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## Drought Stress Alleviation in *Chenopodium quinoa* through Synergistic Effect of Silicon and Molybdenum via Triggering of *SNF1*-Associated *Protein Kinase 2* Signaling Mechanism

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### ABSTRACT

Drought stress negatively impacts agricultural crop yields. By using mineral fertilizers and chemical regulators to encourage plant development and growth, its impact can be mitigated. The current study revealed that exogenous silicon (Si) (potassium silicate;  $K_2Si_2O_5$  at 1000 ppm) and molybdenum (Mo) (ammonium molybdate;  $(NH_4)_6Mo_7O_{24} \cdot 4H_2O$  at 100 ppm) improved drought tolerance in quinoa (*Chenopodium quinoa* Willd). The research was conducted in a randomized complete block design with three biological replicates. The treatments comprised T0 (control, water spray), T4 (drought stress), and T1, T2, T3, T5, T6, and T7, i.e., foliar applications of silicon and molybdenum solutions individually and in combination. Results revealed that drought stress predominantly affected the quinoa yield by decreasing the growth, physiological, biochemical, metabolic, hormonal, antioxidant, and ionic attributes. On the contrary, the supplementation of Si and Mo enhanced the growth attributes (shoot, panicle, and root length, No. of leaves per plant, shoot and panicle fresh/dry weight, root fresh/dry weight, No. of seeds and seeds fresh weight per plant), physiological traits (relative water content, chlorophyll, and carotenoids content), biochemical characteristics (total soluble sugars, protein and lipid content), metabolic attributes (total phenolic, flavonoids, tannins, lycopene, carotene), hormonal contents (indoleacetic acid (IAA), gibberellic acid (GA), salicylic acid (SA)), enzymatic and non-enzymatic antioxidants (catalase, peroxidase and ascorbic acid), and ionic content (potassium (K), (calcium) Ca, (magnesium) Mg, Si and Mo). Under drought stress, Si and Mo reduced electrolyte leakage, abscisic acid (ABA) content,  $H_2O_2$  production, and sodium uptake. In addition, combined Si and Mo supplementation elevated the expression of the *sucrose non-fermenting 1 (SNF1)-associated protein kinase 2 (SnRK2)* (*CqSNRK2.10*) gene in quinoa under drought stress vs. control, signifying an essential regulatory function for Si and Mo-induced drought stress tolerance. These results imply that the exogenous administration of Si and Mo in combination might be an efficient method to alleviate drought stress on quinoa.



**KEYWORDS**Potassium silicate; ammonium molybdate; *Chenopodium quinoa*; drought; stress mitigation**1 Introduction**

Being sessile organisms, plants are susceptible to several environmental stressors, including low temperatures, salinity, as well as drought [1]. Stress-tolerant plant varieties adapt physiologically, morphologically, biochemically, and molecularly to control growth and performance during drought, affecting 21% of the world's land area [2,3]. Drought impacts agricultural output and plant growth, necessitating stress-reduction strategies like hyperactivating Reactive Oxygen Species (ROS) scavenging machinery, increasing antioxidant enzyme activities, and activating stress-tolerant genotypes to prevent cell damage [4].

The xerophytic plants can withstand drought by using strategies like (i) osmolyte biosynthesis and accumulation to regulate turgor pressure and prevent structural membrane damage, (ii) regularizing ionic equilibrium and balancing, (iii) modulating water use efficacy and uptake, (iv) elevated photosynthetic activity, (v) biosynthesis and signaling of phytohormones, (vi) metabolic production related to the growth-promotion and stress-mitigation, (vii) genetic adaptation via modulating the expression of genes that drive stress perception and cell signaling, as well as initiate the expression of genes for metabolite and hormone accumulation, for mitigation of drought stress. Drought stress negatively impacts quinoa growth, development, and yield due to insufficient adaptations in stress-prone plants, indicating that not all plant species can fully handle drought [5].

Quinoa, a genetically diverse crop with strong nutritional capacity, is recognized by the FAO for its potential to ensure human food security in the 21st century. Quinoa is a staple in the human diet due to its high nutritional value, often used as a substitute for rice in various foods [6,7]. As reported earlier [8], drought stress (8% VWC) elevated H<sub>2</sub>O<sub>2</sub> and MDA levels in quinoa while significantly reducing plant growth and relative water content, it is still superior to other environmental factors. Researchers are focusing on exploring the key regulatory genes controlling morpho-physiological changes in quinoa to improve drought resistance. High-quality genome sequencing may help explore molecular underpinnings and enhance plant growth features, despite the lack of research on drought-responsive processes in other crops. For example, *LOC110738152* and *LOC110713661* have been revealed as master genes controlling the drought tolerance responses in quinoa [9]. Another study found that *CqZF-HD14*, a crucial gene regulating drought tolerance, interacted with *CqNAC79* or *CqHIPP34* to ameliorate the drought stress resistance of quinoa seedlings [10]. Furthermore, the sucrose *non-fermenting 1 (SNF1)-associated protein kinase 2 (SnRK2)* gene plays essential roles in biotic and abiotic stresses, which is a member of a small family of plant-specific *serine/threonine (Ser/Thr) protein kinases*. *CqSnRK2.12* overexpression in *A. thaliana* exhibited drought resistance in comparison to the controls [11].

Scientists are striving to develop various biological and non-biological strategies to enhance drought-stress resistance in quinoa. Recently, the induction of drought tolerance response was reported by rhizobacterial inoculation [12], and biochar application in quinoa [13]. Concerning the micronutrients, sodium silicate application for seed priming was reported [14] in improving wheat growth under drought stress. However, there is a lack of comprehensive studies on the impact of micronutrients on drought stress responses in quinoa. Silicon in soil enhances crop quality, production, growth, photosynthesis, nitrogen fixation, and resistance to drought stress in Si-accumulating plants like rice, wheat, maize, sorghum, oilseed rape, lentil, mango, and tomato [15,16]. However, the bioavailable form of Si (silicic

acid) is frequently constrained, as plants absorb silicic acid as a sole silicon source. The exogenous foliar Si supplementation has been shown to reduce drought effects on wheat, pearl millet, and chestnut [15].

Molybdenum is another favorable element that has a key role in increasing tolerance to environmental problems such as drought, cold, and salt stress, according to previous reports [17,18]. Foliar molybdenum application elevated the yield and agronomic characteristics of wheat [19], supported the defense system in *Ricinus communis* by upregulating the expression of enzymes involved in metabolic processes [20,21], and stimulated the growth, yield, and defense system mechanisms of the mung bean [22].

Therefore, the current study was rationalized to explore the potential of silicon and molybdenum as foliar fertilizers to alleviate drought stress in quinoa, its capacity to withstand stress, and its effects on metabolic, antioxidant, phytohormonal, and mineral status rebalancing.

## 2 Methodology

### 2.1 Seed Collection and Plant Growth Experiment

Mature seeds of quinoa (*Chenopodium quinoa* Willd.) variety Q9, were collected from Agriculture University Faisalabad and stored at 5°C. The experiment in the field was done at the Botanical Garden of Abdul Wali Khan University Mardan. Laboratory work was done at the Molecular Biology and Plant Physiology lab, Abdul Wali Khan University Mardan. Soil for plant growth was collected from the Mardan district of Khyber Pakhtunkhwa (KPK) Pakistan, for physicochemical analysis. The soil composition was comprised of sand content (72% to 75%), silt content (10% to 12%), clay (12%–15%), soil pH (7.2 to 7.9), electrical conductivity (0.5% to 5%), organic carbon (4.17%), organic matter (1.3%), carbonates (1.29 meq/L), bicarbonates (2.9 meq/L), and  $\text{Cl}^-$  (1.16 meq/L).

In present work, the silicon (potassium silicate;  $\text{K}_2\text{Si}_2\text{O}_5$ , CAS No. 1312-76-1, Brenntag GmbH, Messeallee 11, 45131, Essen, Germany) at the concentration of 1000 ppm, and molybdenum (ammonium molybdate;  $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}$ , ammonium molybdate (CAS No. 12054-85-2, Sigma Aldrich, Milwaukee, WI, USA) at the concentration of 100 ppm, was supplemented as foliar fertilization.

The healthy seeds of quinoa were surface sterilized by soaking them in 70% ethanol for 1 minute, followed by three washes with distilled water. After 15 days of germination, the evenly developing seedlings were picked and trimmed out to five per container. Every day, the plants were watered with tap water. Sandy loam soil was placed into each earthen pot, each measuring 30 cm × 12 cm × 12 cm (height, length, breadth). The experimental design followed a completely randomized design (CRD), with 48 pots divided into 8 groups representing different treatments, with at least ten technical replicates and three biological replicates each.

The seedlings were subjected to drought stress after germination for three weeks. After the onset of the first signs of wilting, watering and foliar applications of silicon and molybdenum were performed for each plant in the 4th, 6th, and 10th weeks after seed germination. To ensure leaf absorption, the foliar spray was applied in the morning to both sides of the leaves.

Samples were taken after the development phase for biochemical examination. Merck (Darmstadt, Germany), Sigma-Aldrich (Taufkirchen, Germany), Brenntag GmbH, and Fluka (Buchs, Switzerland) provided the chemicals and reagents for the experiment.

#### **The experimental setup included the following treatments:**

Treatment 0: Control (Distilled water).

Treatment 1: Silicon (Potassium silicate 1000 ppm) foliar spray.

Treatment 2: Molybdenum (Ammonium molybdate 100 ppm) foliar spray.

Treatment 3: Silicon + Molybdenum foliar spray.

Treatment 4: Drought stress.

Treatment 5: Drought + Silicon (Potassium silicate 1000 ppm) foliar spray.

Treatment 6: Drought + Molybdenum (Ammonium molybdate 100 ppm) foliar spray.

Treatment 7: Drought + Silicon + Molybdenum foliar spray.

## **2.2 Monitoring Growth Parameters**

For monitoring growth parameters, various agronomic characteristics were studied. The growth parameters included vegetative measurements such as plant root and shoot length, total number of intact leaves, and dry and fresh weight of roots, stems, and leaves. Reproductive attributes like panicle length, fresh and dry mass of the panicle, number of seeds per plant, weight of seeds per plant, and 1000 seeds weight were also measured.

## **2.3 Physiological Parameters**

Electrolyte leakage (EL) was measured according to the method described earlier [23] with some modifications. The relative water content (RWC) of fresh leaf samples was quantified following the method described earlier [24]. Leaf water loss (LWL) was measured according to the method mentioned previously [25].

## **2.4 Biochemical Analysis**

Biochemical analysis was conducted to measure various parameters using standard protocols.

### **2.4.1 Extraction and Estimation of Photosynthetic Pigments**

The estimation of chlorophyll a, b, total chlorophyll, and carotenoid content in fresh leaf samples (1.0 g) from the 15th leaf (counted from the base) of each plant was performed following the process described earlier [26].

### **2.4.2 Analysis of Metabolites and Phytohormones**

Metabolic evaluation was carried out using fresh leaf samples from the 15th leaf (counted from the base) of each plant. Total soluble sugars were quantified by the method indicated earlier [27]. The leaves' total protein content was quantified according to the method mentioned earlier [28]. Lipid content was quantified as described earlier [29]. The proline quantification was conducted as described earlier [30]. Total phenolics were assessed as explained previously [31]. Lycopene and  $\beta$ -carotene were assessed using the method described earlier [32]. Tannin was measured using the method described earlier [33]. The total flavonoid content was calculated using the  $AlCl_3$  technique reported earlier [34]. The estimation of auxin content was carried out using the Salkowski reagent [35], as described previously [36]. Salicylic acid (SA) content was evaluated as mentioned earlier [30]. Endogenous ascorbic acid (AsA) was quantified by the method mentioned earlier [37]. The content of GA3 and ABA content was investigated by the method mentioned previously [38].

### **2.4.3 Quantification of Enzymatic and Non-Enzymatic Antioxidants**

The total antioxidant capacity was quantified using the method devised earlier [39]. Catalase activity was quantified as described [40]. Guaiacol peroxidase activity was quantified according to the procedure described [41]. Ascorbate peroxidase activity was quantified following the method described earlier [42]. The determination of  $H_2O_2$  was performed according to the method mentioned earlier [31]. The detection of ROS was done using the procedure outlined earlier [43].

#### 2.4.4 Detection of Reactive Oxygen Species (ROS) Accumulation through DAB Activity Assay

Detection of ROS through 3,3-diaminobenzidine (DAB) staining was performed as described earlier [44].

### 2.5 Elemental Analyses

Samples of dried ground plants were measured for the concentrations of several elements ( $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{K}^{+}$ , and  $\text{Na}^{+}$ ). A small amount of 0.5 g of pulverized, dry plant material was powdered and put into digesting vials. Each sample received 6.5 mL of a 5:1:0.5 ratio acid solution ( $\text{HNO}_3$ ,  $\text{H}_2\text{SO}_4$ ,  $\text{HClO}_4$ ) for the digestion procedure. For dilution, after adding distilled water, the sample was filtered using No. 1 Whatman filter paper, as described earlier [45]. Following the steps outlined previously [46]. The modified molybdenum blue technique was used to measure silicon uptake. The extraction-photometric approach utilizing Tetrazolium Violet (TV) was used to determine the molybdenum content, as mentioned earlier [47].

### 2.6 RT-qPCR Analysis for Expression of Drought Stress Marker Gene *CqSnRK2.10*

Total RNA was isolated from quinoa leaves and root tissues, followed by DNase treatment to get purified stable RNA that was subsequently used for cDNA synthesis, following the procedure outlined previously [44]. The gene expression analysis of drought stress-related marker gene, *CqSnRK2.10*, reported earlier [11], was performed using qPCR primers designed by Primer3 [48], with oligonucleotide sequences of *CqSnRK2.10* gene as *F-AAGCCCTCGTTCGGATACTTAATGC*, and *R-GCCTCTTGTTCCACCACTAATCTTCTC*.

Due to the stable expression under drought stress in quinoa, the internal control *CqTUB-9* was used with primer sequences; *F-GAGATGTTCCGTCGTGTGAGTGAG*, and *R-ATCGGCAGTTGCATCCTGGTATTG*, as reported earlier [11]. Oligonucleotides for primers were synthesized by Bio Basic (Seoul, Republic of Korea). Gene expression analysis and data normalization procedure was followed as previously mentioned earlier [44].

### 2.7 Statistical Analysis

Biological triplicates were used in the studies. Data analysis was carried out using a two-way ANOVA. The SPSS 20 (SPSS Inc., Chicago, IL, USA) was utilized to conduct the Duncan multiple range test (DMRT) for the differential comparison of mean values. Different statistical bars marked with significant letters at  $p < 0.05$  were used to display significant differences. Fold changes were calculated as described earlier [29].

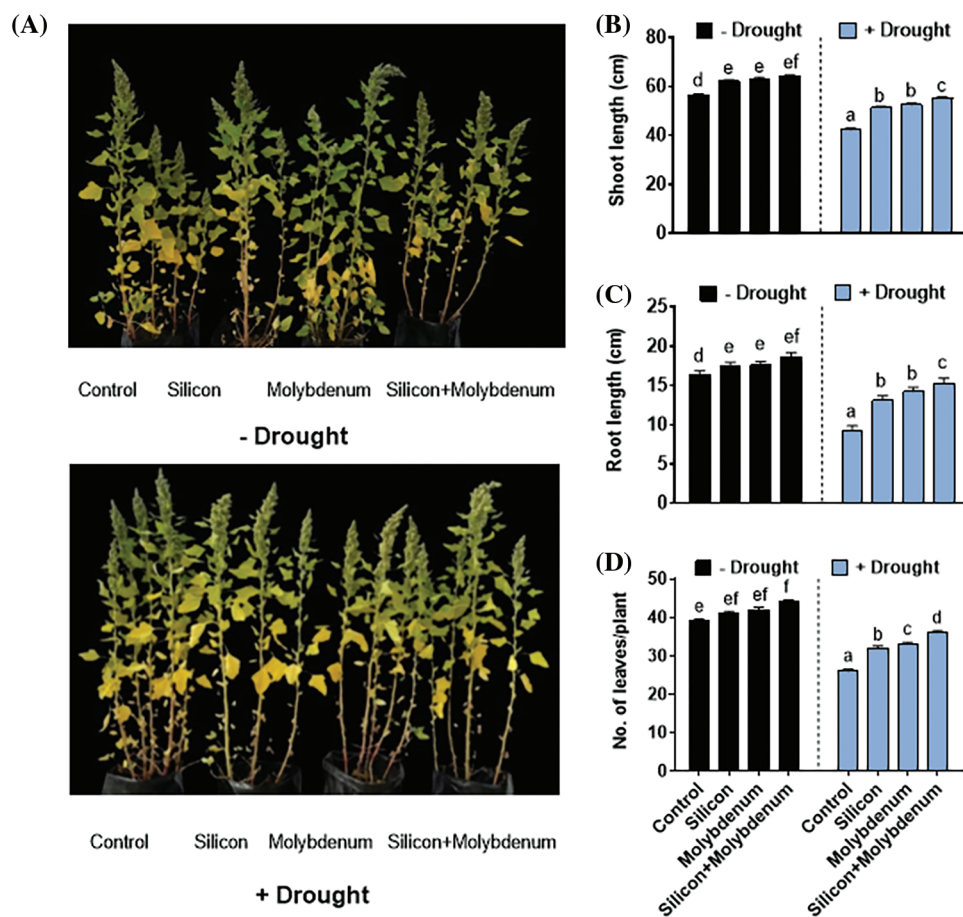
## 3 Results

### 3.1 Effect of Silicon and Molybdenum on Vegetative Growth Attributes of Quinoa under Drought Stress

The findings of this study demonstrate that drought stress exerted a negative impact on the growth of quinoa. The quantitative data for the growth potential of quinoa under drought stress exhibited a statistically significant decrease in shoot length cm [42/57 cm (control)], root length [8.3/17 cm (control)], number of leaves [26/40 (control)], comparable to the respective control plants. Nevertheless, under drought stress the silicon and molybdenum supplementation promoted growth comparable to the respective control plants.

Silicon and molybdenum combined supplementation exhibited statistically significant promotion in shoot length, [54/42 cm (drought control)], root length [(15/8.3 cm (DC), number of leaves [37/26 (drought control)], in quinoa under drought stress, comparable to the respective control plants (Fig. 1A–D).

The quantitative data for the growth performance of quinoa under drought stress showed statistically significant reduction in shoot fresh weight [5.4/10 g (control)], shoot dry weight [1.5/2.9 g (control)], leaves fresh weight per plants [1.6/2.9 g (control)], leaves dry weight per plants [0.29/0.7 g (control)], root fresh weight [1.37/2.35 g (control)], root dry weight [0.37/0.87 g (control)], total fresh weight [8.4/15 g (control)] and total dry weight [2.08/4.5 g (control)], comparable to the respective control plants.



**Figure 1:** Foliar application of silicon and molybdenum on growth phenotype of quinoa under drought stress. Phenotypic analysis (A) shoot length (B) root length (C) and No. of leaves/plant (D). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

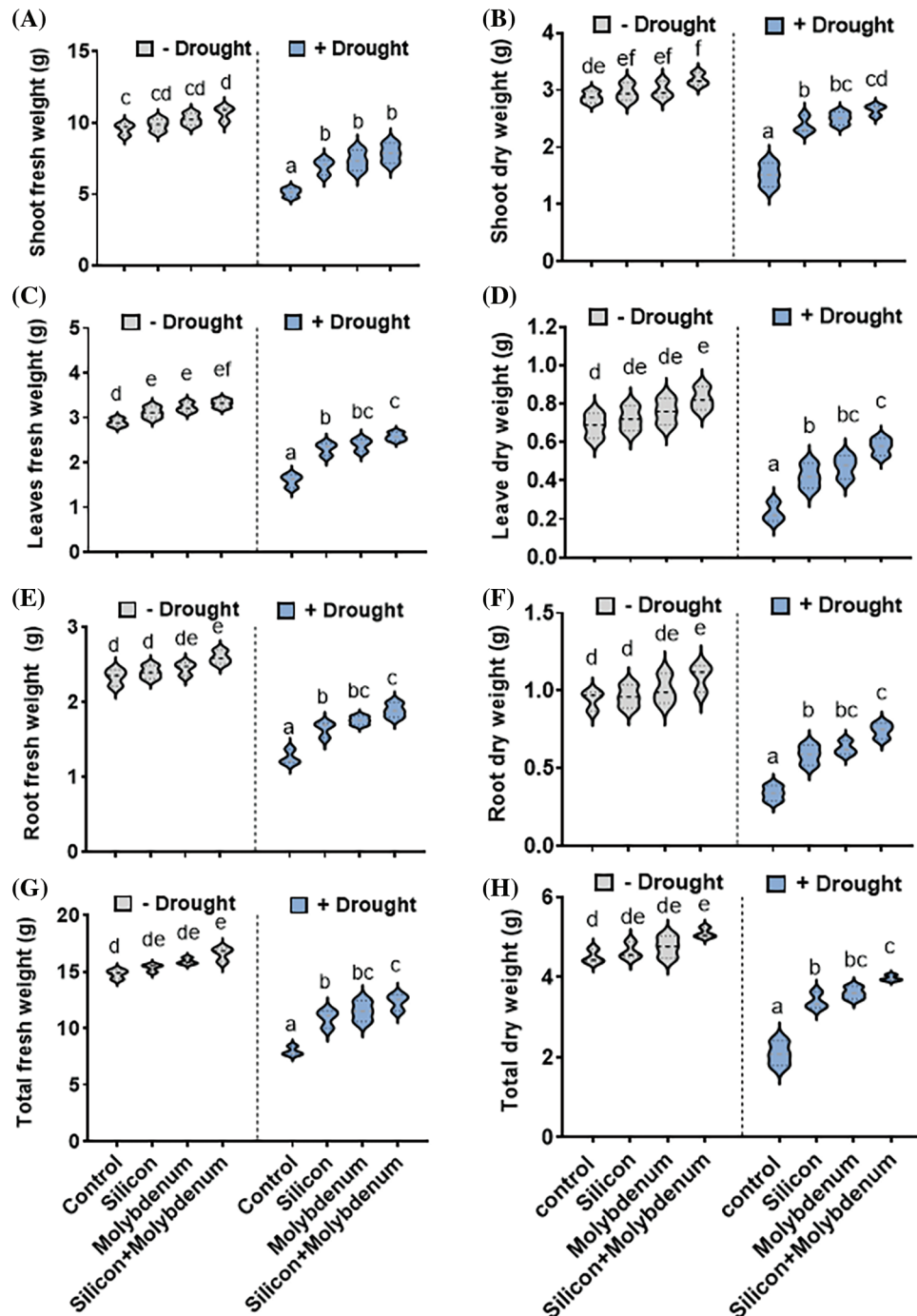
Silicon and molybdenum combined supplementation exhibited statistically significant promotion in shoot fresh weight [8.5/5.4 g (drought control)], shoot dry weight [2.7/1.5 g (drought control)], leaves fresh weight per plant [2.68/1.6 g (drought control)] leaves dry weight per plant [0.58/0.29 g (drought control)], root fresh weight [0.75/0.34 g (DC)], root dry weight [0.79/0.37 g (drought control)], total fresh weight [13/8.4 g (drought control)] and total dry weight [4/2.08 g (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 2A–H).

### 3.2 Effect of Silicon and Molybdenum on Reproductive Growth Attributes of Quinoa under Drought Stress

The findings of the present research exposed that drought stress negatively affects the reproductive growth of quinoa. Parameters such as panicle length, fresh and dry biomass of the panicle, number of seeds per plant, seed weight per plant, and 1000 seeds weight were negatively affected.

Quantitative results for the reproductive growth attributes in quinoa under drought stress showed a statistically significant reduction in panicle length [11.3/23.5 cm (control)], fresh biomass of the panicle [6/17 g (control)], dry biomass of panicle [1.6/3.6 g (control)], number of seeds per plant [620/1001

(control)], total seed weight per plant [1.7/3.8 g (control)], and 1000 seeds weight [2.6/3.8 g (control)], as compared to control (C) without stress.



**Figure 2:** The impact of foliar application of silicon and molybdenum on vegetative growth attributes of quinoa under drought stress. Shoot fresh weight (A) shoot dry weight (B) leaves fresh weight (C) leaves dry weight (D) root fresh weight (E) root dry weight (F) total fresh weight (G) and total dry weight (H). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

Silicon and molybdenum combined supplementation exhibited statistically significant promotion in panicle length [22/11.3 cm (drought control)], fresh biomass of the panicle [12/6 g (drought control)], dry biomass of panicle [3/1.6 g (drought control)], number of seeds per plant [900/620 (drought control)], total seed weight per plant [1.7/3.8 g (drought control)], and 1000 seeds weight [3.7/2.6 g (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 3A–G).

### 3.3 Effect of Silicon and Molybdenum on Physiological Attributes of Quinoa under Drought Stress

In quinoa under drought stress, the quantitative results exposed a statistically significant reduction in relative water content [49/73% (control)], and leaf water loss [32/54% (control)], compared to control (C) without stress, with elevated electrolyte leakage [27/15% (control)].

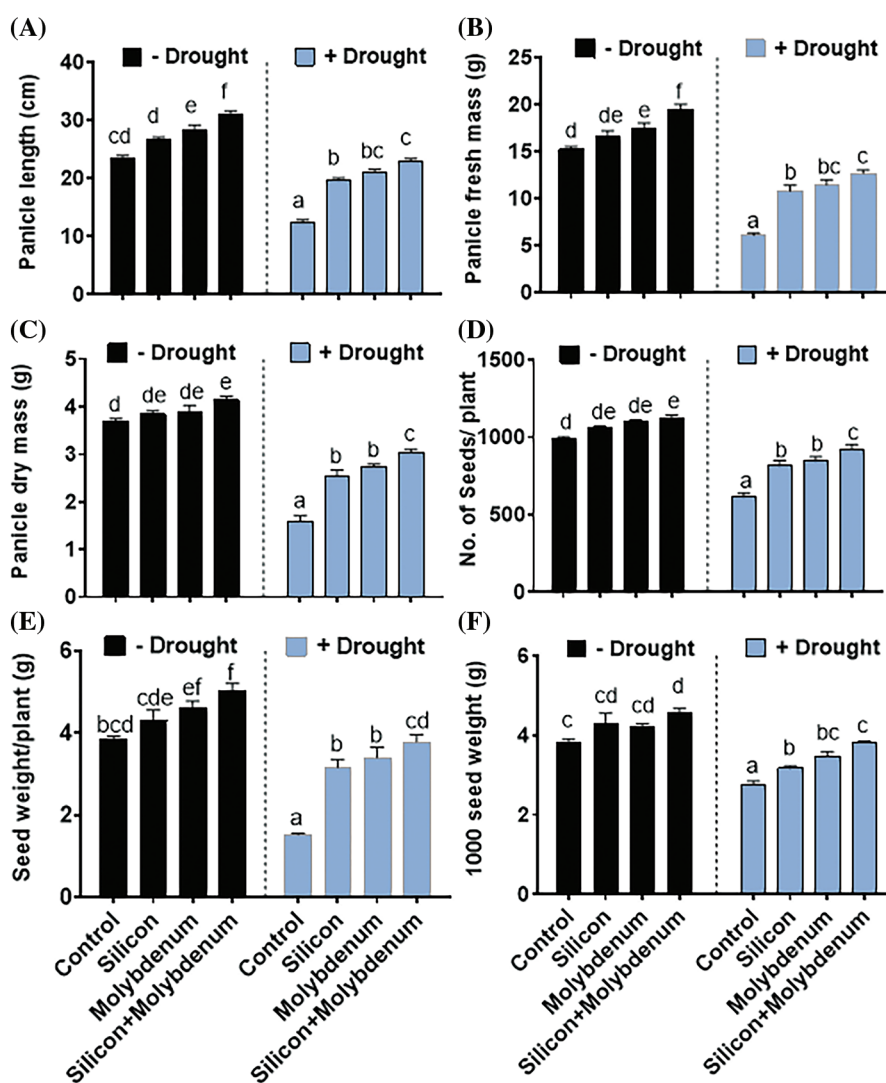
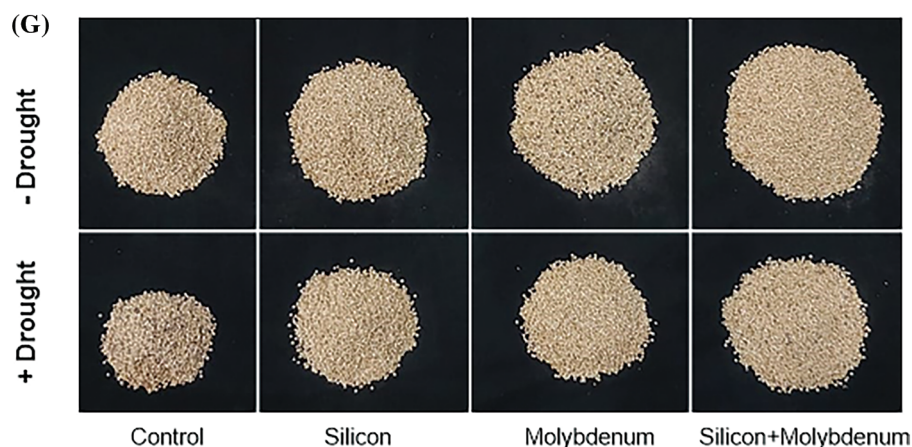


Figure 3: (Continued)





**Figure 3:** Foliar spray effect of silicon and molybdenum on reproductive growth attributes of quinoa under drought stress. Panicle length (A) panicle fresh mass (B) panicle dry weight (C) number of seeds/plants (D) seed weight/plant (E) 1000 seeds weight (F) and visual phenotype of seed mass/plant (G). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates marked with letters denoting significant differences at  $p \leq 0.05$

Nevertheless, silicon and molybdenum combined supplementation exhibited statistically significant promotion in relative water content [75/49% (drought control)], and leaf water loss [53/32% (drought control)] with a decline in electrolyte leakage [20/27% (drought control)] in quinoa plants under drought stress, comparable to the respective control plants (Fig. 4A–C).

### 3.4 Effect of Silicon and Molybdenum on Biochemical Attributes of Quinoa under Drought Stress

#### 3.4.1 Photosynthetic Pigments

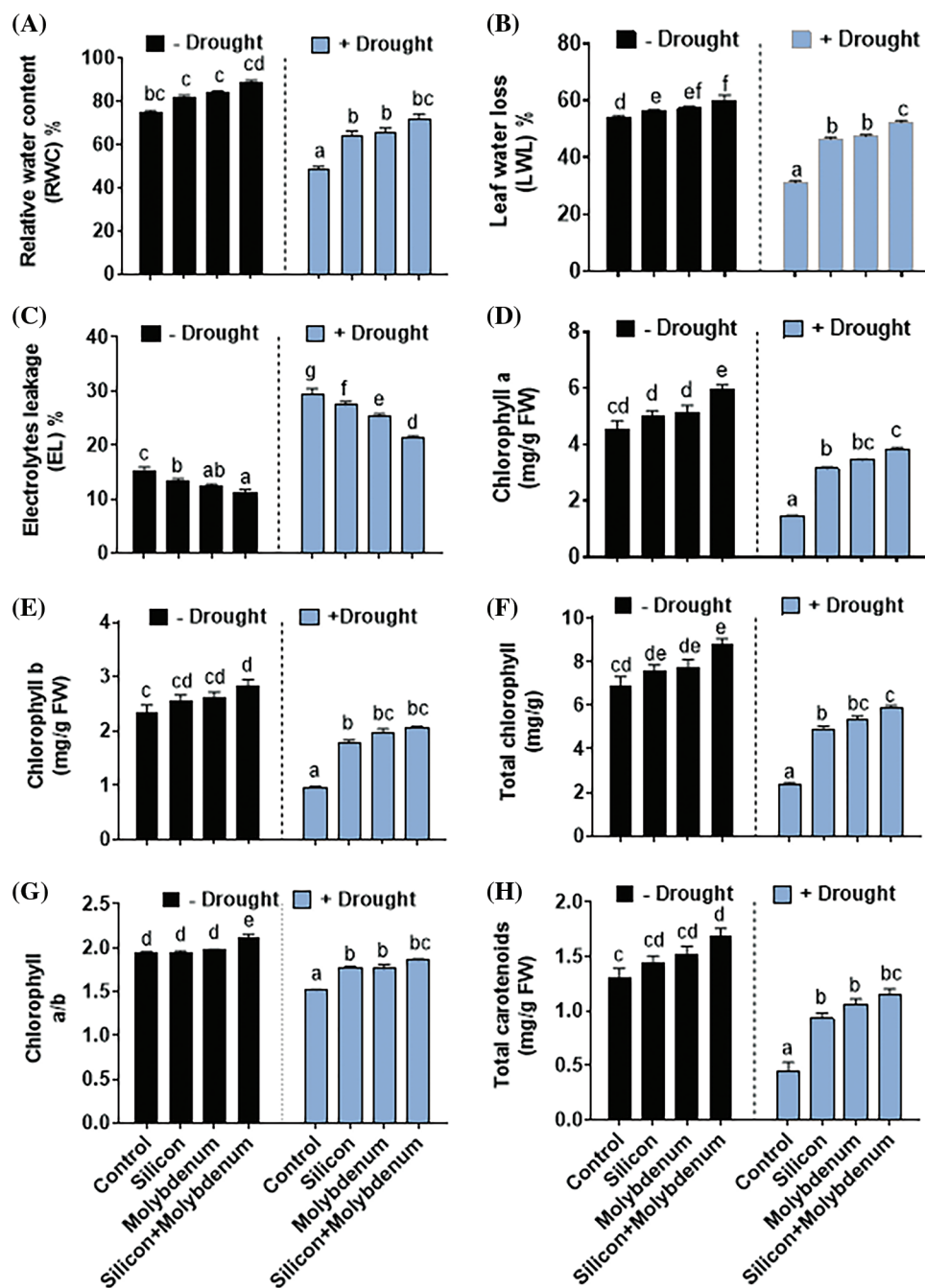
Under drought stress, the quantitative results exposed statistically significant reduction in the chlorophyll a [1.4/4.6 (mg/g FW) (control)], chlorophyll b [0.9/2.3 (mg/g FW) (control)], total chlorophyll [2.3/7.1 (mg/g FW) (control)], chlorophyll a/b ratio [1.4/1.9 (control)], and carotenoid content [0.3/1.3 (mg/g FW) (control)], compared to control (C) without stress.

Silicon and molybdenum combined supplementation exhibited statistically significant promotion in chlorophyll a [3.9/1.4 (mg/g FW) (drought control)], chlorophyll b [2.1/0.9 (mg/g FW) (drought control)], total chlorophyll [6.1/2.3/(drought control)], chlorophyll a/b in quinoa plants under drought stress, comparable to the respective control plants (Fig. 4D–H).

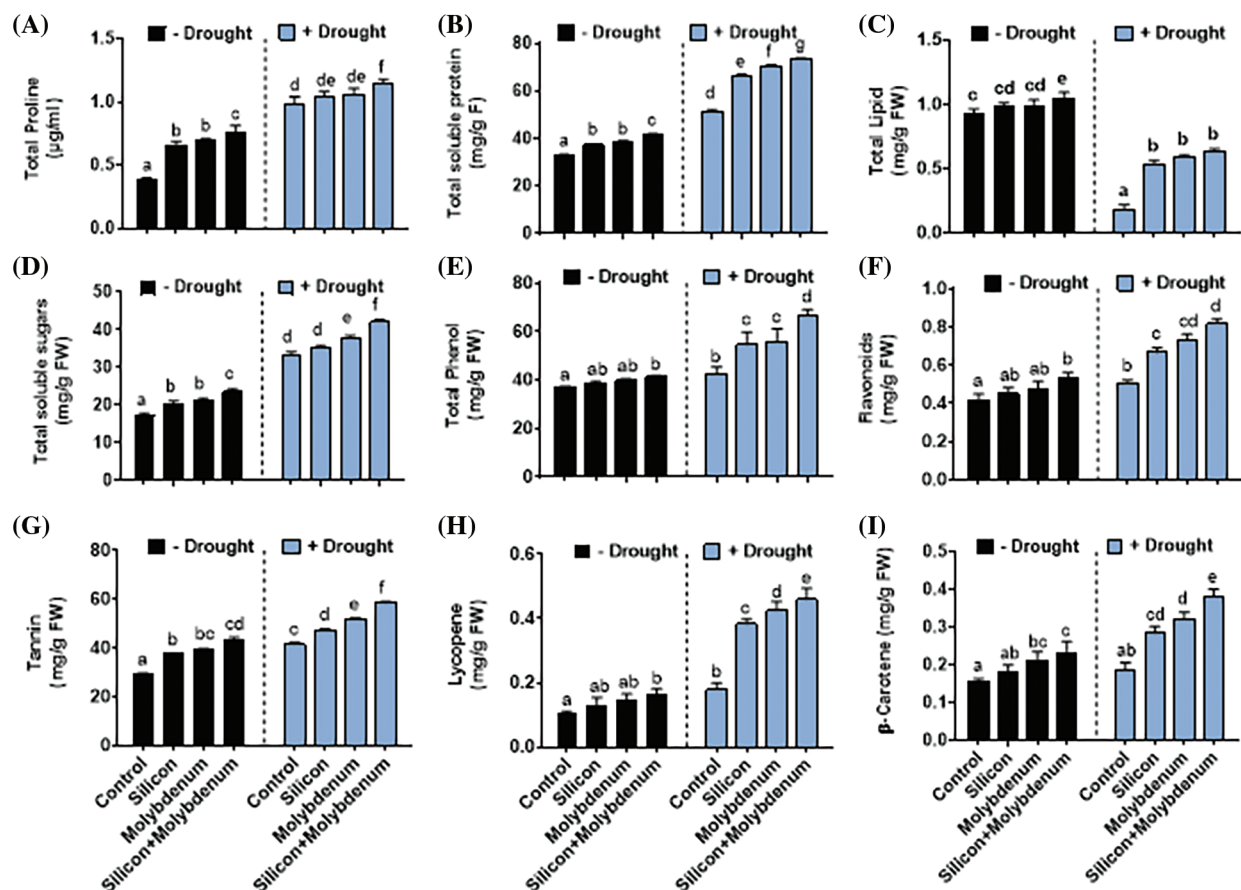
#### 3.4.2 Protein, Lipid and Soluble Sugars Content

Under drought stress, the quantitative results exposed statistically significant promotion in the content of total protein [51/32 (mg/g FW) (control)], proline [0.9/0.3 (mg/mL) (control)], lipids [0.2/0.9 (mg/g FW) (control)], and total soluble sugars [32/72 (mg/g FW) (control)], compared to control (C) without stress.

Silicon, and molybdenum combined supplementation exhibited a statistically significant further promotion in the total protein [73/51 (mg/g FW) (drought control)], proline [1.2/0.3 (mg/mL) (drought control)], lipids [0.5/0.2 (mg/g FW) (drought control)], and total soluble sugars content [43/32 (mg/g FW) (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 5A–E).



**Figure 4:** Foliar spray effect of silicon and molybdenum on physiological attributes and photosynthetic pigments content of quinoa under drought stress. Relative water content (A) leaf water loss (B) electrolytes leakage (C) chlorophyll a (D) chlorophyll b (E) total chlorophyll (F) chlorophyll a/b (G) and total carotenoids (H). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$



**Figure 5:** Foliar spray effect of silicon and molybdenum on biochemical attributes of quinoa under drought stress. Proline (A) total proteins (B) total lipids (C) total soluble sugar (D) total phenols (E) flavonoids content (F) tannin (G) lycopene (H) and  $\beta$ -carotene (I). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

### 3.4.3 Total Phenolics and Flavonoids Content

Under drought stress, the quantitative results exposed statistically significant promotion in the total phenolics [32/17 (mg/g FW) (control)], and total flavonoids [0.5/0.36 mg (mg/mL) (control)], compared to control (C) without stress.

Silicon and molybdenum combined supplementation exhibited a statistically significant further promotion in the total phenolics [43/32 (mg/g FW) (drought control)], and total flavonoids [0.86/0.5 (mg/mL) (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 5F–G).

### 3.4.4 Tannin, Lycopene, and $\beta$ -Carotene Content

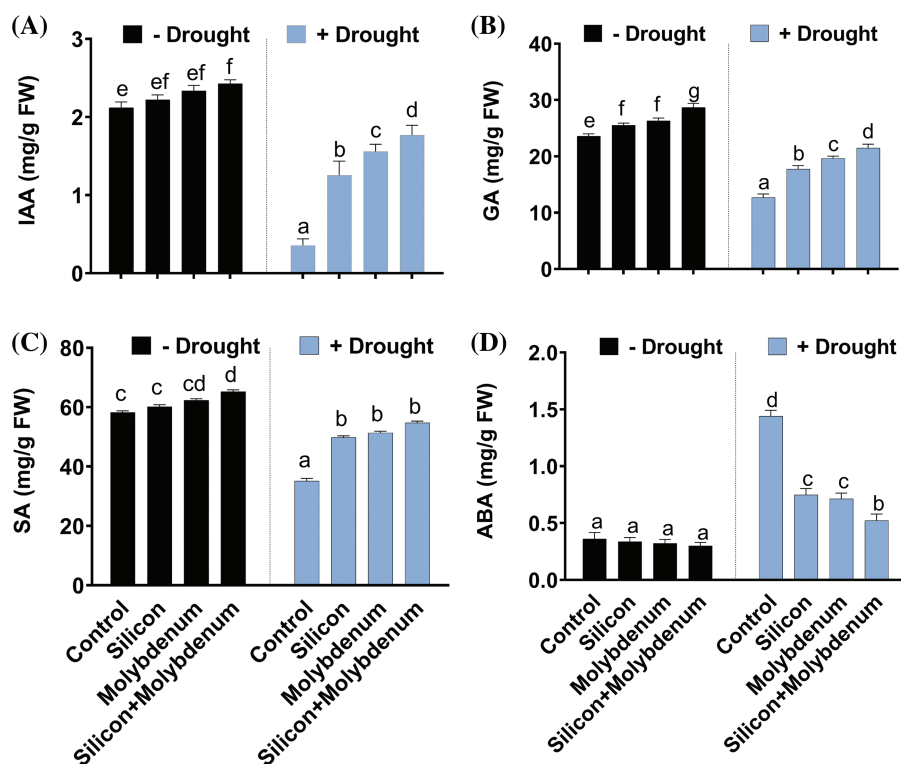
Under drought stress, the quantitative results exposed statistically significant promotion in the tannins [41/30 (mg/mL) (control)], lycopene [0.2/0.1 (mg/mL) (control)], and beta-carotene [0.2/0.1 (mg/g FW) (control)], compared to control (C) without stress.

Silicon, and molybdenum combined supplementation exhibited a statistically significant further promotion in tannins [57/30 (mg/mL) (drought control)], lycopene [0.4/0.2 (mg/mL) (drought control)],

and beta-carotene [0.4/0.1 (mg/g FW) (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 5G–I).

### 3.5 Effect of Silicon and Molybdenum on Hormonal Content of Quinoa under Drought Stress

Under drought stress, the quantitative results exposed a statistically significant decrease in the IAA content [0.5/2.1 (mg/mL) (control)], GA<sub>3</sub> content [12/22 (mg/mL) (control)], and SA content [36/57 (mg/g FW) (control)], compared to control (C) without stress. However, silicon and molybdenum combined supplementation exhibited a statistically significant promotion in IAA content [1.5/0.5 (mg/mL) (drought control)], GA<sub>3</sub> content [25/22 (mg/mL) (drought control)], and SA content [58/57 (mg/g FW) (control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 6A–C).



**Figure 6:** Foliar spray effect of silicon and molybdenum on metabolic content of quinoa under drought stress. IAA content (A) GA<sub>3</sub> content (B) salicylic acid content (C) and ABA content (D). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

The quantitative results exposed statistically significant promotion in the ABA content [1.3/0.4 (mg/mL) (control)], compared to control (C). However, silicon and molybdenum combined supplementation exhibited a statistically significant further promotion in ABA content [0.5/1.3 (mg/mL) (drought control)], in quinoa plants under drought stress, comparable to the respective control plants (Fig. 6D–I).

### 3.6 Effect of Silicon and Molybdenum on Antioxidant Capacity and ROS Status of Quinoa under Drought Stress

Under drought stress, ROS production was studied for assessment of oxidative burst in quinoa upon *silicon and molybdenum* application. To this end, the quantity of  $H_2O_2$  was detected as brown spots in the leaves of the plant using DAB staining in the leaf tissues of quinoa (Fig. 7A).

The highest quantity of  $H_2O_2$  accumulation was recorded in drought-treated plant tissues [48/27 nmol  $g^{-1}$  fresh weight (control)], compared to control (C) without stress (Fig. 7B). However, under drought stress, silicon, and molybdenum combined supplementation exhibited a significantly ( $p \leq 0.05$ ) decreased amount of  $H_2O_2$  tissues [34/48 nmol  $g^{-1}$  fresh weight (drought control)], and intensity of the DAB staining, in quinoa that has been stressed by drought, comparable to the respective control plants (Fig. 7A,B).

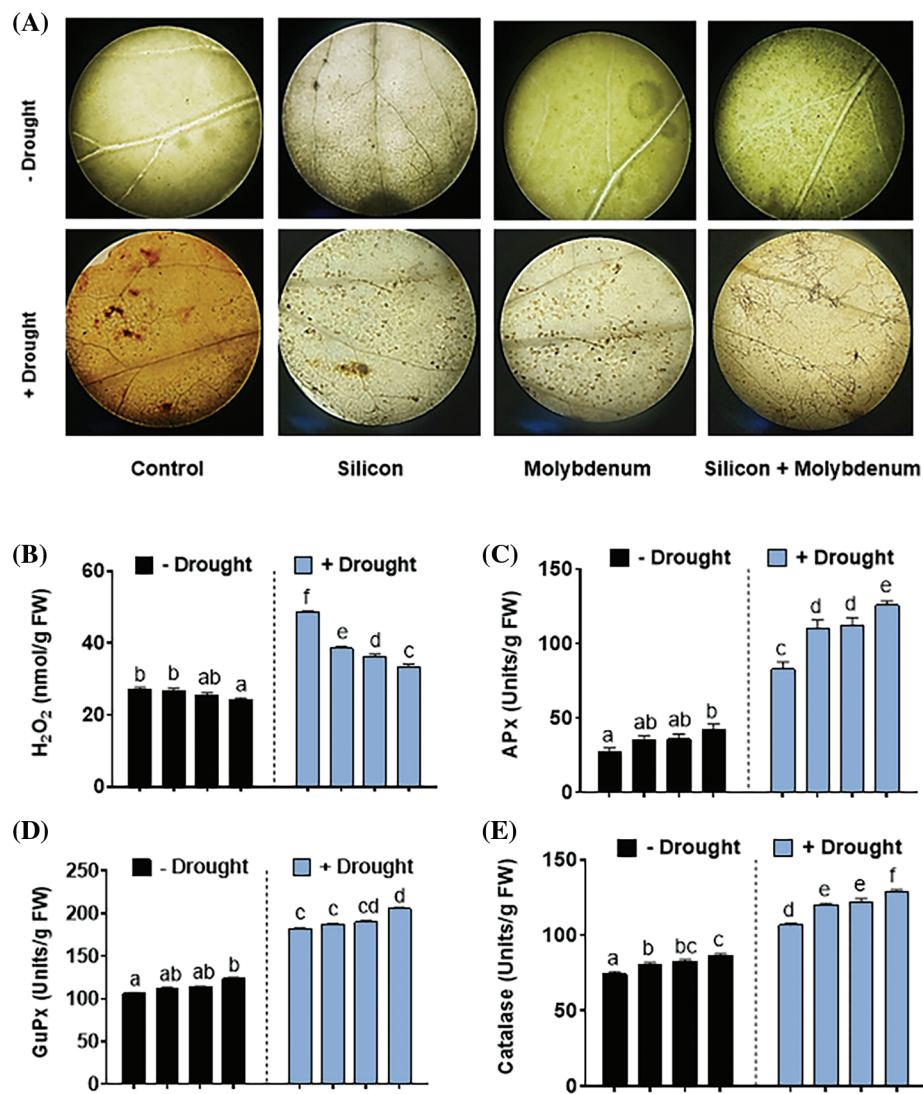
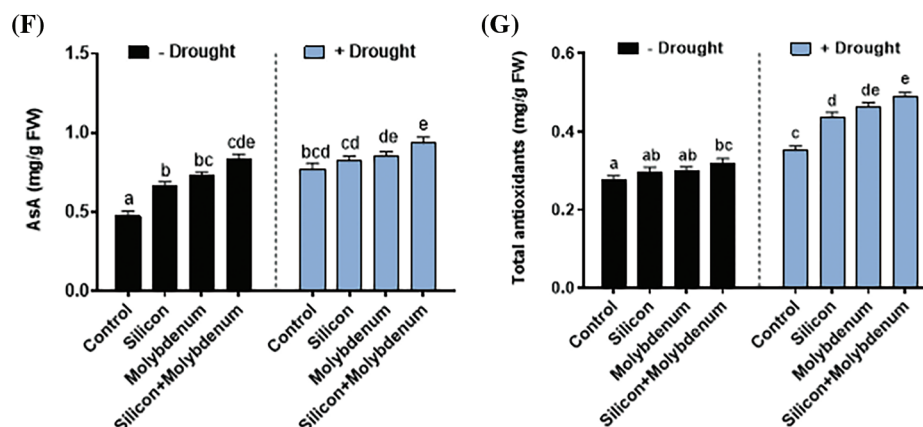


Figure 7: (Continued)



**Figure 7:** Foliar spray effect of silicon and molybdenum on ROS accumulation and antioxidant capacity of quinoa under drought stress. Endogenous ROS accumulation (A)  $H_2O_2$  content (B) ascorbate peroxidase (C) guaiacol peroxidase (D) catalase (E) ascorbic acid content (F) and total antioxidant (G). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

Antioxidants (both non-enzymatic and enzymatic) were investigated, in *quinoa* with separate and collective supplementation of silicon and molybdenum under drought conditions. Fig. 7C–F depicts the differential responses seen under various treatments. The level of enzymatic antioxidants (Ascorbate peroxidase enzyme, Guaiacol peroxidase, Catalase), and non-enzymatic antioxidants (ascorbic acid) was predominantly elevated both under normal and drought stress conditions upon supplementation, both individually and in combination, of silicon and molybdenum compared to control.

Under drought stress, the quantitative results exposed statistically significant promotion in the Ascorbate peroxidase enzyme [75/27 (mg/g FW) (control)], compared to control (C) without stress. Nevertheless, under drought stress, silicon and molybdenum combined supplementation exhibited a significant ( $p \leq 0.05$ ) further promotion in ascorbate peroxidase enzyme [121/75 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 7C). Under drought stress, the quantitative results exposed statistically significant promotion in the Guaiacol peroxidase enzyme [181/108 (mg/g FW) (control)], compared to control (C) without stress. Nevertheless, silicon and molybdenum combined supplementation exhibited a significant ( $p \leq 0.05$ ) further promotion in Guaiacol peroxidase enzyme [205/181 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 7D).

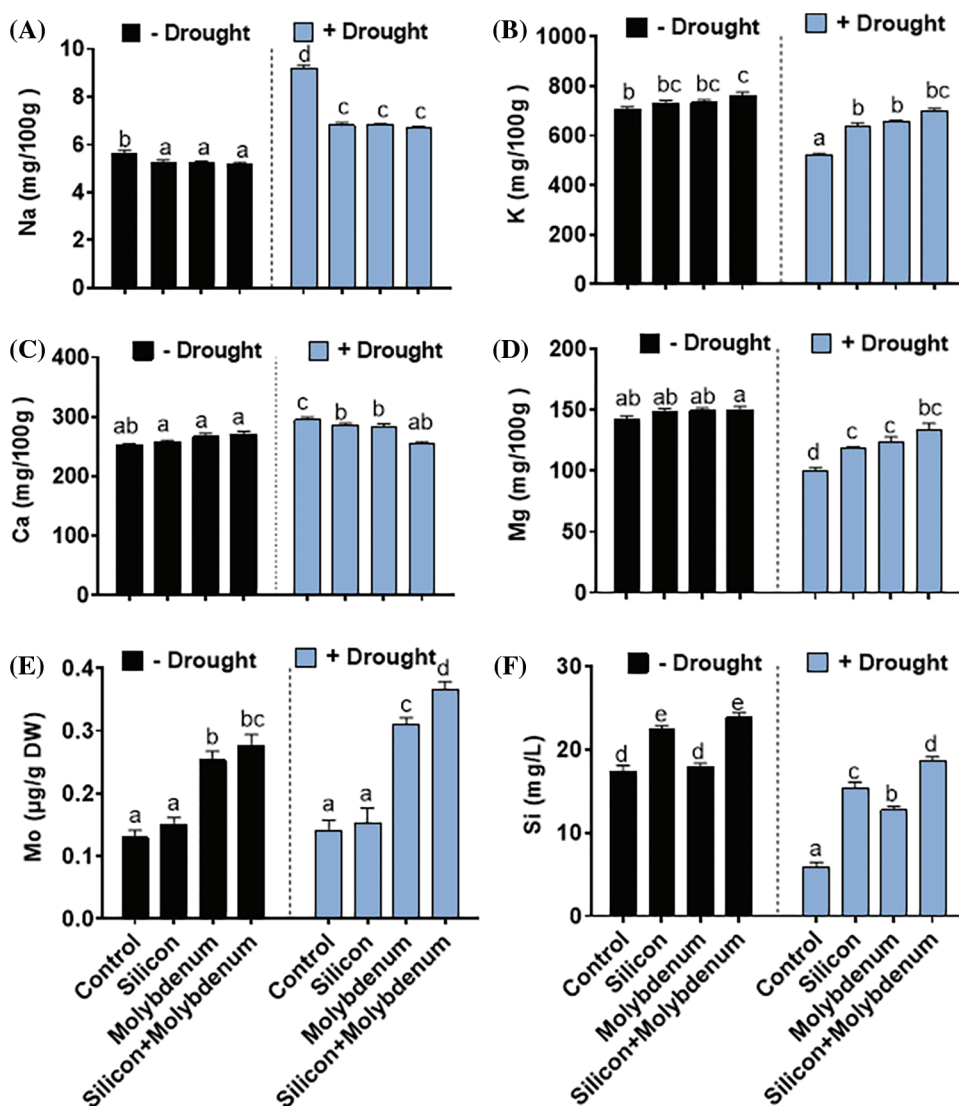
Under drought stress, the quantitative results exposed statistically significant promotion in the catalase enzyme [107/71 (mg/g FW) (control)], compared to control (C) without stress. Nevertheless, silicon and molybdenum combined supplementation exhibited a statistically significant further promotion in catalase enzyme [128/107 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 7E). Under drought stress, the quantitative results exposed statistically significant promotion in the ASA content [0.68/0.51 (mg/g FW) (control)], compared to control (C) without stress. Nevertheless, silicon and molybdenum combined supplementation exhibited a significant ( $p \leq 0.05$ ) further promotion in ASA content [0.85/0.68 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 7F).

Under drought stress, the quantitative results exposed a statistically significant promotion in the Total antioxidants [0.35/0.27 (mg/g FW) (control)], compared to control (C) without stress. Silicon and molybdenum combined supplementation exhibited a significant ( $p \leq 0.05$ ) further promotion in total

antioxidants [0.5/0.35 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 7G).

### 3.7 Effect of Silicon and Molybdenum on Mineral Status of Quinoa under Drought Stress

During the current investigation, it was found that drought stress caused a significant ( $p < 0.05$ ) over-accumulation of  $\text{Na}^+$ , and  $\text{Ca}^{2+}$ , and concentration and a reduction of  $\text{K}^+$ , and  $\text{Mg}^{2+}$  in quinoa compared to control. Under drought stress, the quantitative results exposed a statistically significant promotion in the  $\text{Na}^+$  content [9.4/5.7 (mg/g FW) (control)], compared to control (C) without stress. Nevertheless, under drought stress, silicon and molybdenum combined supplementation exhibited a significantly ( $p \leq 0.05$ ) decreased  $\text{Na}^+$  content [6.8/9.4 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 8A).



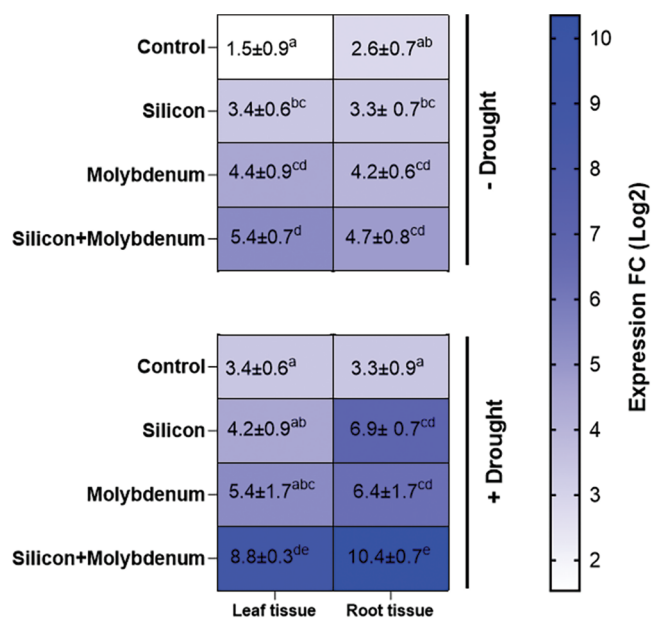
**Figure 8:** Foliar spray effect of silicon and molybdenum on ionic concentration of *Chenopodium quinoa* under drought stress. Sodium content (A) potassium content (B) calcium content (C) magnesium content (D) molybdenum content (E) and silicon content (F). Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$

Under drought stress, the quantitative results exposed a statistically significant decrease in the  $K^+$  content [523/710 (mg/g FW) (control)], compared to control (C) without stress. Silicon and molybdenum combined supplementation exhibited a significantly ( $p \leq 0.05$ ) elevated  $K^+$  content [697/523 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 8B). Under drought stress, the quantitative results exposed a statistically significant promotion in the  $Ca^{2+}$  content [304/257 (mg/g FW) (control)], compared to control (C) without stress. Silicon and molybdenum combined supplementation exhibited a significantly ( $p \leq 0.05$ ) elevated  $Ca^{2+}$  content [697/523 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 8C). Under drought stress, the quantitative results exposed a statistically significant promotion in the  $Mg^{2+}$  content [99/146 (mg/g FW) (control)], compared to control (C) without stress. Silicon and molybdenum combined supplementation exhibited a significantly ( $p \leq 0.05$ ) elevated  $Mg^{2+}$  content [123/94 (mg/g FW) (drought control)] in quinoa under drought stress, comparable to the respective control plants (Fig. 8D–F).

### 3.8 Quantitative Gene Expression Analysis of *CqSnRK2.10* under Drought Stress

To measure the level of expression of selected drought stress marker genes, the RT-qPCR method was used for evaluating the induction of *sucrose non-fermenting1 (SNF1)-associated protein kinase 2 (SnRK2.10)*, which belongs to a comparatively small plant-specific family of serine/threonine (Ser/Thr) protein kinases [11]. In the present study, gene expression analysis for the *CqSnRK2.10*, in the leaf tissues of quinoa grown under drought revealed a statistically significant promotion in *CqSnRK2.10* expression [(3.8-fold/1.9-fold (control)]. Under drought stress, silicon, and molybdenum combined supplementation resulted in a further promotion in *CqSnRK2.10* expression [(8.5-fold/3.8-fold (drought control)] in leaf tissue.

Moreover, gene expression analysis for the *CqSnRK2.10*, in the root tissues of quinoa grown under drought, revealed a statistically significant promotion in the expression [(3.5-fold/2.1-fold (control)]. However, under drought stress, silicon and molybdenum combined supplementation resulted in a further promotion in expression [(10-fold/3.5-fold (drought control)] in root tissue (Fig. 9).



**Figure 9:** Expression profiling of *CqSNRK2.10* gene measured by RT-qPCR using leaf and root tissue of quinoa foliar supplemented with silicon and molybdenum, under drought stress. Numeric data is represented by means and standard errors of three biological replicates with at least four technical replicates, marked with letters denoting significant differences at  $p \leq 0.05$



#### 4 Discussion

Water scarcity is a critical factor that prevents plants from growing, especially during the vegetative and reproductive phases of plant growth of susceptible plants such as quinoa. However, the exploitation of plant drought stress tolerance using the exogenous application of drought stress tolerance mediators and micronutrients, silicon (Si) and molybdenum (Mo) has shown restoration of the sustainable growth of quinoa under drought stress in the present study. Plant nutrition plays an important function in maintaining healthy growth and augmenting stress tolerance. Micronutrients, such as Si and Mo, have been shown to provide tolerance to plants against various stresses [22].

Under drought stress, Si application provides plant tolerance through various mechanisms. Si application can up-regulate aquaporin genes (PIP) and mitigate ROS-induced aquaporin activity inhibition in plants. By raising the concentration of soluble carbohydrates and amino acids contents in the xylem sap, which increases osmotic potential, or by triggering  $K^+$  transport to the xylem sap via the SKOR gene impacting the osmoregulation [49]. Another trace element that is present in soil, and is essential for plant growth is molybdenum (Mo). It exists in different oxidation states, with the most prevalent form found in agricultural soils being Mo (VI). The primary form of molybdenum that is accessible to plants is molybdate. Despite having a major impact on several redox processes, molybdenum is only necessary in very small amounts when compared to other vital micronutrients [50]. Molybdenum is utilized by specific plant enzymes as part of the molybdenum co-factor to participate in oxidation-reduction reactions. Mo is required for the activity of more than 50 enzymes, including five found in plants, involved in nitrogen, sulfur, purine, and phytohormone metabolism [51]. It is part of several enzymes, including nitrate reductase (NR), xanthine dehydrogenase/oxidase (XDH), aldehyde oxidase (AO), and sulphite oxidase (SO), which are involved in nitrogen fixation and assimilation, purine catabolism, phytohormone synthesis, and sulfur metabolism, respectively [52]. Consistently, the present research also showed that under drought stress, individual, as well as combined supplementation of silicon and molybdenum, improved the vegetative and reproductive growth of quinoa in terms of plant length, biomass, seed growth, and yield.

Present results align with previous studies, demonstrating a significant reduction in various growth attributes under drought stress in different plant species, as drought stress disrupts plant metabolism, particularly photosynthetic pigments, by generating excessive reactive oxygen species (ROS) [53]. The decrease in chlorophyll content observed under water shortage is primarily attributed to protein degradation in chloroplast membranes caused by ROS overproduction [54]. Relative water content (RWC) reflects the ability of cell membranes to withstand environmental cues, including drought stress [55].

In the present investigation, comparable to other studies, a decrease in RWC under water shortage conditions is observed. This decline in RWC indicates a loss of turgor, limiting water availability for cell expansion and subsequently suppressing plant development and growth. RWC is considered a reliable indicator of drought stress tolerance in plants. Proline accumulation as a result of water stress has been well-documented as a mechanism to maintain leaf tissue homeostasis in various plant species. In addition to this, the accumulation of sugars serves as a self-protective strategy employed by plants against adverse environmental conditions, and current results also showed a promotion in total soluble sugars and non-reducing sugars under water shortage conditions, in line with the conclusions of previous reports that sugars act as osmoprotectants and confer water stress tolerance in plants. In the present study, drought-stress individuals as well as combined supplementation of silicon and molybdenum, elevated the photosynthetic activity, and relative water content of quinoa grown under drought stress.

It is also known that Si supplementation can improve root hydraulic conductance, modify root growth, increase root/shoot ratio, elevate aquaporin activity, and enhance osmotic driving force. This, in turn, results in elevated water uptake and transport, leading to higher photosynthetic rates and improved plant resistance

to water deficiency [56]. Si application can also alter gas exchange attributes in plants enhance antioxidant defense by increasing the biochemical activities, and protect seedlings from oxidative damage. Furthermore, Si In several crop species, application under water-scarce circumstances boosts photosynthetic rate, leaf and root water and osmotic potential, and water usage efficiency, while lowering transpiration activity and membrane trafficking [57]. Molybdenum (molybdate) is also essential for plant development and plays a vital role in physiological processes [22].

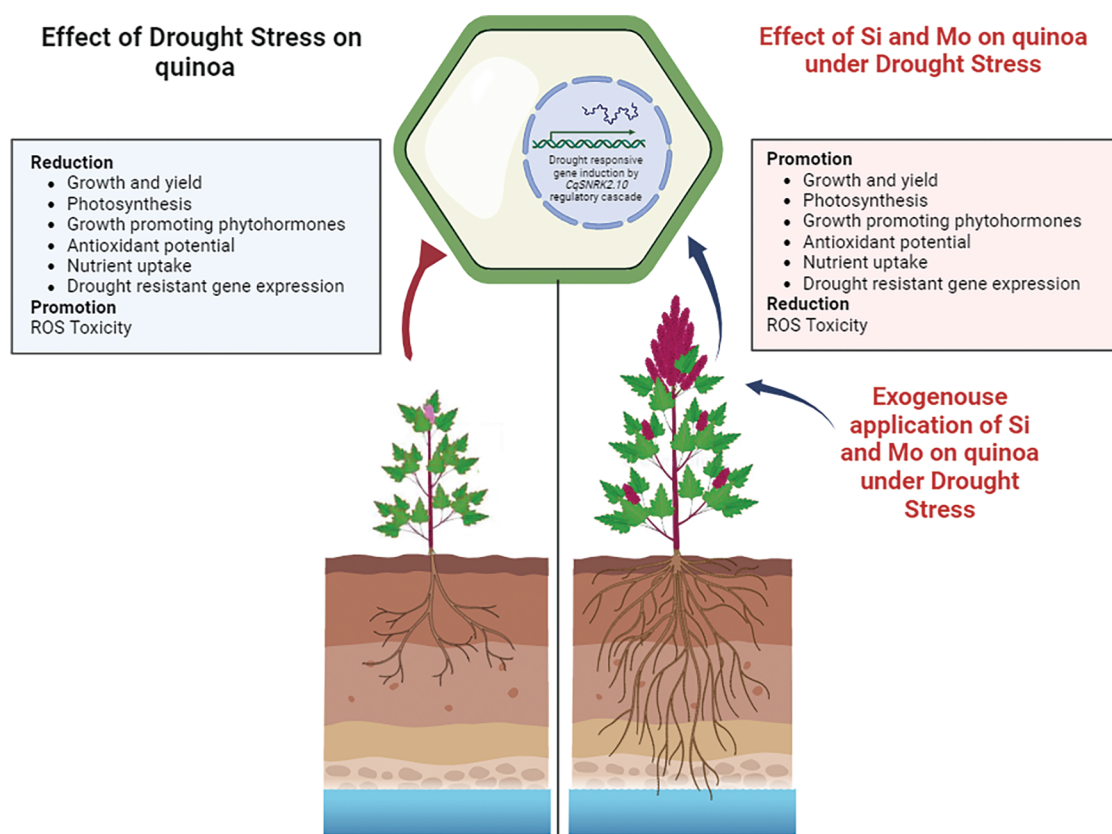
Si has been widely reported to decrease oxidative damage by enhancing the activities of antioxidant enzymes (SOD, APX, CAT, and POD) under drought stress in wheat, sunflower, and tomato plants [58]. Si pretreatment has been shown to up-regulate the expression of drought-specific genes in rice during drought stress [59]. Molybdenum is essential for promoting the production of enzymes including POD, CAT, and SOD as well as raising the proline concentration in cells, thus supporting the plant defense system and playing an important role in mung bean plants under water stress conditions [22]. In the present study, individual as well as combined supplementation of silicon and molybdenum, *also* elevated the antioxidant capacity of quinoa grown under drought stress, thus reducing ROS production and oxidative damage.

Si and Mo translocation in plant tissues also affect the uptake of mineral ions ( $\text{Na}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{K}^+$ , and  $\text{Mg}^{2+}$ ), as it is known that the translocation of one type of micronutrient influences the translocation of others. For example, the translocation of silicon is reported to mitigate the concentration of copper in root and shoot in *A. thaliana* in a water-based culture. Thus, silicon translocation reduces the excessive copper concentration in plant tissue to reduce metal toxicity. The transport of molybdenum aids in the transportation of the micronutrients in the plant's vascular tissues and leaf buds [60]. Different mineral elements also play a crucial role in physiological role in plant development and growth. For example,  $\text{K}^+$  plays a crucial role in water-use efficiency, stomatal control, aerial and underground biomass, and photosynthesis [61]. It also serves as a mineral osmolyte that regulates turgor and osmotic pressure to facilitate plant growth, cell enlargement, and leaf stomata opening and closing. In *Carthamus tinctorius*,  $\text{K}^+$  elevated the leaf's relative moisture content, while magnesium decreased it [62]. The  $\text{Mg}^{2+}$  concentration was affected by drought stress, with a decrease observed in plant leaves as drought stress elevated [63,64]. The same outcomes were reported. in tomatoes, where drought led to a decrease in Mg concentration [65].  $\text{Mg}^{2+}$  levels significantly influence chlorophyll content but exert a negative impact on  $\text{Ca}^{2+}$  levels in safflower [62]. With the severity of drought stress increasing,  $\text{Ca}^{2+}$  concentration decreased, with the lowest concentration being seen in control plants. Similar findings were reported in tomatoes, where water stress reduced  $\text{Ca}^{2+}$  concentrations [65].  $\text{Ca}^{2+}$  ions act as Essential alternative messengers are crucial for different phases of plant development and growth and trigger physiological changes due to drought stress.

Additionally,  $\text{Ca}^{2+}$  is essential for controlling the development of polar tissues and cells as well as plant stress tolerance. It influences the response of plants to drought stress and ABA-induced stomatal closure in plants [66,63,67].  $\text{Na}^+$  concentration elevated with increasing drought stress, with the lowest concentration observed in control plants. The water usage potential of olive leaves altered during drought stress, and elevated  $\text{K}^+$  and  $\text{Na}^+$  levels enhanced the starch content of the leaf.  $\text{Na}^+$  influenced metabolism and the photosynthetic response favorably when  $\text{K}^+$  levels were insufficient [63,64].  $\text{Ca}^{2+}$  concentration elevated with the intensity of drought stress, with the lowest concentration observed in control plants Similar findings were reported in tomatoes, where water scarcity triggered a reduction in  $\text{Ca}^{2+}$  concentrations [65].  $\text{Ca}^{2+}$  ions serve as significant signaling molecules, and messengers that, in response to the stress of drought, promote physiological mechanisms positively, and are essential for various phases of plant development and growth.  $\text{Ca}^{2+}$  plays a significant part in controlling the development of polar tissues and cells as well as plant stress tolerance through adaptability. It affects abscisic acid  $\text{Ca}^{2+}$  concentration elevated with the intensity of drought stress, with the lowest concentration observed in control plants Similar findings were reported in tomatoes, where water stress reduced  $\text{Ca}^{2+}$  concentrations [65,67].  $\text{Ca}^{2+}$

ions act as important signaling molecules that stimulate physiological functions as a result of drought stress and are essential for various stages of plant development and growth.  $\text{Ca}^{2+}$  also plays a key role in regulating polar cell and tissue growth and plant adaptation to stress.

The present study revealed that individual as well as combined supplementation of silicon and molybdenum readjusted the mineral status of quinoa by affecting root-to-shoot transportation. Various transporters have been found in plants up to this point. For instance, several transporters, such as the high-affinity  $\text{K}^+/\text{Na}^+$  transporter (HKT) and the  $\text{K}^+$  transporter (KT), are used to move the ion  $\text{K}^+$ ,  $\text{Ca}^{2+}$ -ATPases,  $\text{Ca}^{2+}/\text{H}^+$  exchanger transports, and transporters  $\text{Ca}^{2+}$  permeable ion channels are used to move the calcium (Ca). Magnesium (Mg) is transported throughout the plant system by the magnesium transporter (MGT). Molybdenum (Mo) is transported throughout the plant via the molybdenum transporter type (MOT). According to a review [60], the molybdenum transporter type 1 (MOT1) is more prevalent in plants. Previously, exogenous Si application was up-regulated. Numerous monocot and dicot plant species have been found to contain the Si-transporter aquaporin genes *AQPs* [68]. In the current study, Si and Mo induced the expression of drought-resistant marker gene (*CqSnRK2.10*) serine/threonine (Ser/Thr) protein kinases in quinoa grown under drought, implying a crucial role in the regulation of drought stress resistance due to the induction of *CqSnRK2.10* expression, triggering the signaling mechanism for readjustment of drought stress tolerance response in quinoa, and the overall findings of present work have been depicted in graphical abstract in Fig. 10.



**Figure 10:** Graphical presentation showing the role of exogenous application of Si and Mo in quinoa under stress. Drought stress inhibited plant growth, while the combined application of Si and Mo ameliorated the drought stress tolerance and promoted the growth and yield by activating the expression of *CqSNRK2.10* (a drought-resistant marker gene), for induction of drought tolerance through metabolic, antioxidant, and ionic reshuffling and rebalancing in quinoa

## 5 Conclusion

Current research concludes the prospective significance of individual and combined supplementation of silicon and molybdenum for growth promotion and drought stress alleviation in quinoa. This is achieved by triggering the signaling mechanism driven by the drought-resistant marker gene (*CqSnRK2.10*) serine/threonine (*Ser/Thr*) protein kinases. Consequently, silicon and molybdenum supplementation readjusted the physiological, biochemical, antioxidant, hormonal, and mineral status of quinoa plants under drought stress to a level well-suited for optimal growth, development, and yield.

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**Availability of Data and Materials:** The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

**Ethics Approval:** This article does not contain any studies with human participants or animals by any of the authors.

**Conflicts of Interest:** All authors declare that they have no conflicts of interest to report regarding the present study.

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