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Cd and Hg Mediated Oxidative Stress, Antioxidative Metabolism and Molecular Changes in Soybean (*Glycine max* L.)

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Received: 16 August 2022 Accepted: 16 December 2022

ABSTRACT

Cadmium (Cd) and Mercury (Hg) is among the heavy metals most hazardous for plant and human health. Known to induce oxidative stress in plants and disbalance equilibrium in the antioxidant defence system, these metals alter plant growth and cause damage at the cellular and molecular levels. Soybean is an important oilseed crop that is raised in soils often contaminated by Cd and Hg. The comparative studies on the deleterious effect of Cd and Hg and the defence system of antioxidants were not studied earlier in soybean plant. In this study, soybean plants were exposed to Cd (100 μ M CdCl₂) and Hg (100 μ M HgCl₂) and studied for physiological, biochemical and molecular responses. Both Cd and Hg treatment increased the magnitude of oxidative stress. Activities of antioxidant enzymes were significantly upregulated in response to Cd and Hg stress. Quantitative and qualitative assessment of isolated RNA showed significant differences in RNA under stress. Integrity values of RNA confirmed alterations. Transcript level of the Actin gene, involved in the morphogenesis of plants and also used as referenced gene in expression studies was analyzed using qRT-PCR just to check its stability and response under heavy metal stress. Results showed significant upregulation of the gene in the presence of Cd. It can be concluded that both Cd and Hg caused oxidative damage to plants, and adversely affected the quality of RNA. However, soybean tried to limit the adverse impacts of Cd and Hg stress by elevating the antioxidant system and upregulating Actin gene.

KEYWORDS

Soybean; heavy metal stress; actin gene; enzymatic antioxidant activity; qRT-PCR

1 Introduction

Soybean is among the important staple food sources all over the world. It is an excellent source of proteins, oil with less saturated fat, and fiber [1]. Hence, soybean serves as an important experimental crop in agricultural research areas [2]. Soy protein has very powerful effects of decreasing plasma cholesterol, cancer prevention, diabetes control, control of obesity, and defense against stomach and kidney disease [3,4]. In the process of water and mineral uptake, roots easily pick up uninvited ionized



toxic heavy metals [5] which have no function in plant metabolism except causing toxicity and reducing plant growth and development [6].

Cadmium (Cd) is one of the most potentially hazardous metal pollutants which caused serious health hazards once absorbed in the body, it is efficiently taken into the human body and accumulated for the entire life of humans [7]. When Cd is released into agricultural lands it affects numerous parameters of plant growth and physiology. Cd alters RNA expression, DNA methylation, and modifies histones [8]. Exposure to Cd, altered the expression pattern of various key genes involved in the regulatory mechanism of plants [9]. Cd affects gene expressions through its ability to change the activities of various pathways of signal transduction [10]. In plants, Cd also changes the levels of metabolic constituents involved in various pathways and ultimately generated reactive oxygen species (ROS) by inducing oxidative stress [11]. The generated ROS reacted with proteins, pigments, lipids and eventually degraded nucleic acids [12].

Mercury (Hg) is another serious environmental pollutant that is toxic to plants and animals and has become a major problem in recent times [13]. Hg is well known for affecting the antioxidant defence mechanism of plants, by modulating various enzymatic and non-enzymatic antioxidant activities [14].

Actin is a housekeeping gene that encodes a protein, which was involved in various significant subcellular processes in plants. Cell shape was determined by actin microfilament cytoskeleton which is concerned with cell division, cell elongation and plane localization [15]. Actin has vital significance for plant growth and development [16]. Actin is commonly used as a reference gene (control) in comparative expression analysis studies due to its stability [17], but several researchers reported that various abiotic factors affected its stability and gene expression which induced changes in the morphogenesis of plants and ultimately affects plant development [12,18]. For the first time in this study, we analyze the transcript level of the soybean plant actin gene under heavy metal (Cd and Hg) treatment to infer its stability and response to stress. This gene was also studied to investigate whether it will be used as a reference gene in future gene expression studies in soybean under stress conditions.

This study analyzes the deleterious effects of most hazardous heavy metals, Cd and Hg on important physiological and biochemical parameters of soybean including defence mechanisms represented by enzymatic and non-enzymatic antioxidants. The impact of Cd and Hg on the quality of RNA and transcript level of the actin gene was also considered to deduce its stability and its role in defence.

2 Materials and Methods

2.1 Plant Growth and Stress Treatment

Soybean (*Glycine max* L.) seeds of genotype JS-335 were officially procured from the ICAR-Indian Institute of Soybean Research (Indore, Madhya Pradesh). Healthy and physically uniform seeds were washed with detergent and sterilized in 1% (v/v) sodium hypochlorite for 8 min followed by 15 washings with double distilled water (DDW). Seeds were germinated in moist condition on paper beds for 5 days. The experiments were designed to form three sets of treatments viz. Control, Cadmium (Cd) and Mercury (Hg). Seedlings were transferred to the pots filled with 0.5 kg Soilrite (sterilized) for full plant growth and watered on alternate days. The first sets of pots were used as Control (only supplied with Hoagland nutrient solution) and experiments were conducted from triplicate pots.

When plants were 30 days old, stress was induced, in the second set of pots 100 μM CdCl_2 was supplied and the third set was supplied with 100 μM HgCl_2 , dissolved in Hoagland nutrient solution. This solution (Hoagland nutrient solution + stress) was supplied to pots once a day. The exposure to stressors continued for 10 days period. All pots were placed inside a growth chamber at 27°C and alternate light (16 h) and dark (8 h) periods.

2.2 Physiological and Biochemical Parameters

After 10 days of treatments, sampling of forty-days-old plants (40 DAS; vegetative stage) was done. Plants were uprooted and rinsed with distilled water. From all the treatments fully expanded mature fresh leaves were harvested and stored in liquid N₂ for performing further experiments on three biological replicates. For metal estimation, root and leaf samples were dried in a hot-air oven at 70°C for 3 days.

2.2.1 Estimation of Metal Accumulation

The root and leaf samples of all treatments were placed in an oven for complete drying at 70°C for 3 days. 100 mL mixture of HNO₃ and HClO₄ in a 5:2 ratio was used for digesting 0.5 g of plant material. The digested mixture was filtered with Whatman filter paper and made up to 50 mL. The estimation of metals (Cd and Hg) was performed by ICP-OES (Inductively Coupled Plasma-Optical Emission Spectrometry) in CIRBSc, Jamia Millia Islamia, New Delhi-110025, India.

2.2.2 Plant Growth

From fresh soybean plants, full height was measured of all sets of Control, Cd and Hg treated plants. All sets of plants were parted into roots, stems and leaves. Plant fresh weight (FW) and dry weight (DW) were measured in grams (g).

2.2.3 Photosynthetic Pigments

Chlorophyll pigments (chlorophyll a, chlorophyll b, total chlorophyll and total carotenoids) in leaves were calculated by Hiscox and Israelstam, 1979 [19]. 1 g of fresh leaves were powdered in liquid N₂ using a mortar and pestle. Added 10 mL of dimethyl sulfoxide (DMSO), and placed in a hot air oven at 65°C for 1 h. Optical density (OD) was recorded at 480, 645, 520, and 663 nm in a spectrophotometer. The concentrations of chlorophyll pigments were estimated by the Arnon formula, 1949 [20].

2.3 Estimation of Thiobarbituric Acid Reactive Substances (TBARS)

The magnitude of oxidative stress was estimated in terms of TBARS by the method of Heath and Packer [21]. 0.5 g of fresh leaves were ground in liquid nitrogen. 1% (w/v) trichloroacetic acid (TCA) was added to this and centrifugation was done for 5 min at 9660 xg. 1.0 mL of supernatant mixed with 4.0 mL of 0.5% (w/v) 2-thiobarbituric acid (TBA). Finally, this mixture was incubated at 95°C for 35 min, then placed on an ice bath and again centrifuged for 5 min. Supernatant OD was measured at 532 nm and 600 nm.

2.4 Histochemical Detection of Hydrogen Peroxide

For physical detection of H₂O₂ effect on leaves, fresh leaves were put in DAB solution (3, 3'-Diaminobenzidine, 1 mg/mL) and kept overnight. All the chlorophyll of leaves was removed by boiling for 10 min in ethanol. The reddish-brown color that appeared on leaves denoted the H₂O₂ content. It was visualized and photographed in white light illuminator [6].

2.4.1 Estimation of H₂O₂ Content

H₂O₂ content on leaves was assessed by Yu et al. method [22]. Leaves were ground and added with 3 mL of potassium phosphate buffer, spun at 9660 xg for 20 min. The supernatant was taken out and added with 0.5 mL of titanium chloride solution and incubated for 10 min at room temperature. Again spun for 10 min. Optical density was recorded at 410 nm for the estimation of H₂O₂ concentration.

2.5 Extraction and Analysis of Antioxidant Enzymes

0.5 g of leaf samples were mixed evenly with 5 mL of cooled phosphate buffer (100 mM, pH 7.4, 1 mM EDTA, 2.5 mM DL-dithiothreitol, 0.5% (v/v) Triton X-100 and 4% (w/v) insoluble polyvinylpyrrolidone) and centrifuged for 30 min at 9660 xg, the supernatant was taken out and used in enzymatic analysis. This Enzyme extraction was done at 4°C. All enzymatic assays were done at 25°C temperature.

2.5.1 Superoxide Dismutase Assay (SOD)

SOD activity was calculated by the process described by Dhindsa et al. [23]. It was estimated by its capacity to restrain photochemical reduction of nitro blue tetrazolium (NBT). In this assay 1.5 mL phosphate buffer (0.1 M) was mixed with 0.2 mL methionine and 0.1 mL extract of the enzyme with an equivalent amount of 1M Na_2CO_3 , 2.25 mM NBT solution, 3 mM EDTA, 60 mM riboflavin and 10 mL DDW (Double Distilled Water). All the mixtures were incubated in dark for 10 min and then absorbance was taken at 560 nm. SOD was expressed in units of EU (Enzyme Unit) $\text{min}^{-1} \text{mg}^{-1}$ protein.

2.5.2 Ascorbate Peroxidase Assay (APX)

Ascorbate peroxidase (APX) assay was done by taking supernatant absorbance at 290 nm according to Qureshi et al. [24]. A reaction mixture of 100 mM potassium phosphate buffer, pH 7.4, 0.5 mM ascorbate, 0.3% H_2O_2 and 100 μL of enzyme extract was formed at a total volume of 1 mL. Enzyme unit is stated as the quantity of APX necessary for the oxidation of 1 μM of ascorbate/min.

2.5.3 Catalase Assay (CAT)

The catalase activity (CAT) was calculated as the decline in supernatant absorbance at 240 nm as described by Aebi [25]. The reaction mixture of 1 mL comprised 50 mM sodium phosphate buffer (pH 7.0), 20 mM H_2O_2 and 100 μL of enzyme extract. Enzyme kinetics was performed for 15 min and expressed EU mg^{-1} protein min^{-1} . Enzyme unit is stated as the quantity of CAT necessary for the decomposition of 1 μM of H_2O_2 /min.

2.5.4 Glutathione Reductase Activity (GR)

The activity of glutathione reductase (GR) was performed by the method of Anderson et al. [26]. The reduction in absorbance at 340 nm was measured to calculate the enzyme activity. Enzyme kinetics was performed for 15 min and enzyme unit is stated as the quantity of GR which catalyzed the reduction of 1 μM of GSSG mg^{-1} protein min^{-1} .

2.6 Estimation of Non-Enzymatic Antioxidants

2.6.1 Proline Estimation

The amount of proline in leaf samples was estimated by the method described by Bates et al. [27]. 0.5 g of fresh leaves were ground and mixed with 10 mL of sulphosalicylic acid and then centrifuged for 10 min at 5590 xg. 2 mL acid ninhydrin mixed with 2 mL of supernatant with glacial acetic acid (2 mL) was added in it and placed in a water bath for 30 min at boiling temperature. After that, all reactions were done in an ice bath and toluene (4 mL) was added. The upper toluene layer was taken out into the cuvette and absorbance was recorded at 520 nm. Proline concentration was estimated by the standard curve of L-proline.

2.7 Statistical Analysis

All data were represented as the mean \pm standard error. Plants' triplicate data were measured for all physiological and biochemical parameters. Statistical analysis was done using one-way analysis of variance (ANOVA) in SPSS-17 statistical software (SPSS Inc., USA) and evaluation of significant differences was calculated for all the data at $p \leq 0.05$. MetaboAnalyst 3.0 (Appendix A) software was used for the generation of heat-map and calculation of Principal component analysis (PCA) and partial least squares discriminant analysis (PLS-DA).

2.8 RNA Isolation

Fresh leaf (1 g) was ground to powder by adding liquid N_2 and 1 mL TRIzol reagent. Centrifugation of lysate was done at 4°C at 12,000 xg for 5 min, supernatant was taken out into a fresh vial and incubated for a further 5 min to dissociate from nucleoproteins. Then 0.2 mL of chloroform was added to the vial and incubated for 2–3 min. Again, centrifuged at 12,000 xg for 15 min at 4°C. The upper RNA layer was

added to a new Eppendorf and 0.5 mL of Isopropanol was added and incubated for 10 min. Again, centrifugation was done at 12,000 xg for 10 min at 4°C. After discarding the supernatant, the pellet was resuspended in 1 mL of 75% ethanol. The sample was vortexed and then centrifuged at 7500 xg for 5 min. The RNA pellet was dried in air for 5 min, then dissolved in 40 µL of DEPC-treated water and 0.1 mM EDTA and further incubated at 60°C for 15 min. 5 µL of RNA solution was added to 495 µL DEPC-treated water.

2.8.1 Quantification of Total RNA

The absorbance of RNA samples was recorded at 260 and 280 nm using a spectrophotometer (IMPLEN, CA, USA). The ratio of OD₂₆₀/OD₂₈₀ and OD₂₆₀/OD₂₃₀ nm measures the purity of nucleic acids. A pure sample of RNA has an OD₂₆₀/OD₂₈₀ ratio lies in 1.8–2.0.

2.8.2 RNA Integrity Number (RIN)

RIN method is applied for electrophoretic RNA integrity measurements, usually obtained using gel electrophoresis with ethidium bromide staining for the visualization of bands of rRNA peaks. RIN for a sample is computed using several characteristics of an RNA electropherogram trace using the Agilent Bioanalyzer machine.

2.8.3 Qualitative Analysis of Total RNA

Isolated RNA from fresh leaves of Control, Cd and Hg treated plants was qualitatively analyzed in agarose (1%) gel electrophoresis stained with EtBr and seen with the help of a gel doc.

2.9 Reverse Transcriptase Polymerase Chain Reaction (RT-PCR) or cDNA Synthesis

cDNA first strand was synthesized using 2.5 µg of total RNA mixed with other reactants like oligo (dT) primer (18 mer, 0.5 µg in 1 µL) and DEPC-treated water (0.1%, 10 µL) were gently mixed, spun and incubated at 65°C/5 min and followed by placing on ice for 1 min. A mixture was prepared to have 4 µL reaction buffer, Ribolock RNase inhibitor (20 units in 0.5 µL), dNTPs mix (2 µL of 10 mM) and M-MuLV reverse transcriptase (200 units in 1 µL) mixed gently and briefly spun. Then this mixture was incubated at 42°C for 60 min, and after that reaction was terminated by heating at 70°C/10 min.

2.9.1 Primer Designing

Primers were designed and synthesized for housekeeping gene plant actin using Primer3 plus software as follows:

Plant-Actin-qFP: 5' – CTCAGTGGTGGTACTACTATGTTCACTGG – 3'

Plant-Actin-qRP: 5' – AGATGGAGCCTCCAATCCAGACACTG – 3'

2.10 RT-PCR Amplification

RT-PCR was done with primers of the actin gene. PCR was performed with initial PCR activation and denaturation done for 5 min at 94°C, followed by 35 cycles of denaturation at 94°C for 1 min, annealing at 52°C for 1 min and extension at 72°C for 1 min. The final extension was performed at 72°C for 10 min. Finally, PCR products picked with actin gene-specific primers were run and analyzed on 2% (w/v) agarose gel and photographed.

2.11 Quantitative Real-Time PCR (qRT-PCR)

qRT-PCR was done using a master mix of SYBR green, actin-specific primer and cDNA to make a volume of 50 µL. All reactions were kept in triplicates & run was taken on ABI Step-One Real-Time PCR machine. The conditions used for amplification were 95°C for 10 min, 40 cycles at 95°C for 15 s, and 60°C for 60 s followed by the conditions needed to calculate the melting curve. The cycle threshold

(Ct) values of all samples were exported from the detection software MxPro™ v.4.1 (Stratagene, Waldbronn, Germany).

3 Results and Discussion

3.1 Physiological Parameters

3.1.1 Metal Accumulation and Growth Attributes

The concentration of metals recorded in soybean root and leaf was shown in (Table 1). Both 100 μM Cd and 100 μM Hg levels recorded the highest accumulation in roots as compared to leaves. Growth parameters were calculated on plant triplicates. It was noted both Cd and Hg caused stunted growth, chlorosis and necrosis in soybean leaves (Fig. 1). 100 μM concentration of CdCl₂ and HgCl₂ significantly decreased growth attributes (Table 2).

Table 1: Metal accumulation in the root and leaf of soybean plant at 40 DAS expressed as $\mu\text{g/g}$ dry weight (DW)

Treatments	Metal concentrations ($\mu\text{g/g}$) in root	Metal concentrations ($\mu\text{g/g}$) in leaf
Control (00)	Not detected	Not detected
Cd (100 μM)	290.24 \pm 9.03 ^a	91.18 \pm 3.12 ^b
Hg (100 μM)	304.46 \pm 10.36 ^a	101.32 \pm 5.71 ^b

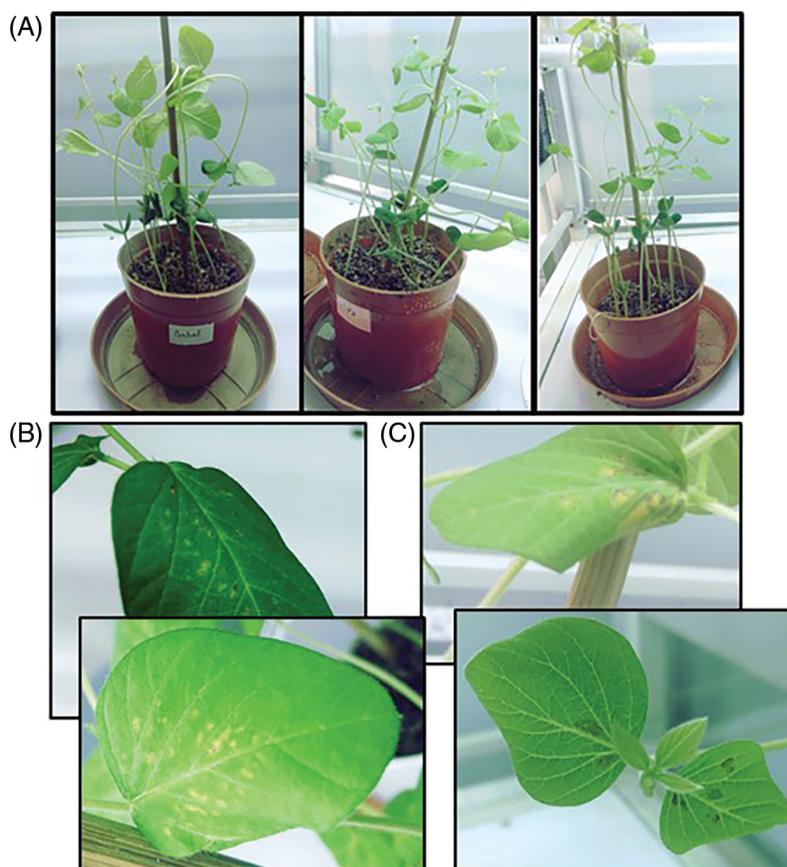


Figure 1: View of mature experimental soybean plants (A) All three sets (Control, Cd, Hg) (B) impact of Cd (100 μM) (C) impact of Hg (100 μM)

Table 2: Effect of Cd and Hg on plant height, fresh weight (FW) and dry weight (DW) of root, stem and leaf in soybean at 40 DAS

Plant	Control	Cd treated	Hg treated
Plant height (cm)	41 ± 1.47 ^a	32 ± 1.15 ^c (-21.9%)	35 ± 1.26 ^b (-14.1%)
Root fresh weight (g)	6.18 ± 0.19 ^a	3.96 ± 0.10 ^c (-36.0%)	4.98 ± 0.29 ^b (-19.4%)
Root dry weight (g)	1.96 ± 0.02 ^a	1.38 ± 0.01 ^c (-29.5%)	1.51 ± 0.02 ^b (-22.1%)
Shoot fresh weight (g)	8.20 ± 0.38 ^a	5.85 ± 0.15 ^c (-28.6%)	6.12 ± 0.18 ^b (-25.3%)
Shoot dry weight (g)	1.35 ± 0.03 ^a	0.85 ± 0.03 ^c (-37.0%)	1.00 ± 0.03 ^b (-25.9%)
Leaf fresh weight (g)	12.66 ± 0.75 ^a	9.06 ± 0.40 ^c (-28.4%)	10.55 ± 0.55 ^b (-16.6%)
Leaf dry weight (g)	1.60 ± 0.05 ^a	0.98 ± 0.03 ^c (-38.7%)	1.24 ± 0.02 ^b (-22.5%)

Each value indicates the mean of three replicates along with ± SE (Standard Error). Different lowercase letters show the presence of significant differences among treatments ($p \leq 0.05$).

Treatment of Cd and Hg significantly decreased growth attributes as shown by earlier studies [28]. In response to Cd and Hg, a drastic reduction of growth parameters included plant height, shoot and root fresh weights (FW) and shoot and root dry weights (DW), leaf dry and fresh weight was observed in soybean plants as compared to Control. Reduction in plant growth might be due to Cd and Hg-induced changes in the plant biochemicals or physiological processes comprising cell division, cell differentiation and morphogenesis [29]. In our findings, Cd caused more damage to the plant physiologically as compared to Hg stress, with used concentrations.

3.2 Biochemical Parameters

3.2.1 Effect on Photosynthetic Pigments

Cd and Hg treated plants revealed a considerably diminished in photosynthetic pigments chlorophyll a (15.1% and 9.6%), respectively, chlorophyll b (28.2% and 16.3%), total chlorophyll (33.8% and 27.1%) and total carotenoids (25% and 16.6%) respectively of soybean leaves as compared with control plants shown in Fig. 2.

Chl a, and chl b are the main photosynthetic pigments that are associated with photosynthetic competence [30]. Contents of chl a, chl b, total chlorophyll, and total carotenoids declined due to Cd and Hg treatments. This decrease could be due to the inhibited synthesis or degradation by oxidative stress [31]. Carotenoids are essential substances that acted as photoreceptor pigments and as a regulator for abolishing singlet oxygen. Several studies have suggested a reduction in these photosynthetic pigments could be owing to the substitution of metal ions from molecules (such as Mg from chlorophyll), denaturing or inactivation of proteins, and disruption of cell membranes or organelles [31–33].

3.2.2 Cd and Hg Induced Changes in Antioxidant Enzymes

The activities of antioxidant enzymes such as superoxide dismutase (SOD), ascorbate peroxidase (APX), glutathione reductase, and catalase (CAT) were altered in response to both Cd and Hg stress. It is noteworthy that Cd and Hg-treated plants showed an increase in SOD, APX, GR, and CAT activities. SOD in the control plant was found to be 32 EU, 48.33 EU (+51%) in Cd and 42.39 EU (+24.5%) in the case of Hg (Fig. 3A). APX was found to be around 1.48 EU in control, 1.95 EU (+31.75%) in Cd and 1.82 EU (+23%) in case of Hg (Fig. 3B). GR was found to be around 0.626 EU in control, 1.12 EU in Cd (+78.9%) and 0.968 EU (+54.6%) in Hg (Fig. 3C). CAT activity was 1.36 EU in control, 1.96 EU (+44%) in Cd, and 1.83 EU (+34%) in Hg treatment (Fig. 3D).

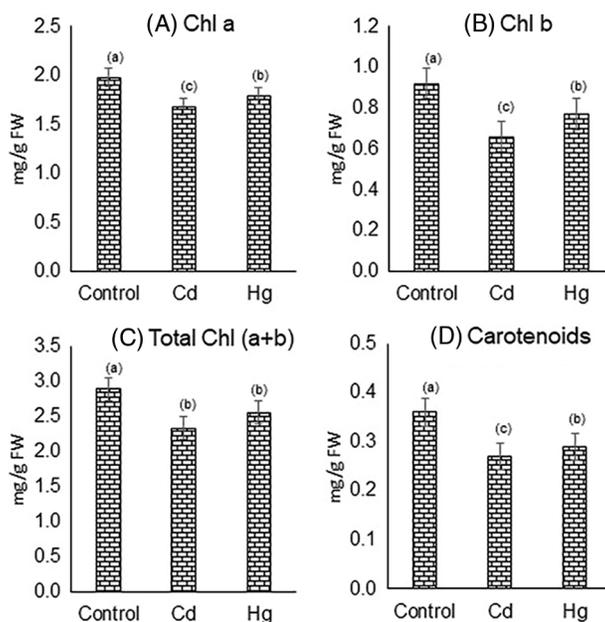


Figure 2: Impact of Cd and Hg on contents of photosynthetic pigments: (A) chlorophyll a, (B) chlorophyll b, (C) total chlorophyll (a + b), and (D) carotenoids. Each value indicates the mean of three replicates along with \pm SE (Standard Error). Different letters show the presence of significant differences among treatments ($p \leq 0.05$)

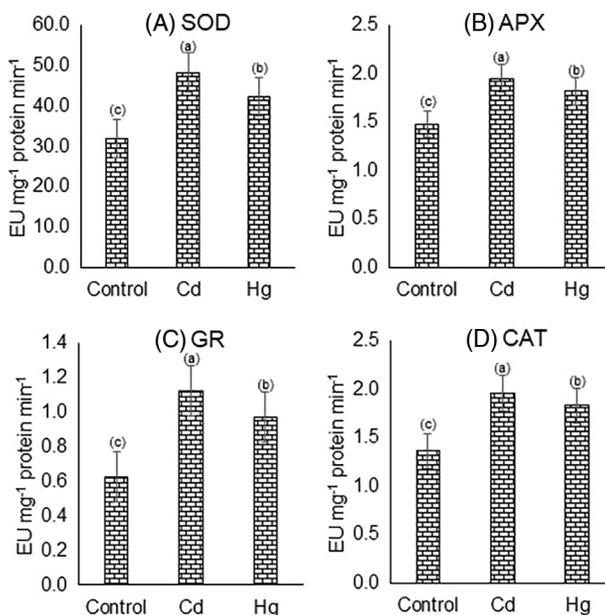


Figure 3: Impact of Cd and Hg on activities of enzymatic antioxidants: (A) Superoxide dismutase (SOD), (B) Ascorbate peroxidase (APX), (C) Glutathione reductase (GR), and (D) Catalase (CAT) activity. Each value indicates the mean of three replicates along with \pm SE (Standard Error). Different letters show the presence of significant differences among treatments ($p \leq 0.05$)

Plants retain a considerable volume of antioxidative enzymes for preventing oxidative damage. It is revealed that Cd and Hg both heavy metals treatments increased oxidative stress levels in plants due to which enzymatic antioxidative activities were enhanced greatly. This study correlated with previous studies in which antioxidative enzyme activities increased under heavy metal stress conditions [34–36]. Cd induces significant changes while Hg did not make much difference as compared to the untreated plant this may be due to the differential nature of both metals in terms of physiochemical properties. Increase in CAT, GR, APX and SOD activities were found in response to Cd and Hg stress. In plants' natural defence system, Superoxide dismutase protects plants against ROS which was produced in stress conditions [6,33]. Conversion of toxic superoxide radical (O_2^-) to less toxic H_2O_2 and O_2 was catalyzed by superoxide dismutase after induction of Cd and Hg stress. SOD perhaps removed more and more O_2^- and reduced the chances of OH^- formation from superoxide radicals [34]. Higher SOD activity might be due to more increase in the content of O_2^- ions or higher formation of H_2O_2 . CAT transforms H_2O_2 to H_2O and O_2 for preventing oxidative damage [33]. APX is a vital H_2O_2 scavenger that works in the Ascorbate Glutathione cycle as an electron donor. APX and GR activities significantly increased in the presence of Cd and Hg-treatment which was also proved by earlier studies [37,38]. Cd gave rise to oxidative damage to trigger anti-oxidative enzyme activities in plants. GR works with MDHAR and DHAR to sustain cellular redox balance [39]. Concerning this, few reports previously showed increased antioxidant resistance capacity in plants by the stimulation of some antioxidant enzyme activities such as GR in maize and other plants also [38,39].

3.2.3 Impact on Oxidative Stress (TBARS, Proline, Hydrogen Peroxide)

Heavy metal Cd and Hg increased the oxidative stress level. Control, Cd and Hg treated plants accumulated 17.38, 34.68 (78.9%), and 25.49 (46.6%) nmol TBARS g^{-1} FW, respectively in Fig. 4A.

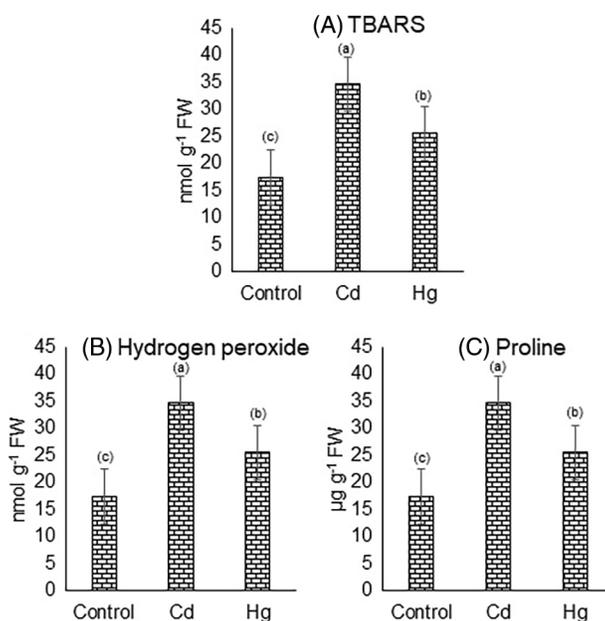


Figure 4: Impact of Cd and Hg on contents of (A) TBARS, (B) Hydrogen peroxide (H_2O_2), and (C) Proline. Each value indicates the mean of three replicates along with \pm SE (Standard Error). Different lowercase letters show the presence of significant differences among treatments ($p \leq 0.05$)

In response to Cd and Hg stress the hydrogen peroxide (H_2O_2) content in soybean leaves was considerably increased. The H_2O_2 level in Control was found to be 34.16 ($\mu mol/g$ DW) which is

increased by 59.80 (75%) and 49.79 (45.7%) in the case of Cd and Hg treated plants respectively in comparison to that of control in (Fig. 4B).

The effect of Cd and Hg stress increased leaf proline content in soybean. Proline content in Control was found to be around 68.05 ($\mu\text{g/g}$ FW) while in the case of Cd and Hg it increased to 102.08 $\mu\text{g/g}$ FW (50%), and 96.70 $\mu\text{g/g}$ FW (42.1%), respectively (Fig. 4C).

In histochemical detection of hydrogen peroxide, both Cd and Hg increased H_2O_2 accumulation which was seen in (Fig. 5).

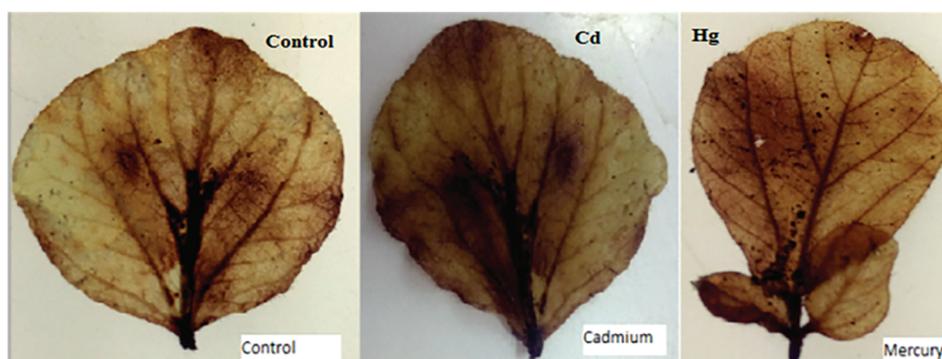


Figure 5: Soybean leaves showing H_2O_2 accumulation under Control, Cd and Hg treatment

TBARS and H_2O_2 contents were estimated in leaves which are considered an important indicator of oxidative stress [6,33,37]. Our results revealed that TBARS and H_2O_2 contents were increased after treatment of Cd and Hg in the soybean plant. Cd generated more oxidative stress as compared to Hg. It has been proposed that H_2O_2 contents escalated in several plants because of metal-induced stress [35]. Under Cd stress conditions, ROS levels and TBARS content increased. Increased ROS damages lipids membranes; besides, causing oxidative damage to other biomolecules in soybean plants [36]. A more abundant amount of proline was formed in stressed conditions. Proline rise in plants is a stress tolerance indicator and serve as osmoprotectant. Presumably, the metal-stressed plants activated the enzymes of proline biosynthesis, consequently producing more proline in leaves [37]. Cd shows more increase as compared to Hg.

3.3 Analysis of Multivariate Data

The connection of metabolites of oxidative stress, antioxidants, pigments, ascorbate peroxidase, glutathione reductase, and proline under Cd and Hg stress was analyzed by multivariate data using MetaboAnalyst software (Figs. 6A–6D). A heatmap of hierarchical clustering was generated for studying the correlation between these metabolites under Cd and Hg treatments concerning control. Hierarchical clustering showed variation between differentially expressed metabolites in the case of control and heavy metal treated plants (Fig. 6A). Principal component analysis loadings and score plots give information on our data and compare the differentially expressed metabolites in a soybean plant (Fig. 6C). Identification of maximum changes in the abundance of metabolites was described by the VIP plot which was formed from the loadings of PLS-DA (Fig. 6B). In the VIP plot 11 components came with VIP scores (0.10–2.0) including components such as proline (pro), H_2O_2 , CAT, TBARS, total chlorophyll (tchl), APX, total carotenoids (tcaro), SOD, chlorophyll a (chl a), GR and chlorophyll b (chl b) in soybean. Graphs showed H_2O_2 content, proline content and level of antioxidants increased (Fig. 6B) (highlighted in red), but chlorophyll and carotenoids content decreased in (Fig. 6B) (seen in blue) in both Cd and Hg stress which suggested that antioxidants, proline, and H_2O_2 helps in plant defence against heavy metal stress.

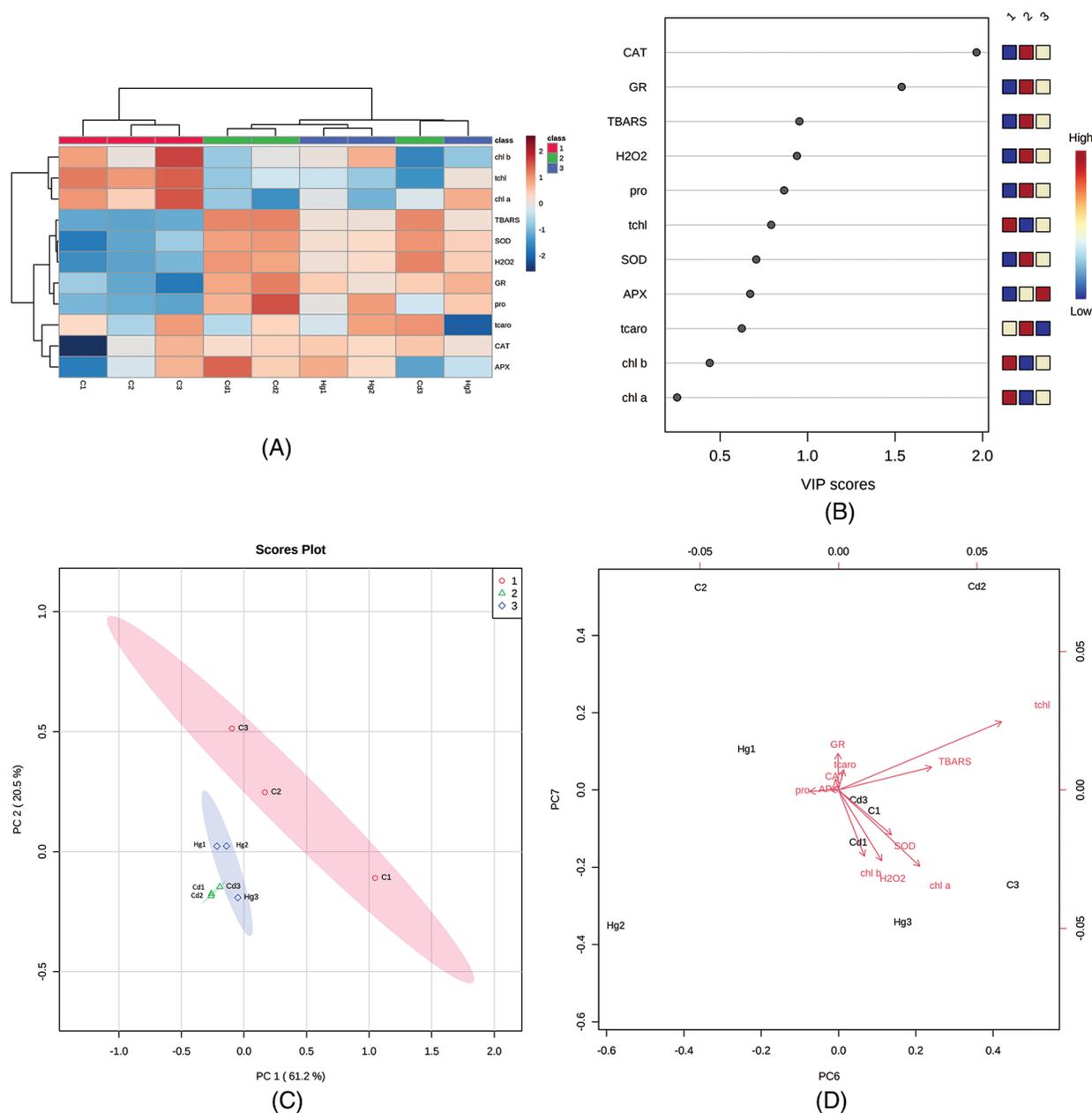


Figure 6: Multivariate data analysis obtained through MetaboAnalyst software (A) Hierarchical clustering (B) VIP plot (C) Scatter plot (D) Loadings plot showing differentially expressed metabolites (Biplot). These plots of PCA analysis indicate a correlation between different metabolic, antioxidant and oxidative stress parameters

The first axis of the loadings score plot has 61.2% of the variance for the total values under Cd and Hg stress in leaves of soybean. TBARS, H₂O₂, and proline contributed to the first axis but on the second axis PCA2 contributed by antioxidants, chlorophyll and carotenoids showed less importance (20.5% of variance). The direction and length of the vectors point toward the potency and show a correlation among them. Large extended vectors of these studied parameters indicated that the vector significantly affects the outcome of our results. From these results, it was clear that the chl a, chl b and total carotenoids grouped

and shows a positive correlation as found on positive loading of the right side of the biplot. Proline, CAT and H₂O₂ show a positive correlation in the biplot (Fig. 6D). The angles found among these different antioxidant parameters are small, demonstrating that they are in positive correlation [29]. However total chlorophyll and carotenoids are segregated in opposite directions with antioxidants, indicating that these factors are negatively correlated (Fig. 6D).

3.4 Molecular Aspects in Response to Cd and Hg Stress

3.4.1 Impact on Isolated RNA and Its Quantification

Total isolated RNA was found to be around 273 µg/g FW in the case of the control leaf. In response to Cd and Hg stress, the concentration of RNA was decreased. Anything higher than 1.8 is considered to be of acceptable purity. A low A₂₆₀/A₂₈₀ ratio in the case of Cd and Hg-treated leaf was likely due to the low yield of RNA (Table 3).

Table 3: Quantification of total isolated RNA from control and treated (Cd and Hg) soybean plants

Plants	Concentration of RNA (µg/g FW)	A ₂₆₀ /A ₂₈₀	A ₂₆₀ /A ₂₃₀	RIN
Control	273 ± 23	2.04 ± 0.016	2.10 ± 0.011	8.6
Cadmium	211 ± 18	1.97 ± 0.006	2.04 ± 0.012	5.3
Mercury	226 ± 15	1.99 ± 0.014	2.06 ± 0.002	6.2

Note: n = 3, $p \leq 0.05$ FW.

Cd seemed to have a major negative effect on the plant at the molecular level but Hg in comparison to Cd seems to have less destructive (Fig. 7) in the used concentration. The effect of Cd and Hg on isolated RNA was revealed. When compared with Control plant RNA, Cd and Hg-affected plants showed low RNA quality, and quantity as well as changed RNA integrity, as seen in the results section and also observed in earlier studies [40,41]. We considered that these responses were aroused through free radical-mediated oxidation, as hypothesized for other cellular constituents [42–45]. Cd causes oxidative stress and leads to the formation of ROS. In the presence of Cd, a major surge in oxidative stress might have led to the degradation of RNA. It has been shown that the 2'OH group, present in RNA, hydrolyzes readily which makes RNA an easily labile molecule [13,41]. Another reason that RNA was prone to degradation by reactive oxygen species (ROS) is that RNA is single-stranded, making it more vulnerable to degradation [44–47].

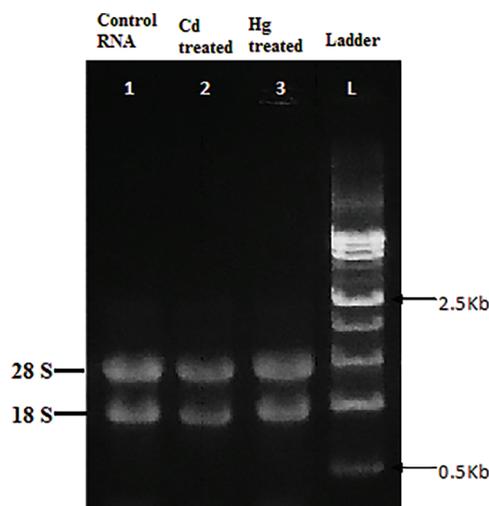


Figure 7: Total RNA isolated from soybean leaves under Control, Cd and Hg stress loaded in 1% Agarose gel with ladder

Destabilization of RNA could also be attributed to the interaction and intercalation of defence-related secondary metabolites with RNA which reduced its yield and quality [47]. RNA was quantitatively and qualitatively examined through spectrophotometry and optimum values of control samples suggested that non-stressed plants' RNA remained stable but compromised in Cd- and Hg-treated plants [13,42,46].

In comparison to Cd, plants treated with Hg had better RNA in terms of quantity and quality. The mechanism which might have led to nucleic acid degradation is increased level of oxyradicals and degradation through a decay pathway called No-Go [43].

3.4.2 Effect on RNA Integrity Number (RIN)

RNA of control plants exhibited the highest RNA integrity value which implied no degradation at all but in the case of Cd, RNA integrity was lesser indicating towards degradation of RNA molecules (Fig. 7) and (Table 3). RNA integrity values (RIN) also may also be decreased due to the degradation of labile RNA molecules [40]. As RNA integrity check is the critical step toward gene expression, the degraded RNA has a direct impact on the transcript levels [44,47].

3.5 Gene Expression Analysis through qRT-PCR

In RT-PCR, cDNA was synthesized using actin gene-specific primers and PCR amplification was done which produced a 142 bp amplified product.

In qRT-PCR, Mean Ct values were calculated from triplicates data of Control, Cd and Hg-treated soybean. The mean Ct of actin gene of the control plant was found to be 23.5150 while in the case of Cd stress it was 19.0940. The mean Ct of the actin gene in response to Hg stress was found to be 21.5760. All Ct values obtained for each sample individually were represented on a box plot diagram (Fig. 8) in which the median Ct value was represented by middle lines.

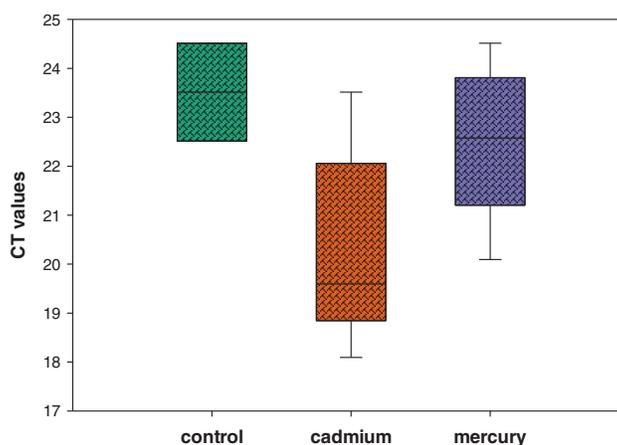


Figure 8: Box plot analysis of Ct values of actin gene obtained from distinct sample sets of Control, Cd and Hg treatment

Mean Ct was used to obtain the expression of actin gene in control and treated plants. Quantitative real-time expression analysis was performed using qRT-PCR. Data showed variations in Ct values of the actin gene in response to Cd and Hg stress [48] as compared to the control. In terms of gene expression, Ct values were negatively related to gene expression level, so the lower the Ct the higher the gene expressed [49,50]. Actin gene showed significant upregulation in response to Cd stress which might be due to the presence of metal-responsive signals produced inside the cell which ultimately leads to transcriptional

factors activation and initiation of transcription to undo the damage caused by heavy metals [51,52]. Cd and Hg might have affected actin filaments through ROS which has activated repair mechanisms that increased actin gene expression directly. Stability and expression of actin gene which was determined by mean Ct values revealed upregulation in expression level (lowest Ct value) in response to Cd stress as compared to Control which had the highest Ct value and showed stable expression [53–57].

Ideally, the expression of this internal control gene was not changed. However abiotic stress factors can alter gene expression in plants [56,58–60]. Our study was also validated by previous studies which suggested that internal standards, mostly housekeeping genes used in the quantification of RNA expression in qRT-PCR could vary with many biotic and abiotic stress factors [58–60].

4 Conclusion

Our study contributed to comparative physiological, biochemical and some molecular aspects of Cd and Hg stress in the soybean plant at the vegetative stage (pre-flowering). Plants reacted differentially when exposed to Cd and Hg and generated high levels of oxidative stress. Our findings concluded that the soybean plant showed more response in case of Cd stress compared to Hg and proved to be more disastrous. Soybean plants responded to heavy metal stress by accumulating more proline and increased enzymatic and non-enzymatic cellular antioxidant activities for their defence. It can be concluded that both Cd and Hg caused oxidative damage to plants which adversely affected the quality of RNA. The actin gene showed a differential gene expression pattern in response to Cd and Hg, especially Cd upregulated the actin gene expression. qRT-PCR results contributed to the choice of selection of a more stable and better housekeeping gene to be used as a reference gene in qRT-PCR in plants exposed to heavy metal stress.

Acknowledgement: The authors would like to extend their sincere appreciation to the researchers supporting Project Number RSP2023R186, King Saud University, Riyadh, Saudi Arabia.

Funding Statement: The study has been supported by Project Number RSP2023R186, King Saud University, Riyadh, Saudi Arabia.

Author Contributions: Conceptualization, S. N. and M. I. Q. Methodology, S. N. and N. A. Software, S. N., and M. I. Validation, S. N., A. A. A. H. and M. I. Data curation, S. N., A. A. A. H and M. I. Q. Writing—original draft preparation, S. N., and F. K. Writing—review and editing, S. N., A. A. A. H., and M. I. Q. Supervision, N. A. and M. I. Q. All authors have read and agreed to the published version of the manuscript.

Conflicts of Interest: The authors declare that they have no conflicts of interest to report regarding the present study.

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Appendix AThe *p*-value correlation table

	tchl	tcaro	pro	TBARS	SOD	H ₂ O ₂	CAT	APX	GR
tchl		0.15817	0.098089	0.004413	0.02508	0.018474	0.37778	0.65195	0.86571
tcaro	0.15817		0.52573	0.44534	0.66828	0.5645	0.50895	0.6998	0.47898
pro	0.098089	0.52573		0.01878	0.015293	0.021586	0.57798	0.31949	0.32052
TBARS	0.004413	0.44534	0.01878		0.000113	0.00011	0.19853	0.39165	0.80934
SOD	0.02508	0.66828	0.015293	0.000113		4.03E-07	0.12271	0.34706	0.36109
H ₂ O ₂	0.018474	0.5645	0.021586	0.00011	4.03E-07		0.22346	0.45864	0.45231
CAT	0.37778	0.50895	0.57798	0.19853	0.12271	0.22346		0.16306	0.35073
APX	0.65195	0.6998	0.31949	0.39165	0.34706	0.45864	0.16306		0.099853
GR	0.86571	0.47898	0.32052	0.80934	0.36109	0.45231	0.35073	0.099853	