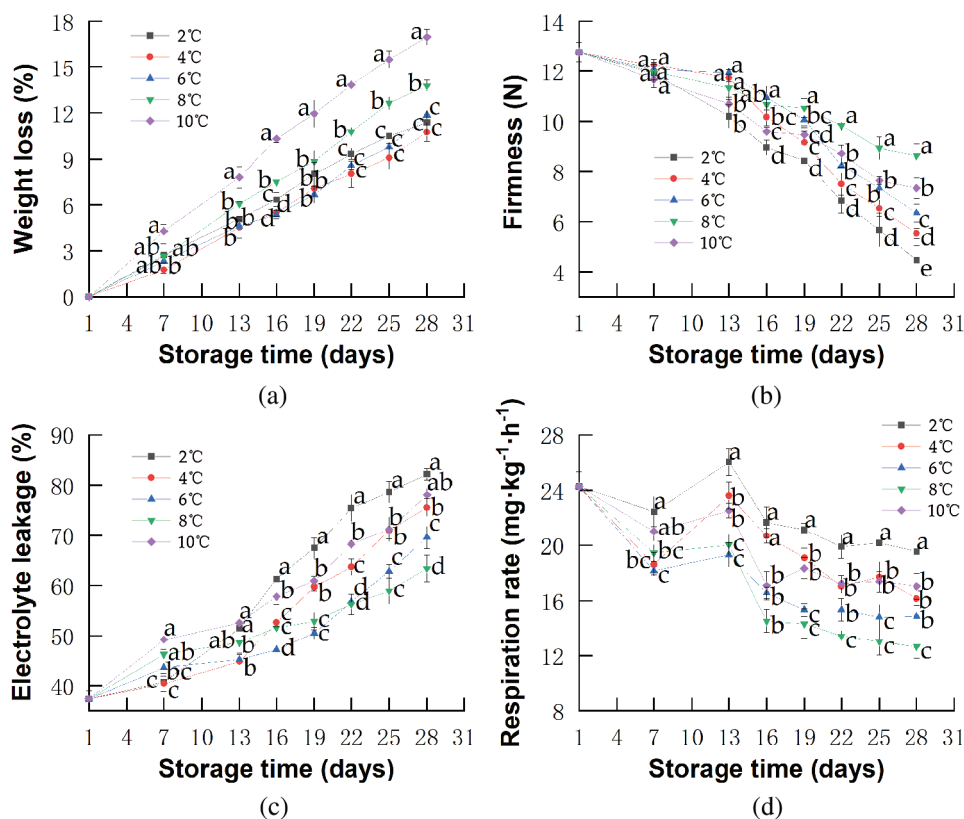


**Figure 1:** Changes in CII (a), Lightness (b), Chroma (c), Hue angle (d) of red pitahayas during cold storage (2°C 1-■; 4°C 1-●; 6°C 1-▲; 8°C 1-▼; 10°C 1-◆) and an extra period of 1 day at 23°C. The data show means  $\pm$  SD of six replicates. Different letters on the same day indicate significant differences ( $p < 0.05$ )

### 3.3 WL Results

The WL is considered a limiting factor for marketing because it causes the skin to wrinkle and fruits to have lower visual appeal [43]. As shown in Fig. 2a, there was a continuous increase in WL in all samples during storage. The WL in red pitahayas stored at 10°C was 17.0% at the end of storage, which was higher than that in other samples. Cold storage could inhibit the evaporation from the tissue surface (a result of transpiration processes) and decrease the WL [44]. Brunini et al. [45] reported the WL in

pitahayas stored at 13°C was 2.1% and higher than stored at 8°C on the 25th day. However, red pitahayas stored at 2°C had higher WL than that at 4 and 6°C and there was no significant difference among these treatments. Probably because decay caused by CI increased the loss of water so that the effect of protecting WL by low temperatures was covered. Proulx et al. [46] found that the WL of papayas stored at 0°C maintained a slightly higher level compared with the those stored at 5°C.



**Figure 2:** Changes in WL (a), Firmness (b), EL (c), Respiration rate (d) of red pitahayas during cold storage (2°C□; 4°C●; 6°C▲; 8°C▼; 10°C◆) and an extra period of 1 day at 23°C. The data show means ± SD of six replicates. Different letters on the same day indicate significant differences ( $p < 0.05$ )

### 3.4 Fruit Firmness

The fruit firmness is closely related to the fruit ripeness, and is also used to determine the suitability of fruits for critical steps in postharvest handling, including storage in bulk bins, passing across a grading line, or whether a particular line of fruits meets the requirement for export. All these processes are problematic when the fruits are too soft [47]. The firmness of all red pitahaya samples decreased slightly from the 1st day to the 13th day (Fig. 2b) presumably because of low pectinase enzyme activities controlled by cold temperature [48], but after storage at low temperatures for over 2 weeks, a quick decrease of firmness turned to be notable owing to continuous cell wall degradation induced by pectinase and oxidative damage caused by senescence and chilling disorder. After 27 days of cold storage, the firmness of red pitahayas stored at 10°C decreased rapidly from 12.8 N on the 1st day to 7.3 N at the end of storage, while the decrease was delayed by storing at 8°C, slowly from 12.8 N to 8.6 N. Kan et al. [49] reported that peaches kept at 5°C showed higher firmness than those stored at 20°C. Red pitahayas showed a higher reduction in firmness at high temperatures due to a higher activity of cell wall degrading enzymes

that lead to fruit softening [50]. On the other hand, the reduction of red pitahaya firmness after 18 days at 2, 4, or 6°C was higher than that stored at 8 and 10°C. Ming et al. [51] reported that the Hami melons stored at 3°C had higher firmness than those stored at 0.5°C, which was explained by disorders as a result of exposure to near-threshold temperatures over a long period, producing irreversible consequences such as the damage of membranes, rupture of cell compartments, and EL [39].

### 3.5 EL Results

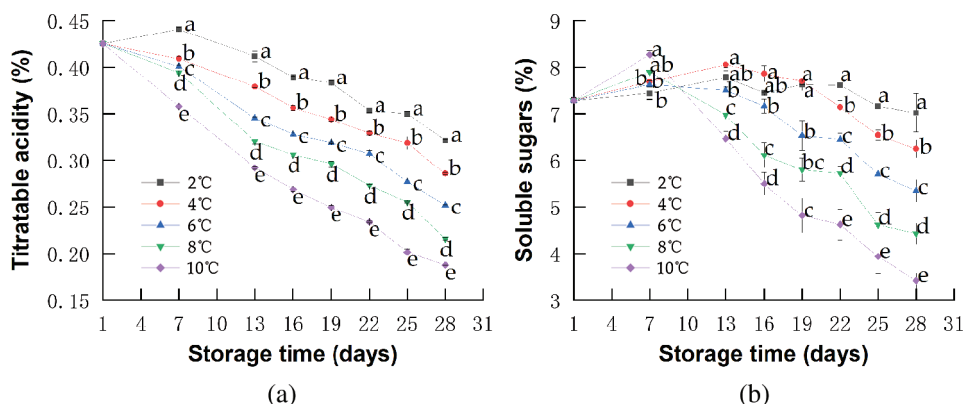
EL usually reflects the permeability and integrity of the cell membrane, which is considered to be the primary site for the development of CI, and thus membrane penetrability might be involved in improving resistance to cold stress [52]. In this study, EL in red pitahayas increased slightly from the 1st day to the 7th day and then increased sharply (Fig. 2c). A major increase in the fruits stored at 2°C was observed on the 13th day of storage. A relatively smooth increase in the fruits stored at 8 and 10°C, suggesting that low-temperature storage (2, 4, and 6°C) could cause cellular membrane damage in red pitahayas. Probably because chilling storage led to the damage of cellular compartmentalization and increase of EL [53]. Furthermore, the rapid increase of EL of red pitahayas stored at 10°C could be attributed to the cell integrity loss caused by the senescence process and mold contamination [54]. EL was significantly ( $p < 0.05$ ) lower at 8°C and storing at 8°C could inhibit the loss of membrane integrity caused by both CI and senescence process.

### 3.6 Respiration Rate

Fruit respiration rate is indicative of the intensity of metabolic activity and biochemical reactions taking place in fruits, which can greatly affect fruit quality [55]. As showed in Fig. 2d, low-temperature storage significantly ( $p < 0.05$ ) impacted the respiration rate. The respiration rates in red pitahayas from all samples first decreased, then reached a peak on the 13th day and finally descended at a slow pace until the end of the storage. Rosas-Benítez et al. [56] showed a similar respiration pattern in pitahayas stored at 12°C. Red pitahayas stored at 8°C showed a lower respiration rate than those stored at 10°C. Raza et al. [57] found that mango stored at 10°C showed a 1-fold lower respiration rate during storage as compared to the fruits stored at 12°C. Presumably, down-regulation of the respiration at low temperatures might save internal O<sub>2</sub> and relieve hypoxic conditions in the fruits [58]. The lowest respiration rates observed were at 8°C, not at 2, 4 or 6°C and the respiration rate values for the fruits ranged in ascending order as follows: 8°C (12.6 mg·kg<sup>-1</sup>·h<sup>-1</sup>) < 6°C (14.9 mg·kg<sup>-1</sup>·h<sup>-1</sup>) < 4°C (16.1 mg·kg<sup>-1</sup>·h<sup>-1</sup>) < 2°C (19.5 mg·kg<sup>-1</sup>·h<sup>-1</sup>). The higher respiration rate at 2°C compared to 8°C was probably due to the change of mitochondria and their electron transport pathway leading to CI development during 28-day storage [11,59]. Alborno et al. [60] reported that the respiration rate values of cherry tomato stored at 5°C were higher than that at 12.5°C.

### 3.7 TA Results

TA is the indicator of acidity content of fleshy fruits [61]. The TA value on the 1st day was over 0.4% and showed a significant decrease ( $p < 0.05$ ) in all samples during storage (Fig. 3a). The highest value of TA was observed at 2°C at the end of storage, while the lowest level was found those in at 10°C. This indicated that lower storage temperatures could delay the loss of acidity during storage. Pal Singh et al. [62] found that the extent of decrease in TA concentration of Japanese plums was higher at 5°C than that at 0°C, which probably due to using organic acids as substrates during the metabolic reactions. Additionally, TA slightly increased in red pitahayas stored on the 7th day at 2°C compared to those on the 1st day resulting from increasing the ability to accumulate malate and decreasing leakage of solutes, such as protons or protonated forms of organic acids by low temperature [63]. Ding et al. [64] reported that mango fruits stored at 5°C for 10 days had higher TA values than those at harvest.



**Figure 3:** Changes in TA (a) and SS (b) of red pitahayas during cold storage (2°C1-■; 4°C1-●; 6°C1-▲; 8°C1-▼; 10°C1-◆) and an extra period of 1 day at 23°C. The data show means  $\pm$  SD of six replicates. Different letters on the same day indicate significant differences ( $p < 0.05$ )

### 3.8 SS Results

SS changes are easily found in fruits under cold stress conditions, which plays a crucial role in osmoregulation and cryoprotection in fruits [65]. There was an increase in the SS at the beginning and then had a decrease (Fig. 3b). The increased total SS of red pitahayas resulted from starch breakdown or hydrolysis of mucilage of the fruits as a response to low temperatures [38,66]. The decrease in total SS could be explained by metabolic processes and lacking the accumulation of carbohydrates in the fruits [10,67]. The red pitahayas stored at 6, 8, and 10°C diminished SS rapidly from the 7th day and those stored at 2 and 4°C remained significantly ( $p < 0.05$ ) high during storage. Probably because higher levels of reducing-sugar played a positive role in improving chilling tolerance, which contributed to signal pathway regulation [68]. Hong et al. [69] found that higher total SS was also observed in summer pineapple fruits held at 6°C than those at 10 and 25°C due to the rapid sugar conversion rate during storage [57]. The accumulated SS acted as an important anti-freeze solute in response to low-temperature stress [52,70].

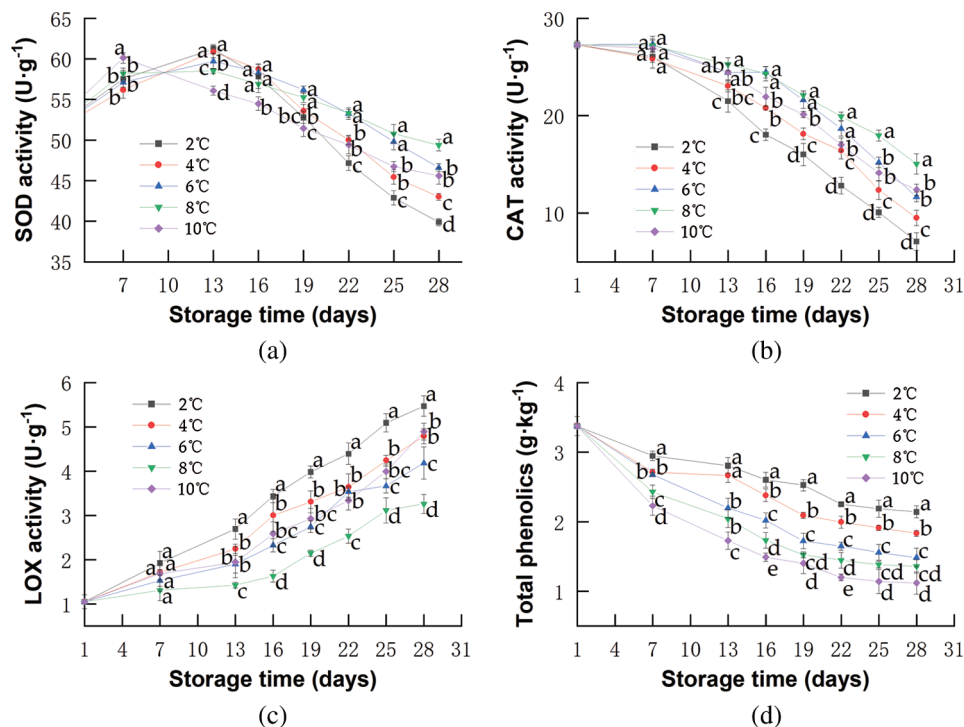
### 3.9 Enzyme Assays

Oxidative damage is regarded as an early response to low temperatures. To get rid of oxidative stress, fruits have applied ROS scavenging strategy. SOD and CAT play essential roles in this defense [4,71]. Besides, LOX regulates fruit development at different stages, which is tightly associated with the generation and identification of stress signals [51].

SOD is a major actor in eliminating highly reactive and toxic superoxide radicals generated during metabolic processes in fruits [72]. As shown in Fig. 4a, SOD activities of red pitahayas stored at 8 and 10°C increased sharply at the beginning in response to the presence of growing toxic ROS, such as superoxide radicals, hydrogen peroxide, hydroxyl radicals, and simple oxygen [73]. On the 13th day, the level of SOD activities in red pitahayas stored at 10°C was significantly lower than that at 8°C ( $p < 0.05$ ), which resulted from further senescence and water loss during storage resulting in more production of ROS and afterward the destruction of the dynamic balance of protective enzyme including SOD. Yuanzhi et al. [74] reported that higher levels of superoxide dismutase activity were observed in the rambutan fruits stored at 10°C than that at 25°C. Moreover, the faster-decreased SOD activities were noticed at lower temperatures (Fig. 4a) caused by severe chilling stress and resulted in more accumulation of superoxide anions, then leading to greater CI incidence [15,75]. Besides, we observed that low-temperature storage could delay the increase rate of SOD activities probably owing to the reduction of the



reaction rate of enzyme production and catalysis by low temperature. Qian et al. [76] also demonstrated that low temperature effectively delayed the increase of SOD activities in stored blueberries.



**Figure 4:** Changes in SOD activity (a), CAT activity (b), LOX activity (c), TP (d) of red pitahayas during cold storage (2°C□; 4°C●; 6°C▲; 8°C▼; 10°C◆) and an extra period of 1 day at 23°C. The data show means  $\pm$  SD of six replicates. Different letters on the same day indicate significant differences ( $p < 0.05$ )

CAT protects cells against ROS by catalyzing the decomposition of hydrogen peroxide to oxygen and water, which prevents  $H_2O_2$  from accumulating in tissues [77]. As shown in Fig. 4b, CAT activities of red pitahayas decreased in all samples during storage. At the end of storage, the CAT activities of red pitahayas stored at 2, 4, 6, and 8°C were 7.09, 9.51, 11.66, and 15.05  $U \cdot g^{-1}$ , respectively. This showed that higher temperatures inhibited the decrease of CAT activities. Huang et al. [78] reported that the CAT activity in the navel orange stored at 20°C was slightly higher than that at 6°C. The high CAT activity has been proposed to impart cold tolerance [62], which could partly explain why there was no CI discovered in red pitahayas stored at 8°C. In contrast, red pitahayas stored at 10°C showed a sharper decrease in CAT activity than those at 8°C, indicating being less capable of alleviating the formation of hydroxyl radical perhaps owing to senescence [76].

LOX can introduce oxygen into unsaturated fatty acids to form unstable hydroperoxides that continually form free radicals, leading to cell damage and apoptosis [79]. LOX activity increased in all samples during storage (Fig. 4c) owing to the deoxygenation of polyunsaturated fatty acids, which resulted in toxic hydroperoxy fatty acids and membrane damage. A similar effect was also reported by Handong et al. [43]. LOX activity of red pitahayas stored at lower temperatures had higher values indicating severer lipid membrane peroxidation under cold stress. Ming et al. [51] also found Hami melons stored at 0.5°C had higher LOX activity values than those at 3°C, which reflected a more severe oxidation level of lipid

membranes due to the development of CI. LOX activity of red pitahayas stored at 10°C was also higher than that at 8°C due to rotting, which was coincident with the result of EL. Xiujuan et al. [79] reported that the LOX activity of peaches stored at 25°C was much higher and underwent greater changes than that at 4°C.

### 3.10 TP Results

As ROS usually accumulates in reaction to chilling stress, phenols are antioxidant compounds that protect fruit cells from oxidative damage attributed to ROS [80]. The TP decreased in all samples due to the oxidation of monohydric or dihydric phenols and the degradation of phenolic as a result of enzymatic activities that occurred in the fruits [81]. As shown in Fig. 4d, the TP content of red pitahayas stored at 2, 4, 6, 8, and 10°C decreased from 3.37 to 2.14, 1.83, 1.48, 1.36, and 1.12 g·kg<sup>-1</sup>, respectively, on the 28th day. The TP contents of kiwifruit and kinnow also showed temperature dependence and the degradation of phenolic occurred faster with higher temperatures [82,83]. Therefore, low-temperature storage is effective to limit the degradation of phenolic compounds.

Pearson correlation was of great significance ( $p < 0.01$ ) between total phenols and antioxidant enzymes SOD, CAT, and LOX. Total phenols content was directly correlated with SOD activities ( $R = 0.38$ ) and CAT activities ( $R = 0.43$ ) while inversely correlated with LOX activity ( $R = -0.36$ ). It demonstrated that the content of total phenols was probably related to the activities of antioxidant enzymes such as SOD, CAT, and LOX during low-temperature storage of red pitahayas. And weak correlation was maybe because the mechanism of response to CI was slightly different between phenolic compounds and antioxidant enzymes (Fig. 4).

## 4 Conclusions

Low-temperature storage had an impact on red pitahaya metabolism during storage. Cold storage at temperatures below 10°C could maintain fruits a good quality by reducing the key physiological and biochemical parameters studied. However, red pitahayas are susceptible to CI. Worse appearance evaluation, lower firmness, higher EL and respiration rate, and more toxic ROS indicated by TP and enzyme activities were observed during low-temperature storage (2, 4, and 6°C). Low-temperature (8 and 10°C) storage contributed to remarkably better fruit quality of red pitahayas. Compared with storage at 10°C, red pitahaya fruits stored at 8°C maintained higher TA, SS, SOD activity, and CAT activity and inhibited LOX activity, cell membranes damage, and respiratory metabolism, thereby effectively extending the shelf time. In conclusion, our current study demonstrated that the better storage temperature for red pitahaya fruits was 8°C for 27 days without CI.

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**Conflicts of Interest:** The authors declare that they have no conflicts of interest to report regarding the present study.

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