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Evaluation of Pre-Harvest Sprouting (PHS) Resistance and Screening of High-Quality Varieties from Thirty-Seven Quinoa (*Chenopodium quinoa* Willd.) Resources in Chengdu Plain

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ABSTRACT

Pre-harvest sprouting (PHS) will have a serious effect both on the yield and quality of quinoa (Chenopodium quinoa Willd.). It is crucial to select and breed quinoa varieties with PHS resistance and excellent agronomic traits for guidance production and utilization of quinoa. A comprehensive evaluation of the PHS resistance and agronomic traits of 37 species of quinoa resources was conducted in Chengdu Plain. The evaluation used various methods, including grain germination rate (GR), grain germination index (GI), total spike germination rate (SR), total grain germination index (SI), grey correlation analysis (GCA), cluster analysis and correlation analysis. Results showed significant differences in PHS resistance and agronomic traits amongst the 37 quinoa resources. CDU-23 was most resistant to PHS within 24 h, with a germination rate of 2.67% and 0% according to the GR and SR results, respectively. However, in the same time, CDU-31 showed the maximum susceptibility to PHS based on the SR of 31.07%, while CDU-34 was the most sensitive to PHS according to the GR of 100%. The comprehensive evaluation identified one and nine kinds of high resistance species for grain and whole spike germination, respectively. In both cases, the coefficients of variation (CV) for these parameters were 34.78% and 82.13%, respectively. GCA results showed that the magnitude of the association between each trait and yield in the thirty-seven quinoa resources was in the following order: thousand grain weight > seed length > seed area > seed width. Although the seed weight of CDU-18 reached 3.7010 g, the seed weight of CDU-5 was only 1.6030 g. However, the size of the seeds, their width and area did not correlate with their 1000-grain weight. There was a complex correlation between PHS resistance index and agronomic traits. Based on clustering analysis, thirty-seven quinoa resources were classified into three taxa. It was found that various taxa differed in PHS resistance and agronomic traits. Several comparisons of the aggregated data led to the selection of five varieties of quinoa, of which CDU-2 presented excellent agronomic qualities and strong PHS resistance. This study has provided a reference for breeding excellent quinoa varieties with PHS resistance.

KEYWORDS

Quinoa; pre-harvest sprouting (PHS); agronomic traits; cluster analysis



1 Introduction

Quinoa (Chenopodium quinoa Willd.), an annual dicotyledonous plant of the Chenopodium genus in the Amaranthaceae family, is native to the Andes region of South America and an agricultural crop similar to grains [1]. About 7000 years ago, quinoa was domesticated and cultivated by the local population as a main food crop [2]. Quinoa can withstand extreme conditions that common crops (rice, wheat, corn, sorghum) cannot survive [3], including drought salinity and frost tolerance, and can be grown at high altitude areas. Quinoa is rich in starch (52.0%-69.0%), lipids (2.0%-9.5%), protein (13.8%-16.5%), dietary fiber (7.0%-9.7%) [4] and nutrients that meet the body's daily intake [5]. Despite the high cultivation, nutritional and functional value of quinoa, it was neglected for thousands of years [6,7]. Until the last 50 years, as the global population grew dramatically and the natural environment and agricultural production conditions became increasingly severe, there was an urgent need to find a high-quality grain to satisfy the daily consumption of people [8]. In recent years, quinoa has been rediscovered due to its excellent agronomic property and nutritional value, so its area of cultivation and yield are rapidly increasing. In recent years, quinoa has been grown in China, France, India, Sweden, Denmark, Netherlands, Italy, and other countries [9,10]. Due to the Andean people's contribution to the development of quinoa, the International Food and Agriculture Organization (FAO) has declared 2013 the "International Year of Quinoa" [1].

However, in Southwest China [11], especially Chengdu Plain, the harvest season of quinoa is often accompanied by high temperatures and high humidity, and Quinoa is extremely vulnerable to PHS if it is not harvested in time or not handled properly after harvested [8,12,13]. PHS will promote the hydrolysis of starch in grain endosperm [8,14,15], the change of protein spatial structure, the damage of seed internal structure which reduce the nutritional value of grain, create a favorable environment for saprophytic fungi, and lead to seed spoilage and deterioration [16] and loss of vitality. The annual global economic loss due to PHS is up to \$1 billion [15]. Consequently, it is crucial to find a solution to the quinoa PHS method as soon as possible, both in terms of boosting agricultural production and promoting economic development.

Through natural variation and artificial selection, selective breeding is an effective method of evaluating PHS resistance of quinoa resources with excellent resistance in years of cultivation (the conventional methods to identify pre-harvest sprouting resistance mainly include whole-spike germination test and grain germination test) [17–19]. Varieties with excellent agronomic characters are selected from the screened resources for planting, so as far as possible to reduce the grain PHS caused by agricultural production loss. Quinoa yield is generally determined by effective spike length, number of spike branches and thousand grain weight [20]. Thousand grain weight is closely related to the grain phenotypic traits, which can be divided into grain size (grain area, grain width, grain length) and grain morphology (aspect ration, roundness value) [21,22]. The improvement of quinoa yield and quality can be achieved by improving the phenotypic traits of the grain of quinoa. In this study, varieties with PHS resistant were selected through grain germination and whole spike germination experiments, varieties with high quality agronomic traits were selected through thousand grain weight, grain length, grain width and grain area. The data were integrated and 37 quinoa resources were analyzed, and the excellent quinoa varieties suitable for planting in the Chengdu Plain were screened, which provided a reference for developing quinoa resources and improving quinoa yield.

2 Materials and Methods

2.1 Preparation and Treatment of Test Materials

In this experiment, 37 quinoa resources were provided by the Key Laboratory of Coarse Cereal Processing, Ministry of Agriculture and Rural Affairs at Chengdu University. Approximately 100–110 days were the most fertile period for the thirty-seven quinoa resources. The experimental site was

located in Yuanba Village, Xinshi Street, Jianyang City, East New District, Chengdu City, Sichuan Province $(104^{\circ}56'59''N, 30^{\circ}32'90''E; 387 \text{ m})$. The average temperature of the test site from March to July is 25°C in two years. All the materials were sowed in a plot of measured 3 m × 3.3 m, with a 30 cm × 30 cm plant spacing, cultivated 1 quinoa plant every 30 cm and cultivated 11 quinoa plants within 3.3 m. The experiments were conducted in 2021 and 2022, respectively, and three biological replicates were performed in each year. All the quinoa materials were sowed on March 15 and were harvested on July 11 in the first year and sowed on March 8 and were harvested before July 10 in the second year. Conventional field management was adopted during the growth period. During harvest, each plot was divided into five parts, and a top spike approximately 15 cm long was randomly selected from each part. Hence, each quinoa material was numbered and five spikes were selected in a plot.

All the quinoa spikes were dried in an oven at 37°C for 72 h until the moisture content of the grain less than 12%. An evaluation of spike germination was conducted on two spikes of each quinoa. Ninety grains were randomly selected from the remaining dried quinoa spikes for the grain germination test after manual threshing. Furthermore, 37 quinoa resources were assessed for their grain agronomic traits (including thousand grain weight, grain length, grain width, and grain area).

All the test materials were disinfected by immersion with 0.5% NaClO (grains immersion for 3 min and spikes immersion for 10 min), rinsed slowly with distilled water to remove residual reagents. A dark climate chamber at 25°C and 85% relative humidity was used to test germination of grains and spikes [23].

2.2 Germination Test

2.2.1 Whole Spike Germination Test

A whole spike germination test was conducted using wet tissue wrapping [23,24]. For each spike, wet paper towels were wrapped around it and it was placed upright in a 250 mL beaker for one hour until completely moist. During incubation, the samples were placed in a 25°C artificial climate chamber. In order to prevent the paper from drying out, 10 mL of distilled water was added to the beaker. The germination status of the spikes in the paper towels was observed every four hours. The 100% humidity of the paper towels was maintained at all times to provide a moist environment for the germination of quinoa spikes. The number of germinated seeds was observed and counted at 8, 16, 24, 32, 40, 48, 72 and 96 h respectively (the grain coat was considered to be broken or buds of 1–2 mm appeared named germinate) [25] and the whole spike germination rate (SR represents the percentage of PHS grains in total grains) and index (SI represents the average germination degree of quinoa PHS) were calculated. Among them, the whole spike germination resistance test was conducted within 8–48 h and the whole spike vitality test was carried out at 72, 96 and 120 h.

2.2.2 Grain Germination Test

Each treated grain of quinoa was placed in a Petri dish covered with double filter paper (diameter 9 cm), 30 seeds were placed in each dish and 5 mL of distilled water was added, and the process was repeated three times. The Petri dishes were observed every two hours in a constant light incubator. Each grain germination rate (GR) and grain germination index (GI) was recorded every 4 h for 32 h on the filter paper with a moisture level of 100%. Subsequent, seed vitality test was carried out at 44, 56 and 68 h.

2.2.3 Agronomic Trait Determination

The length, width and area of each quinoa resource was performed using a WinRHIZO Reg STD4800 root system analyzer, while the thousand grain weight was determined with an electronic analytical balance FA2004.

2.2.4 Statistics and Analysis

Germination rate (%): GR = $n_{(1 \sim k)}/N \times 100\%$.

Germination index: GI = $(K \times n_1 + \dots + 3 \times n_{(k-2)} + 2 \times n_{(k-1)} + 1 \times n_k)/K \times N$ [26].

In the formula: $n_{(1 \sim k)}$ denotes the number of grains sprouting each time from the 1st to the next k times. The k represents the total number of germination tests. The N represents the total number of grains used for germination.

A modified version of the standard method for wheat spike germination resistance (NY/T1739-2009) was used for the whole spike germination resistance test [27,28]. References to relevant literature were used to develop the grading method for quinoa grains germination resistance in Table 1. The results of dormancy grade and grain germination resistance grade are consistent.

Resistance level	Classifica	tion criteria
	Germination resistance of grains	Germination resistance in spikes
High resistance (HR)	$GR \le 10\%$	$GI \le 5\%$
Resistance (R)	$10\% < GR \le 30\%$	$5\% < GI \le 20\%$
Moderately resistance (MR)	$30\% < GR \le 50\%$	$20\% < GI \leq 40\%$
Moderately susceptible (MS)	$50\% < GR \le 70\%$	$40\% < GI \leq 60\%$
Highly susceptible (HR)	70% < GR	60% < GI

Table 1: Resistance grading criteria

Coefficient of variation (CV) is defined as mean standard deviation (SD)/mean value (Mean) × 100%.

PHS resistance indicators were used for cluster analysis. Agronomic traits were used for grey correlation analysis and cluster analysis.

All the results were the average of two years. All statistical results were analyzed and plotted using Microsoft Excel 2019 software, SPSS Statistics 25 and Origin 2021. All data were averaged, and in this study, single factor ANOVA test method were used for significance analysis.

3 Results and Analysis

3.1 Evaluation and Analysis of Whole Spike Germination Trials

The results of the whole spikes germination trials were shown in Table 2. There were significant differences in whole spike germination resistance within 48 h among 37 quinoa resources treated under the same conditions. According to the standard, nine highly resistant (HR) varieties, seven resistant (R) varieties, nine moderately resistant (MR) varieties, seven moderately susceptible (MS) varieties and five highly susceptible (HS) varieties were selected. Among high resistance (HR) varieties, the spike germination index was in the order of CDU-23 < CDU-15 < CDU-1 < CDU-36 < CDU-5 < CDU-2 < CDU-18 < CDU-6 < CDU-14. CDU-23 spike had the lowest germination rate of 1.05% and germination index of 0.44%. It was followed by CDU-15 with 1.35% and 0.70% spike germination rate and index. The spike germination rate and index of CDU-1 were 2.33% and 1.56%, respectively. The germination rate of CDU-23 spikes was 0% during the first 32 h, and slowly increased to 1.05% within next 16 h. The germination process of CDU-15 was similar to that of CDU-23, with 0% of spikes germinating in the first 8 h and rose to 1.35% within next 40 h and remained unchanged. For CDU-1, the spike germination rate was 0% during the first 16 h and rose to 2.33% in the subsequent period. In a comprehensive analysis, CDU-31 had the lowest spike germination resistance among the 37 quinoa resources, followed by CDU-16 and CDU-35, which the spikes germination rates were 76.43%, 75.63% and 56.99%, respectively. CDU-31 and CDU-16 had 0 germination in the first 8 h, while CDU-35 had the same germination in the first 8 h. However, the spike germination rate of the three varieties

aforementioned gradually increased in the subsequent time, and by 24 h, the spike germination rate had far exceeded that of the other varieties.

3.2 Evaluation and Analysis of Grain Germination Trials

Table 2 showed that there were significant differences in grain germination rates within 32 h among 37 quinoa resources treated under the same conditions. According to the standard, CDU-23 was screened as a highly PHS resistance (HR) variety with the grain germination rate and index of 2.67%. For CDU-23, there were zero grain germinated in the first 16 h and the germination rate increased to 2.67% in the second 16 h with no change in the subsequent time. CDU-2 was selected as a resistant (R) variety with a grain germination rate of 16.00% and grain germination index of 7.83%. Moreover, the germination process of CDU-1 was similar to that of CDU-23, with no germination during the first 16 h and no change after the germination rate reached 16.00% in the subsequent time. Twenty out of thirty-seven quinoa materials were screened as high-sensitive (HS) varieties. The top three grain germination rates were CDU-34, CDU-26 and CDU-31. According to the above sequence, with 100.00%, 97.33% and 96.00% of germination rates and 260.33%, 227.17% and 200.83% of germination indices, respectively. The CDU-34 grains had all sprouted within 20 h, while CDU-26 and CDU-31 reached the peak within 24 and 28 h, respectively. CDU-15 was selected as one of the medium PHS resistant (MR) varieties, which germination rates and germination indices were 45.33% and 77.33%. Moreover, when CDU-15 was subjected to a 32 h of germination treatment, its germination rate was the highest and tended to increase continuously.

3.3 Resistance Analyses of Grain Germination Rate and Whole Spike Germination Index

As a statistical indicator of seed germination resistance, grain germination rate indirectly indicated the dormancy resistance of each resource in Table 3. The variation in grain germination resistance among the 37 quinoa resources ranged from 2.67% to 100.00%, with a coefficient of variation of 34.78% and germination was discrete and highly variable among the species. There were only one highly resistant (HR) and one resistant (R) variety in seed germination, which were CDU-23 and CDU-2, respectively. There were twenty seed germination high susceptibility (HS) varieties, accounting for 54.05% of the total resources, with a variation range of 72.00%–100.00%, which was the most representative part. However, the coefficient of variation of the high-sensitive (HS) varieties was only 9.64% with a good degree of dispersion, indicating that the differences in germination moderately susceptible (MS) varieties with the least variation range from 53.33% to 65.33% and the least dispersion of coefficient of variation of 7.60%, indicating that the germination rates of each moderately susceptible variety were very close.

The whole-spike germination index was used as a grading index. In the whole spike germination resistance results, the variation of germination index of 37 quinoa resources ranged from 0.44% to 81.50% with a coefficient of variation as high as 82.13%, indicating that there were significant differences in quinoa spike resistance among the resources. The coefficient of variation of the medium-sense (MS) variety of spike germination was the smallest at 11.30%, and the range of variation was also small at 41.73% to 56.30% for a total of seven resources. The second was the highly sensitive (HS) varieties with a slightly larger coefficient of variation. There were five varieties with a coefficient of variation for 13.18%. Spike germination high resistance (HR) varieties were the most numerous, with a total of nine varieties and a minimum variation range of 0.44% to 4.66%, but the coefficient of variation was as high as 61.86%. Among highly resistant (HR) varieties, spike seed germination resistance differed significantly among highly resistant (HR) varieties, while highly resistant (HR) varieties differed significantly in spike seed vigor, they were all capable of inhibiting PHS to varying degrees.

Resources		Whole s _f	oike gern	Whole spike germination rate (%)	e (%)	Res	ance			U	Grain germination rate (%)	ination rat	e (%);			Dormat	Dormancy Germination	
	8 h 16 h 24 h	1 32 h	40 h	1 48 h	h 72 h 96 h	h 120 h	de ranking	4 h 8 h	12 h	16 h	20 h 2	24 h 2	28 h 32	h 44 h	56 h	68 h grade	rate ranking	germination index (%)
CDU-1	0.00a 0.00a 0.10a	a 0.49a	1.25a 1.25a		2.33a 10.97 17.67	.67 62.69 HR	35	0.00d 1.33d	10.67d	34.67c 4	48.00b 5	53.33ab (62.67a 64.	64.00a 72.67	84.00	100.00 MS	24	1.56/98.33
CDU-2	0.00c 0.00c 0.00c	c 1.27c	s 4.67b		9.21a 18.94 30.45	.45 74.39 HR	32	0.00d 0.00d	0.00d	0.00d	1.33cd (6.67bc	10.67ab 16.00a	00a 36.33	42.67	100.00 R	36	3.87/7.83
CDU-3	0.00b 1.48b 4.75b		10.29ab 14.74ab		25.91a 39.47 50.33	.33 100.00 R	22	1.33e 2.67e	4.00e	13.33de	28.00cd 3	34.67bc 5	52.00ab 57.33a	33a 71.33	80.00	100.00 MS	25	19.81/62.17
CDU-4	0.18d 0.78cd 1.79cd	cd 5.49bc	oc 9.19b		13.40a 28.69 42.27	.27 85.73 R	26	0.00d 1.33d	6.67cd	22.67c	65.33b (68.00b 8	85.33a 89.	89.33a 100.0	100.00 100.00 1	100.00 HS	14	10.11/111.00
CDU-5	0.00b 0.29b 0.51b	b 0.98b	o 1.87bc		4.00a 13.33 26.04	.04 68.67 HR	33	0.00c 2.67c	8.00bc	14.67abc	14.67abc 16.00abc 22.67ab		22.67ab 32.00a	00a 47.33	64.67	100.00 MR	35	1.89/43.67
CDU-6	0.50c 0.63bc 0.79bc	bc 1.36bc	oc 2.25ab		3.42a 11.46 24.98	98 67.79 HR	30	1.33d 2.67d	5.33d	9.33d	21.33c 3	32.00bc 3	37.33b 54.	54.67a 66.67	78.67	100.00 MS	28	3.96/52.33
CDU-7	0.00c 0.35c 5.65c		13.66bc 22.41ab		.23a 50.23 64.	35.23a 50.23 64.29 100.00 MR	21	0.00e 0.00e	4.00e	10.67de	17.33cd 2	29.33bc 3	36.00ab 48.00a	00a 61.33	70.67	100.00 MR	31	23.32/44.33
CDU-8	0.00c 0.40c 2.73c		13.75b 16.82ab		24.15a 38.29 48.67	.67 100.00 R	23	4.00d 12.00cd	25.33c	45.33b (60.00ab 6	68.00a	73.33a 73.	73.33a 100.0	100.00 100.00 1	100.00 HS	19	18.34/144.83
CDU-9	0.00c 3.45bc 14.61abc 22.03ab 28.01a	labc 22.03	ab 28.0		.57a 54.28 68.	35.57a 54.28 68.78 100.00 MS	8	1.33f 22.67e	60.00d	69.33c	76.00b 8	89.33a 9	92.00a 93.	93.33a 100.0	100.00 100.00 100.00 HS	100.00 HS	10	53.64/215.67
CDU-10	0.00c 0.00c 22.46bc	6bc 37.25	37.25ab 46.68ab		61.19a 76.49 89.63	.63 100.00 HS	4	0.00d 2.67d	13.33d	36.00c	48.00bc 5	57.33b (61.33b 85.	85.33a 100.0	100.00 100.00 100.00 HS	100.00 HS	15	63.08/158.33
CDU-11	0.00d 0.18d 1.50cd		8.97c 18.87b		.85a 52.37 66.	36.85a 52.37 66.18 100.00 R	24	0.00d 0.00d	0.00d	4.00cd	8.00cd	12.00c 2	25.33b 41.	41.33a 54.67	68.67	100.00 MR	34	17.93/22.50
CDU-12	0.00c 0.18c 7.38bc		10.54abc 21.55ab		28.28a 43.78 59.45	.45 100.00 MR	18	0.00c 0.00c	5.33c	21.33bc	37.33abc 52.00ab		56.00ab 64.00a	00a 80.00	87.33	100.00 MS	22	23.92/77.50
CDU-13	0.00b 0.60b 12.32ab		22.43ab 36.37ab		.94a 66.13 80.	50.94a 66.13 80.94 100.00 MR	. 13	0.00e 1.33e	6.67e	17.33d	30.67c ²	42.67b ²	46.67b 65.33a	33a 80.67	90.00	100.00 MS	21	39.93/68.17
CDU-14	0.14b 0.18b 0.40b		2.15b 4.78b		10.71a 19.13 27.68	.68 70.38 HR	29	0.00d 0.00d	8.00d	29.33c	44.00b ²	49.33b	54.67ab 64.00a	00a 78.00	85.33	100.00 MS	22	4.66/56.50
CDU-15	0.00c 0.02c 0.02c		0.36bc 0.70b		15a 11.17 23.	1.35a 11.17 23.42 67.58 HR	36	1.33d 5.33d		12.00cd 25.33bc	33.33ab 3	38.67ab 3	38.67ab 45.33a	33a 59.33	70.00	100.00 MR	32	0.70/77.33
CDU-16	0.00d 3.25d 36.43c		47.48bc 63.03ab		.63a 90.89 100	75.63a 90.89 100.00 100.00 HS	2	0.00c 1.33c	21.33c	46.67b	62.67ab	69.33ab 8	80.00a 81.	81.33a 100.0	100.00 100.00 100.00 HS	100.00 HS	16	80.80/133.83
CDU-17	0.00d 0.71d 11.72cd		21.19bc 35.17ab		.48a 60.72 71.	45.48a 60.72 71.56 100.00 MS	12	0.00d 0.00d	34.67c	56.00bc	62.67ab	70.67ab	77.33ab 78.67a		100.00 100.00 100.00 HS	100.00 HS	17	41.73/148.00
CDU-18	0.00c 0.44c 0.88bc		2.06bc 2.85b		5.23a 14.78 29.23	23 72.49 HR	31	0.00d 1.33cd	10.67c	52.00b	89.33a 9	96.00a 9	96.00a 96.	96.00a 100.0	100.00 100.00 100.00 HS	100.00 HS	4	3.91/58.33
CDU-19	0.07d 0.77d 5.74cd		9.86bc 14.10ab		.25a 30.47 48.	19.25a 30.47 48.06 100.00 R	25	6.67d 10.67d	10.67cd 20.00cd 26.67bc		42.67ab 4	48.00a ²	48.00a 53.	53.33a 65.33	73.33	100.00 MS	30	16.76/105.67
CDU-20	0.00c 0.77c 9.34bc		15.71abc 21.98ab		.23a 48.53 62.	29.23a 48.53 62.53 100.00 MR	. 15	2.67f 5.33f	25.33e 49.33d		66.67c 7	78.67b 9	93.33a 94.	94.67a 100.0	100.00 100.00 100.00 HS	100.00 HS	5	33.03/155.17
CDU-21	0.00c 0.18c 4.40bc		9.10abc 15.19ab		21.51a 39.24 56.44	.44 100.00 MR	19	2.67f 2.67f	13.33ef	13.33ef 28.00de	44.00cd 2	52.00bc	66.67ab 76.00a		100.00 100.00 100.00 HS	100.00 HS	20	23.55/100.17
CDU-22	0.09d 0.73d 1.74cd	-cd 3.94bc	oc 6.80ab		8.41a 20.39 34.17	.17 80.27 R	27	0.00c 1.33c	42.67b	62.67b	88.00a 9	90.67a 9	93.33a 94.	94.67a 100.0	100.00 100.00 100.00 HS	100.00 HS	5	8.12/185.50
CDU-23	0.00b 0.00b 0.00b	b 0.23b	o 0.50ab		1.05a 10.32 19.32	.32 53.67 HR	37	0.00a 0.00a	0.00a	0.00a	1.33a 2	2.67a 2	2.67a 2.67a	7a 10.67	30.00	100.00 HR	37	0.44/2.67
CDU-24	0.00c 2.79c 10.41bc		22.06abc 27.99ab		32.44a 68.47 79.47	.47 100.00 MS	11	0.00d 9.33d	46.67c	57.33bc	65.33bc 7	74.67ab 8	89.33a 90.	90.67a 100.0	100.00 100.00 100.00 HS	100.00 HS	13	44.76/173.33

Table 2: Evaluation of 37 quinoa resources for germination trial

Resources	-	Whole spik	e germina.	Whole spike germination rate (%)		Resistance Index	Index				Grain ger.	Grain germination rate (%)	ite (%)			Dormé	Dormancy Germination	
	8 h 16 h 24 h	32 h	40 h	48 h 72 h 96 h 120 h	h 96 h	120 h	rankıng	4 h 8 h	12 h	16 h	20 h	24 h	28 h	32 h 4	44 h 56 h	68 h grade	rate ranking	germination index (%)
CDU-25	0.00b 0.37b 6.88ab 13.21ab 19.99ab	13.21ab	, 19.99ab	28.47a 46.59 62.19 100.00 MR	59 62.19	100.00 MR	14	0.00d 1.33d		2.67cd 16.00bc 26.67b	26.67b	41.33a	44.00a	54.67a 7	77.33 87.33	100.00 MS	28	37.86/59.83
CDU-26	2.04c 6.25c 8.16bc 14.28ab 15.24ab	14.28ab	15.24ab		17.04a 38.54 58.54 100.00 MR	100.00 MR	16	0.00e 20.00d	00d 64.00c	c 73.33b	89.33a	93.33a	96.00a	97.33a 1	96.00a 97.33a 100.00 100.00 100.00 HS	100.00 HS	2	29.27/227.17
CDU-27	0.00c 5.30bc 17.83abc 27.52ab 34.03a	bc 27.52ab	, 34.03a	39.53a 59.5	39.53a 59.33 73.47 100.00 MS	100.00 MS	6	2.67e 21.33d	33d 40.00c	c 69.33b	69.33b	90.67a	93.33a	94.67a 1	94.67a 100.00 100.00 100.00 HS	100.00 HS	5	48.52/198.50
CDU-28	0.44d 3.36cd 5.13bcd 11.58abc 13.66ab	d 11.58ab	ic 13.66ab		20.18a 38.16 59.81 100.00 MR	100.00 MR	20	5.33c 13.33c	33c 44.00b	b 80.00a	89.33a	89.33a	94.67a	94.67a 1	94.67a 94.67a 100.00 100.00 100.00 HS	100.00 HS	S	23.54/213.67
CDU-29	0.00c 4.38c 24.15bc 36.08abc 44.84ab	c 36.08ab	ic 44.84ab		55.85a 74.57 84.57 100.00 MS	100.00 MS	7	10.67f 16.00f		s 58.67cd	72.00bc	86.67ab	94.67a	94.67a 1	45.33e 58.67cd 72.00bc 86.67ab 94.67a 94.67a 100.00 100.00 100.00 HS	100.00 HS	S	55.16/199.33
CDU-30	0.00d 4.72cd 17.58c 33.31b 47.02a	33.31b	47.02a	56.91a 77.4	56.91a 77.43 87.56 100.00 MS	100.00 MS	9	0.00d 1.33d	d 9.33cc	1 18.67bc	d 25.33bco	l 33.33abc	54.67ab	45.33a 6	9.33cd 18.67bcd 25.33bcd 33.33abc 54.67ab 45.33a 61.33 72.00 100.00 MR	100.00 MR	32	56.30/64.33
CDU-31	0.00d 5.66d 31.07c 40.51c 60.65b	40.51c	60.65b	76.43a 96.7	76.43a 96.79 100.00 100.00 HS	SH 00.00	1	1.33e 12.0	1.33e 12.00e 38.67d 76.00c	d 76.00c		92.00ab	96.00a	96.00a 1	84.00bc 92.00ab 96.00a 96.00a 100.00 100.00 100.00 HS	100.00 HS	б	81.50/200.83
CDU-32	0.00d 2.08d 9.38cd 12.81bc 16.80ab	12.81bc	; 16.80ab	23.73a 44.33 58.33 100.00 MR	33 58.33	100.00 MR	17	1.33e 5.33e		17.33d 61.33c	76.00b	86.67a	90.67a	92.00a 1	90.67a 92.00a 100.00 100.00 100.00 HS	100.00 HS	11	24.64/162.00
CDU-33	0.00d 0.03d 1.31cd		6.32b	3.61bc 6.32b 13.78a 23.79	32.65 75.68	75.68 R	28	0.00d 1.33d		10.67cd 20.00bc	30.67b	49.33a	56.00a	57.33a 7	56.00a 57.33a 70.67 81.33 100.00 MS	100.00 MS	25	7.32/76.67
CDU-34	2.22e 11.28d 16.40d 30.52c 43.62b 56.78a 83.32	30.52c	43.62b	56.78a 83.32	100.00	100.00 100.00 HS	5	2.67d 22.67c		69.33b 97.33a	100.00a	100.00a	100.00a	100.00a 1	100.00a 100.00a 100.00a 100.00 100.00 100.00 HS	100.00 HS	1	62.14/260.33
CDU-35	0.00c 3.68c 21.56bc 36.50ab 47.18ab 56.99a 89.19	c 36.50ab	, 47.18ab	56.99a 89.19	100.00	100.00 100.00 HS	3	0.00e 6.67e		14.67de 36.00cd	52.00bc	60.00ab	74.67a	74.67a 1	60.00ab 74.67a 74.67a 100.00 100.00 100.00 HS	100.00 HS	18	68.64/115.83
CDU-36	0.00c 0.25c 0.36c 0.94bc 1.83b 3.44a 14.74	0.94bc	1.83b	3.44a 14.74	28.47	28.47 70.56 HR	34	1.33e 1.33e	se 8.00e	21.33d	32.00cd		46.67ab	57.33a 7	40.00bc 46.67ab 57.33a 70.00 79.33 100.00 MS	100.00 MS	25	1.88/71.67
CDU-37	0.00c 1.73c 11.69bc 28.54ab 33.65a 42.11a 55.37	c 28.54ab) 33.65a	42.11a 55.37	65.87	65.87 100.00 MS	10	0.00e 0.00e		13.33d 53.33c	68.00b	76.00b	89.33a	92.00a 1	89.33a 92.00a 100.00 100.00 100.00 HS	100.00 HS	П	46.56/139.67

lote: Different lowercase letters indicate significant differences in germination rates of the same variety (grain and whole spike) at different germination stages ($p < 0.05$). Among them, the whole spike germination resistance test	was conducted within 8-48 h, the whole spike germination index, germination index ranking and resistance grade classification were calculated in addition. The whole spike continuous germination test was carried out at 72,	6 and 120 h. The grain germination resistance test was conducted within 4-32 h, the grain germination index, germination index ranking and dormancy grade classification were calculated in addition. The grain germination test	was carried out at 44, 56 and 68 h. Only the germination resistance experimental data were significantly labeled.	
Note: Different lowercase letters indicate signi	was conducted within 8–48 h, the whole spike	96 and 120 h. The grain germination resistance	was carried out at 44, 56 and 68 h. Only the	

			Pi	rojects		
Indicators		Grain germina	tion rate	Wł	nole spike germ	nination index
	Number	Range of variation (%)	Coefficient of variation (%)	Number	Range of variation (%)	Coefficient of variation (%)
HR	1	2.67	100.00	9	0.44~4.66	61.86
R	1	16.00	100.00	7	7.32–19.81	37.86
MR	5	32.00-48.00	14.82	9	23.32-39.93	22.99
MS	10	53.33-65.33	7.60	7	41.73-56.30	11.30
HS	20	72.00-100.00	9.64	5	62.14-81.50	13.18
Mean ± standard deviation (%)	70.45 ± 2	24.50		29.27 ±	24.04	
Coefficient of variation (%)	34.78			82.13		

Table 3: Analysis of the diversity of PHS resistance in 37 quinoa resources

3.4 Analysis of High-Quality Agronomic Traits of Quinoa Resources

Fig. 1 showed that the phenotypic trait characteristics of quinoa seeds showed a roughly normal distribution trend. There was an R^2 greater than 0.9 in grain length (Fig. 1A), grain width (Fig. 1B), grain area (Fig. 1C), and thousand grain weight (Fig. 1D), suggesting that the phenotypic traits were well modeled and the overall variation was small.

Agronomic statistics and analysis of quinoa grains were presented in Tables 4 and 5. According to the 37 quinoa varieties tested, variety CDU-18 had the maximum weight of 3.70 grams, while variety CDU-5 had the smallest weight of 1.60 grams, indicating there was a difference of 2.10 g between the two varieties. The CDU-2 variety had a maximum grain length of 0.25 cm and a maximum grain area of 0.05 cm². There was a coefficient of variation of 19.79% for thousand grain weight, followed by a coefficient of variation of 18.35% for grain area. The results showed a high degree of dispersion in phenotypic data, indicating a large difference in yield between varieties. There was relatively little variation in grain length and width, both less than 10%, indicating that phenotypic traits were relatively stable in quinoa.

Based on the results of Table 5, it was apparent that agronomic traits were more closely related to quinoa yields. For each variety, thousand grain weight, grain length, grain area, and grain width correlated with yield, in descending order. According to the classification principle, the higher the correlation degree, the closer the relationship between the traits and the yield. Among phenotypic traits, thousand grain weight was a decisive factor for quinoa yield, while seed length and seed area had a weaker effect.

3.5 Correlation Analyses of Each Index of Quinoa Spike Germination with Agronomic Traits

The correlation analysis results between the PHS indicators and agronomic traits of quinoa are shown in the Table 6. The indicators had degrees of correlation. Especially, each of the evaluation indices of spike germination was highly significant and positive correlated, only the whole spike germination rate and seed germination rate (r = 0.399) were significantly and positively correlated. All indicators of agronomic traits were highly significantly positively correlated. The correlation between different types of indicators was also evident. Grain germination rate was highly significantly positively correlated with thousand

grain weight (r = 0.436), and grain germination index was significantly positively correlated with thousand grain weight (r = 0.391), grain area (r = 0.340), and grain width (r = 0.349). Seed area, seed length, and seed width negatively correlated with whole spike germination rate, correlations between the remaining indicators were not significant.

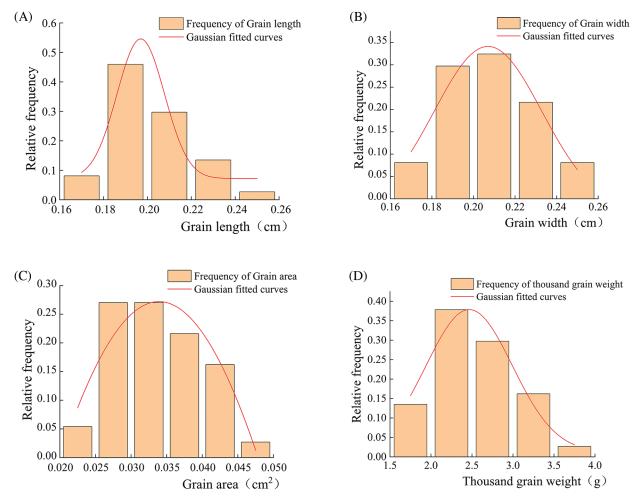


Figure 1: Frequency distribution of agronomic traits in the grains of 37 quinoa resources. (A) Grain length; (B) Grain width; (C) Grain area; (D) Thousand grain weight

Table 4:	Mean	values	of basic	agronomic	traits for	or 37	quinoa	resources
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			Trait	
Species	Grain length (cm)	Grain width (cm)	Grain area (cm ²)	Thousand grain weight (g)
CDU-1	$0.2042 \pm 0.0119 defghijkl$	$0.2262\pm0.0119 bcdef$	$0.0370\pm0.0015 defghi$	$2.3510 \pm 0.0273 lm$
CDU-2	$0.2526 \pm 0.0110 a$	$0.2450 \pm 0.0198 ab$	$0.0478 \pm 0.0064a$	$3.0804 \pm 0.0474 def$
CDU-3	$0.1802 \pm 0.0118 lmno$	$0.1856\pm0.0096mnop$	$0.0264\pm0.0027 pqrs$	$2.0718 \pm 0.0535 p$
CDU-4	$0.1966 \pm 0.0089 efghijklm$	$0.2092 \pm 0.0139 fghijk$	$0.0322 \pm 0.0031 ghijklmnop$	$2.1905 \pm 0.0495o$
CDU-5	$0.1856 \pm 0.0103 klmno$	$0.1836\pm0.0084nopq$	$0.0264\pm0.0019 pqrs$	$1.6030 \pm 0.0314 t$

(Continued)

Table 4 (continued)

		,	Trait	
Species	Grain length (cm)	Grain width (cm)	Grain area (cm ²)	Thousand grain weight (g)
CDU-6	$0.1980 \pm 0.0110 efghijklm$	$0.1996 \pm 0.0104 ijklmn$	$0.0306 \pm 0.0031 ijklmnopq$	$2.0669 \pm 0.0576 p$
CDU-7	$0.1980 \pm 0.0129 efghijklm$	$0.1984 \pm 0.0062 ijklmno$	$0.0312 \pm 0.0028 hijklmnopq$	$2.2209 \pm 0.0718 no$
CDU-8	$0.1914 \pm 0.0524 hijklmn$	$0.2144 \pm 0.0114 efghi$	$0.0372\pm0.0052 defgh$	$2.8483 \pm 0.0609 g$
CDU-9	$0.2180 \pm 0.0086 bcdef$	$0.2376 \pm 0.0152 abcd$	$0.0406 \pm 0.0036 bcde$	$2.8046 \pm 0.0474 g$
CDU-10	$0.1916 \pm 0.0101 hijklmn$	$0.1918 \pm 0.0117 jklmnop$	$0.0288 \pm 0.0035 mnop qrs$	$2.3518 \pm 0.0717 lmn$
CDU-11	$0.2054 \pm 0.0087 defghijk$	$0.2032 \pm 0.0108 ghijklmn$	$0.0330 \pm 0.0033 ghijklmno$	$2.8199 \pm 0.0783 g$
CDU-12	$0.2156 \pm 0.0166 bcdefgh$	$0.2070 \pm 0.0053 fghijkl$	0.0354 ± 0.0020 efghijkl	$2.6395 \pm 0.0462 hi$
CDU-13	$0.1650 \pm 0.0038o$	$0.1650 \pm 0.0054 q$	$0.0226 \pm 0.0005 s$	$1.6921 \pm 0.0448 st$
CDU-14	$0.1788 \pm 0.0124 mno$	$0.1882 \pm 0.0089 klmnop$	$0.0276 \pm 0.0031 nopqrs$	$1.8869 \pm 0.0617 r$
CDU-15	0.2056 ± 0.0064 defghijk	$0.2070 \pm 0.0053 fghijkl$	0.0342 ± 0.0012 efghijklm	$2.4525 \pm 0.0773 jk$
CDU-16	$0.1714 \pm 0.0184 no$	$0.1738 \pm 0.0066 pq$	$0.0240 \pm 0.0028 rs$	$2.5260 \pm 0.0930 jk$
CDU-17	$0.2196 \pm 0.0087 bcdef$	$0.2072\pm0.0157 fghijkl$	$0.0356 \pm 0.0030 efghijk$	$3.1470 \pm 0.0465 de$
CDU-18	$0.2156 \pm 0.0139 bcdefgh$	$0.2362 \pm 0.0178 abcd$	$0.0404 \pm 0.0036 bcde$	$3.7010 \pm 0.0744 a$
CDU-19	$0.2270 \pm 0.0061 bcd$	$0.2488 \pm 0.0152a$	$0.0436 \pm 0.0017 abc$	$2.5008 \pm 0.0773 j$
CDU-20	$0.2206 \pm 0.0073 bcde$	$0.2174 \pm 0.0205 defghi$	0.0364 ± 0.0021 defghij	$3.4470 \pm 0.0730 b$
CDU-21	$0.1992 \pm 0.0204 efghijklm$	$0.1866 \pm 0.0147 lmnop$	$0.0290 \pm 0.0036 lmnopqrs$	$2.7010 \pm 0.0797 h$
CDU-22	$0.2104 \pm 0.0062 defghij$	$0.2236 \pm 0.0107 cdefg$	$0.0380 \pm 0.0024 bcdefg$	$2.7110 \pm 0.0902 h$
CDU-23	$0.1828\pm0.0128klmno$	$0.1984 \pm 0.0123 ijklmno$	$0.0272 \pm 0.0040 nopqrs$	$2.0701 \pm 0.0659 pq$
CDU-24	$0.1928 \pm 0.0119 ghijklmn$	$0.2148 \pm 0.0160 efghi$	$0.0318 \pm 0.0015 ghijklmnop$	$2.3820 \pm 0.0647 hl$
CDU-25	$0.1864 \pm 0.0048 jklmno$	$0.2020\pm0.0083 hijklmn$	$0.0296 \pm 0.0014 klmnopqr$	$2.3039 \pm 0.0814 lm$
CDU-26	$0.2346\pm0.0058abc$	$0.2414 \pm 0.0115 abc$	$0.0442 \pm 0.0023 ab$	$3.3968 \pm 0.1103 c$
CDU-27	$0.2386 \pm 0.0250 ab$	$0.2300 \pm 0.0210 abcde$	$0.0422 \pm 0.0055 abcd$	$2.9530 \pm 0.0405 f$
CDU-28	$0.1978 \pm 0.0074 efghijklm$	$0.1970 \pm 0.0091 ijklmno$	$0.0300 \pm 0.0015 jklmnopqr$	$3.0668 \pm 0.0868 ef$
CDU-29	$0.2168 \pm 0.0159 bcdefg$	$0.2224 \pm 0.0058 cdefgh$	$0.0378 \pm 0.0035 cdefg$	$2.8016 \pm 0.0403 g$
CDU-30	$0.2028 \pm 0.0069 defghijklm$	$0.2110 \pm 0.0107 efghij$	0.0344 ± 0.0015 efghijklm	$2.3140 \pm 0.0858 mn$
CDU-31	$0.2208 \pm 0.0128 bcde$	$0.2312 \pm 0.0115 abcde$	$0.0398 \pm 0.0040 bcdef$	$3.1839 \pm 0.0679 d$
CDU-32	0.1916 ± 0.0124 hijklmne	$0.1894 \pm 0.0189 klmnop$	$0.0274 \pm 0.0027 nopqrs$	$2.5683 \pm 0.0755 ij$
CDU-33	0.1966 ± 0.0081 efghijklm	$0.2062 \pm 0.0142 fghijklm$	$0.0306 \pm 0.0030 ijklmnopq$	$1.9680 \pm 0.0474 qr$
CDU-34	$0.2142 \pm 0.0074 cdefghi$	$0.2364 \pm 0.0123 abcd$	$0.0400 \pm 0.0021 bcde$	$2.0534 \pm 0.0476 pq$
CDU-35	$0.1954 \pm 0.0101 fghijklmn$	$0.2158 \pm 0.0108 efghi$	$0.0334 \pm 0.0021 fghijklmn$	$2.3580 \pm 0.0499 lmn$
CDU-36	$0.1902 \pm 0.0097 ijklmn$	$0.1932 \pm 0.0052 jklmnop$	$0.0268 \pm 0.0007 opqrs$	$1.7618 \pm 0.0533s$
CDU-37	$0.1802 \pm 0.0115 lmno$	$0.1784 \pm 0.0099 opq$	$0.0250 \pm 0.0017 qrs$	$2.3094 \pm 0.0603 mn$

Note: Different lowercase letters indicate significance of differences at the 0.05 level.

3.6 Cluster Analyses of PHS Indicators and Agronomic Traits of Quinoa Material

Results of the spike germination index by cluster analysis were shown in Fig. 2A. It was possible to divide the 37 quinoa resources into three major categories when the squared Euclidean distance was 9.66. First-class quinoa materials included CDU-2 and CDU-23, accounting for 5.41% of the test materials. The average spike and seed germination rate were 5.13% and 9.34%, and the average spike and seed germination index were 0.22 and 0.05. Based on the spike germination index, it was evident that a class of materials are highly resistant to spike germination. A total of 19 materials were included in the second

category, representing 51.35 percent of the materials tested. According to the results, pre-harvest germination rate, grain germination rate, pre-harvest germination index, and grain germination index of these materials were 39.02%, 89.75%, 0.43% and 1.70%, respectively. All the relevant pre-harvest germination indices were high, belonging to the materials with low pre-harvest germination resistance, indicating that they were easy to sprout. The third category contained 16 materials that accounted for 43.24% of all test materials. It was 19.72%, 55.17%, 0.16% and 0.74% for the spike germination index, seed germination index, respectively, and there was some resistance to PHS at the middle level.

Agronomic traits	Maximum value	Minimum value	Range	Mean ± standard deviation (%)	Coefficient of variation (%)	Relevance	Ranking
Thousand grain weight	3.7010	1.6030	2.0980	2.5216 ± 0.4491	19.7912	0.911	1
Grain length	0.2526	0.1651	0.0880	0.2025 ± 0.0188	9.2818	0.685	2
Grain area	0.0478	0.0226	0.0250	0.0335 ± 0.0061	18.3451	0.674	3
Grain width	0.2488	0.1654	0.0830	0.2087 ± 0.0206	9.8904	0.638	4

 Table 5: Agronomic traits and grey correlation analysis of quinoa resources

Table 6:	Correlation	analysis of	each indicator	of quinoa PHS	with agronomic traits
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Indicators	Whole spike germination rate	Whole spike germination index	Grains germination rate	Grains germination index	Grain length	Grain width	Grain area	Thousand grain weight
Whole spike germination rate	1							
Whole spike germination index	0.955**	1						
Grains germination rate	0.399*	0507**	1					
Grains germination index	0.431**	0.573**	0.857**	1				
Grain length	-0.054	0.009	0.150	0.298	1			
Grain width	-0.079	0.027	0.194	0.349*	0.859**	1		
Grain area	-0.035	0.037	0.185	0.340*	0.963**	0.963**	1	
Thousand grain weight	0.110	0.170	0.436**	0.391*	0.564**	0.564**	0.645**	1

Note: "*" indicates a significant level of correlation (p < 0.05), "**" indicates a highly significant correlation (p < 0.01).

Results of the agronomic traits by cluster analysis were shown in Fig. 2B. The 37 quinoa resources could be divided into three categories based on the squared Euclidean distance of 9.66. Three materials were in the first category, representing 8.11% of the test material, namely CDU-18, CDU-26, and CDU-19. There was an

average grain length of 0.22, a grain width of 0.23, a grain area of 0.04 and a thousand grain weight of 3.52 g. It was a quinoa resource with yield and quality potential, as its agronomic traits were higher than the other two categories. A total of 12 materials were tested in the second category, representing 32.43 percent of all materials tested. In terms of grain length, grain width, grain area, and thousand grain weight, the average grain was 0.22, 0.228, 0.04, and 2.85, which was an excellent quinoa resource for transformation, all agronomic indexes place it slightly below the first category, but significantly higher than the third category. The third category contains another 22 materials, accounting for 59.46%. Approximately 0.18, 0.19, 0.03 and 2.11 grains were averaged for length, width, grain area and thousand grains.

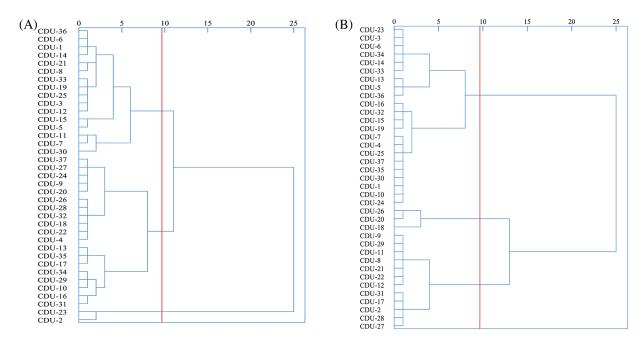


Figure 2: Cluster analysis of 37 quinoa resources. (A) Cluster analysis of PHS resistance indicators; (B) Cluster analysis of agronomic traits

4 Discussion

PHS in grains is an irreversible and extremely damaging natural disaster, the imbalance between grain dormancy and germination is the main reason for cause of PHS [13,29,30], which can lead to a decrease in grain yields and quality. Agronomic traits of germplasm resources that can be used for estimating grain yield and quality. Grain yield is determined by thousand grain weight, while grain quality is determined by seed length, seed width, and seed area [28,31]. One of the most effective ways to reduce PHS and improve nutrition of grain is by selecting and breeding varieties that have excellent agronomic attributes and are resistant to PHS.

This study selected quinoa varieties with PHS resistance based on seed germination and whole spike germination test referring to the experimental methods for identifying PHS resistance in wheat, rice and barley [23,32–34] adjusted for quinoa germination characteristics. By analyzing thousand grain weight, seed length, seed width, and seed area, excellent agronomic traits were selected. Dormancy could explain seed PHS resistance, which is closely related to seed dormancy characteristics. The strength of seed dormancy characteristics could be determined by grain germination rate, grain germination index, whole spike germination rate and whole spike germination index. According to the seed germination rate and the whole spike germination index, PHS resistance of quinoa varieties were divided into five categories:

high resistance (HR), medium resistance (MR), resistance (R), medium sensitivity (MS), and high sensitivity (HS). After analysis of variation coefficient, seed germination rate, whole spike germination rate, seed germination rate and whole spike germination index, it was found that the seed germination rate of 37 quinoa varieties was much higher than the germination rate of the whole spike, and the grain germination index was higher than the germination index of the whole spike, indicating that significant differences in PHS resistance among different varieties. Correlation analysis revealed that PHS resistance indexes of all varieties were significantly and positively correlated.

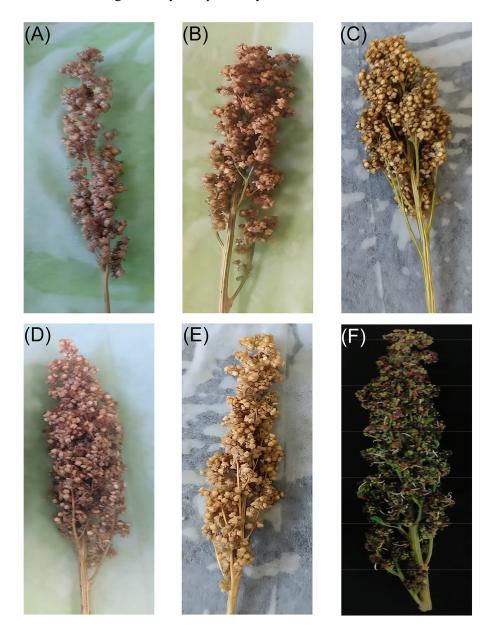


Figure 3: The picture of whole spike germination in 48 h. The excellent agronomic traits and PHS resistance of quinoa varieties were as follows: (A) represents CDU-2 with germination rate of 9.21%, (B) represents CDU-11 with germination rate of 36.85%, (C) represents CDU-12 with germination rate of 28.28%, (D) represents CDU-21 with germination rate of 21.51%, (E) represents CDU-8 with germination rate of 24.15%. (F) represents CDU-31 as the most sensitive variety to PHS, with germination rate of 76.43%

Furthermore, during the experimental process, it was found that the middle and bottom of the spike were the main germination sites. A first possibility is that there are substances in the peripheral perianth of grains that inhibit or promote grain germination, thus, the variety acquired PHS resistance, which was consistent with previous studies [35]. Secondly, water in the tissue at the top of the spike may flow downward due to gravity, the spike bottom stayed wet for a long time and provided the conditions for grain germination. The third reason might be that the seeds at the top of the spike were exposed to more sunlight and had lower water content, which dormancy of seed was enhanced and obtained stronger PHS resistance. The last reason might be that a-amylase activity and endogenous hormone content in the embryo of the seed dormancy [12,36–38]. This study of agronomic traits has been shown to follow a normal distribution. According to the coefficient of variation, there was a large difference between thousand grain weight and seed area. It was found that the traits were highly significantly correlated in the correlation analysis. Gray correlation analysis revealed that thousand grain weight, grain length, and grain area were the three most important traits affecting yield. Variations in genetic variation may lead to differences in yield and quality under the same treatment. Cluster analysis was used to classify 37 quinoa resources based on agronomic traits and PHS resistance at a squared Euclidean distance of 9.66. The germplasm resources with similar PHS resistance and genetic proximity were clustered into one category, for cultivation, PHS resistant varieties with excellent agronomic qualities (CDU-2) were selected to alleviate damage caused by quinoa PHS and to increase yield and quality, and promote the healthy development of quinoa industry.

5 Conclusion

In this study, 37 quinoa resources were tested for PHS resistance in 48 h, excellent agronomic traits and were compared. Five quinoa materials, CDU-2, CDU-11, CDU-12, CDU-21 and CDU-8, were selected from the comprehensive multiple comparison analysis, and all had better PHS resistance and excellent agronomic traits. The CDU-31 was selected as the most sensitive material to PHS. The photos of the 6 quinoa varieties were shown in Fig. 3. There was one material, CDU-2, which had the best PHS resistance and thousand grain weight, length, width, and area of seed. Besides screening high quality and PHS resistant quinoa varieties suitable for low altitude cultivation in Chengdu plain, this study also provides some theoretical support for the creation of subsequent quinoa germplasm resistant to PHS, which has important implications in regards to cultivation extension and production guidance.

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Availability of Data and Materials: The datasets used and/or analyzed during the current study are available from the author and/or corresponding author on reasonable request.

Ethics Approval: Not applicable.

Conflicts of Interest: The authors declare that they have no conflicts of interest to report regarding the present study.

References

- 1. Zurita-Silva, A., Fuentes, F., Zamora, P., Jacobsen, S. E., Schwember, A. R. (2014). Breeding quinoa (*Chenopodium quinoa* Willd.): Potential and perspectives. *Molecular Breeding*, 34(1), 13–30.
- Bazile, D., Pulvento, C., Verniau, A., Al-Nusairi, M. S., Ba, D. et al. (2016). Worldwide evaluations of quinoa: Preliminary results from post international year of quinoa FAO projects in nine countries. *Frontiers in Plant Science*, 7, 850–868.
- Reguera, M., Conesa, C. M., Gil-Gomez, A., Haros, C. M., Perez-Casas, M. A. et al. (2018). The impact of different agroecological conditions on the nutritional composition of quinoa seeds. *PeerJ*, 6, e4442–e4462.
- 4. Vega-Galvez, A., Miranda, M., Vergara, J., Uribe, E., Puente, L. et al. (2010). Nutrition facts and functional potential of quinoa (*Chenopodium quinoa* willd.), an ancient Andean grain: A review. *Journal of the Science of Food and Agriculture*, 90(15), 2541–2547.
- 5. Navruz-Varli, S., Sanlier, N. (2016). Nutritional and health benefits of quinoa (*Chenopodium quinoa* Willd.). *Journal of Cereal Science*, 69, 371–376.
- Angeli, V., Miguel Silva, P., Crispim Massuela, D., Khan, M. W., Hamar, A. et al. (2020). Quinoa (*Chenopodium quinoa* Willd.): An overview of the potentials of the "golden grain" and socio-economic and environmental aspects of its cultivation and marketization. *Foods*, 9(2), 216–247.
- 7. Hussain, M. I., Farooq, M., Syed, Q. A., Ishaq, A., Al-Ghamdi, A. A. et al. (2021). Botany, nutritional value, phytochemical composition and biological activities of quinoa. *Plants*, *10(11)*, 2258–2276.
- 8. Sohn, S. I., Pandian, S., Kumar, T. S., Zoclanclounon, Y. A. B., Muthuramalingam, P. et al. (2021). Seed dormancy and pre-harvest sprouting in rice–An updated overview. *International Journal of Molecular Sciences*, 22(21), 11804–11826.
- 9. Bhargava, A., Shukla, S., Ohri, D. (2006). Chenopodium quinoa—An Indian perspective. *Industrial Crops and Products*, 23(1), 73–87.
- Pulvento, C., Riccardi, M., Lavini, A., D'Andria, R., Iafelice, G. et al. (2010). Field trial evaluation of two chenopodium quinoa genotypes grown under rain-fed conditions in a typical mediterranean environment in South Italy. *Journal of Agronomy and Crop Science*, 196(6), 407–411.
- 11. Zhang, X., Wang, S. (2020). Long-term trend of precipitation days for Southeast Tibetan Plateau China. *Journal of Agricultural Meteorology*, *76(2)*, 111–118.
- 12. Nonogaki, M., Sall, K., Nambara, E., Nonogaki, H. (2014). Amplification of ABA biosynthesis and signaling through a positive feedback mechanism in seeds. *The Plant Journal*, 78(3), 527–539.
- 13. Tai, L., Wang, H. J., Xu, X. J., Sun, W. H., Ju, L. et al. (2021). Pre-harvest sprouting in cereals: Genetic and biochemical mechanisms. *Journal of Experimental Botany*, 72(8), 2857–2876.
- 14. Simsek, S., Ohm, J. B., Lu, H., Rugg, M., Berzonsky, W. et al. (2014). Effect of pre-harvest sprouting on physicochemical properties of starch in wheat. *Foods*, *3*(2), 194–207.
- 15. Nonogaki, H., Barrero, J. M., Li, C. (2018). Editorial: Seed dormancy, germination, and pre-harvest sprouting. *Frontiers in Plant Science*, *9*, 1783–1786.
- 16. Li, C., Ni, P., Francki, M., Hunter, A., Zhang, Y. et al. (2004). Genes controlling seed dormancy and pre-harvest sprouting in a rice-wheat-barley comparison. *Funct Integr Genomics*, 4(2), 84–93.
- 17. Martinez, S. A., Godoy, J., Huang, M., Zhang, Z., Carter, A. H. et al. (2018). Genome-wide association mapping for tolerance to preharvest sprouting and low falling numbers in wheat. *Frontiers in Plant Science*, *9*, 141–157.
- 18. Upadhyay, M. P., Morris, C. F., Paulsen, G. M. (1988). Characterization of preharvest sprouting resistance in clark's cream white winter wheat. *Euphytica*, 38(1), 85–92.
- 19. George, D. W. (1967). High temperature seed dormancy in wheat (*Triticum aestivum* L.). Crop Science, 7(3), 249–253.
- Daniel, I. O., Adeboye, K. A., Oduwaye, O. O., Porbeni, J. (2012). Digital seed morpho-metric characterization of tropical maize inbred lines for cultivar discrimination. *International Journal of Plant Breeding and Genetics*, 6(4), 245–251.

- 21. Evers, A. D., Cox, R. I., Shaheedullah, M. Z., Withey, R. P. (1990). Predicting milling extraction rate by image analysis of wheat grains. *Aspects of Applied Biology*, 25, 417–426.
- 22. Gu, Y., Qian, X., Sun, B., Ma, S., Tian, X. et al. (2022). Nutritional composition and physicochemical properties of oat flour sieving fractions with different particle size. *LWT*, *154*, 112757–112767.
- 23. Hagemann, M. G., Ciha, A. J. (1984). Evaluation of methods used in testing winter wheat susceptibility to preharvest sprouting. *Crop Science*, 24(2), 249–254.
- 24. Ching, T. M., Foote, W. H. (1961). Post-harvest dormancy in wheat varieties. Agronomy Journal, 53(3), 183-186.
- 25. Mares, D. J. (1984). Temperature dependence of germinability of wheat (*Triticum aestivum* L.) grain in relation to pre-harvest sprouting. *Australian Journal of Agricultural Research*, 35(2), 115–128.
- 26. Reddy, L. V., Metzger, R. J., Ching, T. M. (1985). Effect of temperature on seed dormancy of wheat. *Crop Science*, 25(3), 455–458.
- 27. Yang, Y., Zhao, X. L., Xia, L. Q., Chen, X. M., Xia, X. C. et al. (2007). Development and validation of a *Viviparous-1* STS marker for pre-harvest sprouting tolerance in Chinese wheats. *Theoretical and Applied Genetics*, 115(7), 971–980.
- 28. Gupta, P. K., Rustgi, S., Kumar, N. (2006). Genetic and molecular basis of grain size and grain number and its relevance to grain productivity in higher plants. *Genome*, 49(6), 565–571.
- 29. Penfield, S. (2017). Seed dormancy and germination. Current Biology, 27(17), R874-R878.
- Ceccato, D. V., Daniel Bertero, H., Batlla, D. (2011). Environmental control of dormancy in quinoa (*Chenopodium quinoa*) seeds: Two potential genetic resources for pre-harvest sprouting tolerance. *Seed Science Research*, 21(2), 133–141.
- 31. Moles, A. T., Ackerly, D. D., Webb, C. O., Tweddle, J. C., Dickie, J. B. et al. (2005). Factors that shape seed mass evolution. *Proceedings of the National Academy of Sciences of the United States of America*, 102(30), 10540–10544.
- 32. Schwarz, P., Horsley, R., McNamara, H. (2018). Preharvest sprouting in the 2002 midwestern barley crop: Occurrence and assessment of methodology. *Journal of the American Society of Brewing Chemists*, 62(4), 147–154.
- 33. Shorter, S. C., Munro, C. A., Hodgkinson, J. (2005). Predicting pre-harvest sprouting susceptibility in New Zealand wheat cultivars. *Euphytica*, 143(3), 309–312.
- 34. Yanagisawa, A., Nishimura, T., Amano, Y., Torada, A., Shibata, S. (2005). Development of winter wheat with excellent resistance to pre-harvest sprouting and rain damage. *Euphytica*, 143(3), 313–318.
- 35. Liu, D. C., Lan, X. J., Wang, Z. R., Zheng, Y. L., Zhou, Y. H. et al. (1998). Evaluation of *Aegilops tauschii* Cosson for preharvest sprouting tolerance. *Genetic Resources and Crop Evolution*, 45(6), 495–498.
- 36. Suriyasak, C., Oyama, Y., Ishida, T., Mashiguchi, K., Yamaguchi, S. et al. (2020). Mechanism of delayed seed germination caused by high temperature during grain filling in rice (*Oryza sativa* L.). *Scientific Reports*, 10(1), 17378–17389.
- 37. Phan, P. D. T., Van Vu, B. (2021). Improving the pre-harvest sprouting resistance of rice cultivar IR36 using wild rice (*Oryza rufipogon*) W630. *Cereal Research Communications*, 50(1), 37–43.
- Lee, J. S., Chebotarov, D., McNally, K. L., Pede, V., Setiyono, T. D. et al. (2021). Novel sources of pre-harvest sprouting resistance for japonica rice improvement. *Plants*, 10(8), 1709–1722.